

BMJ Open

BMJ Open is committed to open peer review. As part of this commitment we make the peer review history of every article we publish publicly available.

When an article is published we post the peer reviewers' comments and the authors' responses online. We also post the versions of the paper that were used during peer review. These are the versions that the peer review comments apply to.

The versions of the paper that follow are the versions that were submitted during the peer review process. They are not the versions of record or the final published versions. They should not be cited or distributed as the published version of this manuscript.

BMJ Open is an open access journal and the full, final, typeset and author-corrected version of record of the manuscript is available on our site with no access controls, subscription charges or pay-per-view fees (<http://bmjopen.bmj.com>).

If you have any questions on BMJ Open's open peer review process please email editorial.bmjopen@bmj.com

BMJ Open

Oral cannabinoids in people living with HIV on effective antiretroviral therapy—CTN PT028: Study protocol for a pilot randomized trial to assess safety, tolerability and effect in on immune activation

Journal:	<i>BMJ Open</i>
Manuscript ID	bmjopen-2018-024793
Article Type:	Protocol
Date Submitted by the Author:	15-Jun-2018
Complete List of Authors:	<p>Costiniuk, C; Chronic Viral Illness Service, McGill University Health Centre; Research Institute, McGill University Health Centre</p> <p>Ware, Mark; Department of Family Medicine, McGill University Health Centre; Department of Anesthesia, McGill University Health Centre</p> <p>Saneei, Zahra; Chronic Viral Illness Service, McGill University Health Centre</p> <p>Routy, JP; Chronic Viral Illness Service, McGill University Health Centre; Research Institute of the McGill University Health Centre</p> <p>Margolese, Shari; Canadian Institutes of Health Research Canadian HIV Trials Network</p> <p>Mandarino, Enrico; Canadian Institutes of Health Research Canadian HIV Trials Network; WILLL Cannabis Group</p> <p>Singer, Joel; Canadian Institutes of Health Research Canadian HIV Trials Network; School of Population and Public Health, University of British Columbia, Vancouver, BC, Canada</p> <p>Lebouché, Bertrand; Chronic Viral Illness Service, McGill University Health Centre; Department of Family Medicine, McGill University</p> <p>Cox, Joseph; Chronic Viral Illness Service, McGill University Health Centre; Department of Family Medicine, McGill University</p> <p>Szabo, Jason; Chronic Viral Illness Service, McGill University Health Centre; Department of Family Medicine, McGill University</p> <p>Brouillette, Marie-Josée ; Chronic Viral Illness Service, McGill University Health Centre; Department of Psychiatry, McGill University Health Centre</p> <p>Klein, Marina; Chronic Viral Illness Service, McGill University Health Centre; Research Institute, McGill University Health Centre</p> <p>Chomont, Nicolas; Centre de Recherche du CHUM and Department of microbiology, infectiology and immunology, Université de Montréal</p> <p>Jenabian, Mohammad-Ali; Université du Québec à Montréal, Biological Sciences</p>
Keywords:	cannabinoids, cannabis, safety and tolerability, inflammation, immune activation, HIV reservoir



Oral cannabinoids in people living with HIV on effective antiretroviral therapy—CTN

PT028: Study protocol for a pilot randomized trial to assess safety, tolerability and effect on immune activation

Cecilia T. Costiniuk^{1,2}, Mark Ware^{2,4}, Zahra Saneei¹, Jean-Pierre Routy^{1,2}, Shari Margolese⁵, Enrico Mandarino⁶, Joel Singer⁷, Bertrand Lebouché^{1,3}, Joseph Cox^{1,3}, Jason Szabo^{1,3}, Marie-Josée Brouillette^{1,2,8}, Marina B. Klein^{1,2}, Nicolas Chomont⁹, Mohammad-Ali Jenabian¹⁰

¹Chronic Viral Illness Service, McGill University Health Centre, Montreal, QC, Canada

²Research Institute of McGill University Health Centre, Montreal, QC, Canada

³Department of Family Medicine, McGill University Health Centre, Montreal, QC, Canada

⁴Department of Anesthesia, McGill University Health Centre, Montreal, QC, Canada

⁵CIHR Canadian HIV Trials Network

⁶CIHR Canadian HIV Trials Network; WILL Cannabis Group

⁷Canadian Institute of Health Research Canadian HIV Trials Network, Vancouver, BC, Canada;

School of Population and Public Health, University of British Columbia, Vancouver, BC,

Canada

⁸Department of Psychiatry, McGill University Health Centre, Montreal, QC, Canada

⁹Centre de Recherche du CHUM and Department of microbiology, infectiology and immunology, Université de Montréal, Montreal, QC, Canada

¹⁰Department of Biological Sciences and BioMed Research Centre, University of Quebec at Montreal (UQAM), Montreal, Quebec, Canada

Key words: cannabinoids; cannabis; HIV; inflammation; immune activation; HIV reservoir, microbiome

Short title: Cannabinoids and HIV

Word count: 6,936

Correspondence: Cecilia Costiniuk, MD, MSc, Division of Infectious Diseases and Chronic Viral Illness Service, McGill University Health Centre, 1001 boulevard Décarie, Montreal (QC) Canada, H4A 3J1 Tel: 1 (514) 843-2090; Fax: 1 (514) 843 2092; Email: cecilia.costiniuk@mcgill.ca

Abstract:

Introduction: Despite antiretroviral therapy(ART), people living with HIV have higher rates of non-infectious chronic diseases. These conditions are driven by relatively high levels of inflammation persisting on ART compared to uninfected individuals. Chronic inflammation also contributes to HIV persistence during ART. Cannabis when taken orally may represent a way to reduce inflammation and strengthen immune responses. Before planning large interventional studies, it is important to ensure that cannabis taken orally is safe and well-tolerated in people living with HIV. We propose to conduct a pilot randomized trial to examine the safety and tolerability of cannabis oils containing tetrahydrocannabinol(THC) and cannabidiol(CBD) consumed orally in people living with HIV. We will also measure inflammatory markers, markers of HIV persistence in peripheral blood cells and changes in the gastrointestinal microbiome.

Methods and Analysis: Twenty-six people living with HIV having undetectable viral load for at least three years will be randomized to receive TN-TC11LM(THC:CBD in 1:1 ratio) or TN-TC19LM(THC:CBD in 1:9 ratio) capsules daily for 12 weeks. Safety and tolerability of these capsules will be assessed through haematological, hepatic and renal blood tests, face-to-face interviews and questionnaires. Proportions of participants without any signs of significant

toxicity(Grades 0-2 scores on the WHO toxicity scale) and who complete the study, as well as scores on quality of life and mood will be examined using descriptive statistics. The effects on inflammatory markers, markers of peripheral blood reservoir size and effect on the composition of the gastrointestinal microbiome will be assessed before and after study completion.

Ethics and Dissemination: This study has been approved by the Research Institute of the McGill University Health Centre. A Data Safety Monitor will review safety information at regular intervals. The final manuscript will be submitted to an open-access journal within 6 months of study completion. This trial is registered with clinicaltrials.gov([NCT03550352](https://clinicaltrials.gov/ct2/show/study/NCT03550352)).

Strengths and limitations of this study

- Will provide important preliminary data of cannabinoids on safety and tolerability in people living with HIV
- The first and only randomized clinical trial to date which will examine the effects of oral cannabinoids on markers of inflammation and HIV reservoir, as well as the gut microbiome
- Identified by Community Advisory Committee for people living with HIV as a key interest of study and study commencement coincides with cannabis legalization in Canada
- The number of participants should provide insight into the degree of variability for continuous outcomes we are measuring and should help guide future sample size calculations for larger studies

1

2

3

4 **Background**

5

6

7

8 Despite antiretroviral therapy (ART), persistent immune activation is associated with

9

10 increased risk of non-opportunistic complications in people living with HIV such as

11

12 cardiovascular, pulmonary, renal and hepatic events^{1 2}. HIV pathogenesis and persistence appear

13

14 to be related to chronic inflammation and immune activation³, driven by microbial translocation

15

16 of bacterial products across the gut mucosa⁴⁻⁶. Even when ART is initiated in the primary or

17

18 early phase of HIV, gut integrity is not fully restored⁷. Furthermore, there is recent evidence to

19

20 suggest that inflammatory features of the enteric microbiota, and not just increased permeability

21

22 alone, is driving chronic inflammation in people living with HIV. Indeed, some studies have

23

24 shown correlations between specific enteric bacteria and immune activation markers in gut and

25

26 blood^{8 9}. Persistent immune activation may also contribute to the persistence of HIV during

27

28 ART¹⁰⁻¹⁵. HIV reservoirs are the reason why HIV remains an incurable infection. Although

29

30 HIV may also persist in myeloid cells, CD4+ T cells are the best-characterized and the most

31

32 abundant reservoirs¹⁶⁻¹⁸.

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47 Attenuation of immune activation and levels of inflammation may portend several

48

49 therapeutic benefits to people living with HIV and novel strategies are needed to achieve this

50

51 goal. Having both anti-inflammatory and anti-fibrotic properties¹⁹, cannabis may represent a

52

53

54

55

56

57

58

59

60

feasible method to reduce immune activation and enhance immune profile. When consumed orally, notably soluble in oil, it may also temper inflammation at the gastrointestinal mucosa²⁰. This, in turn, may hasten the progression of non-opportunistic complications associated with HIV. Furthermore, cannabis use, mainly through smoking, is already extremely common amongst PLWHIV. A study conducted by our team involving 1072 participants in the Canadian Co-infection Cohort Study (CTN 222), a multi-centered longitudinal study of HIV-Hepatitis C co-infected individuals, revealed that 53% of individuals had smoked marijuana in the past 6 months²¹. A clearer understanding of the safety, tolerability and feasibility of using cannabis in a clinical trial is an important first step before designing interventional studies to ameliorate specific conditions of importance to people living with HIV. Furthermore, as cannabis use will be legalized in Canada July 2018, understanding its safety profile is very important.

Pharmacology and medical properties of cannabis

Cannabinoids, found in the hemp plant *Cannabis sativa*, have been recognized for centuries for their analgesic, anticonvulsant, bronchodilatory, sedative, hypnotic and antispasmodic properties^{22 23}. Their biological activity is conferred by cannabinoid receptors CB1 and CB2 through activation of heterodimeric G-proteins that function as signaling and

regulatory proteins to operate or modulate intracellular signaling pathways^{24 25}. While CB1 receptors are expressed predominantly in the central nervous system, they are also found in the lung, liver and kidneys. The endocannabinoid system also plays a key role in the gastrointestinal tract's neural and molecular control mechanisms. Indeed, the endocannabinoid system plays a role in normal physiological functions of the gastrointestinal tract including motility, gut-brain-mediated fat intake, hunger signaling, inflammation and gut permeability²⁶. Furthermore, there is some evidence for and interactions between the endocannabinoid system and the gut microbiota²⁶. In lean mice who received 4 weeks of a cannabinoid-receptor agonist HU-210, plasma lipopolysaccharides (LPS) were significantly increased^{26 27}. When obese mice with disrupted gastrointestinal musoca were treated with rimonabant, an inverse CB1 agonist, for nearly 2 weeks, reductions in plasma LPS were observed^{26 28}. Furthermore, improvements in localization of tight junction proteins, occluding and zonula occludens-1 were measured, suggestive of improvement in endothelial barrier function^{26 28}.

The presence of cannabinoid receptors in the central nervous system accounts for the psychoactive effects of cannabis^{24 25}. In contrast, CB2 receptors are abundant on immune cells including T and B cells, natural killer cells, monocytes and neutrophils as well as the liver^{24 29}. Two primary active constituents found in hemp plants include Δ9-tetrahydrocannabinol (THC) and cannabidiol (CBD). Δ9-THC is a partial agonist at both CB1 and CB2^{3 30 31} while CBD is

thought to mediate its effects through a variety of signaling mechanisms and act as a negative allosteric modulator at CB1^{32 33}.

Cannabis is perhaps best known for its effect on stimulating appetite via CB1 receptor activity and was historically used for this purpose in AIDS wasting syndrome. A variety of randomized controlled trials have shown that smoked cannabis is efficacious in the management of chronic pain, including in painful HIV-associated sensory neuropathy³⁴⁻³⁶. In animal models, Δ^9 -THC prevents atherosclerosis by inhibiting macrophage migration into atheromas through CB2 activation^{37 38}. Endocannabinoids may also play a role in liver disease, where CB1 receptors are present in endothelial cells and hepatocytes and CB2 receptors are distributed in Kupffer cells^{39 40}. CB1 receptors have an important role in non-alcoholic fatty liver disease and alcoholic liver disease while anti-inflammatory action of CB2 receptors can be useful in inflammatory liver disease. CB2 receptors show antifibrinogenic properties and administration of its agonists in fibrotic rats resulted in improvement in liver fibrosis, decreased inflammation and increased apoptosis of hepatic myofibroblasts⁴⁰⁻⁴⁴.

Cannabis, the immune system and HIV

1

2

3

4 ***In vitro* studies**

5

6

7 Cannabinoids have been shown to inhibit productive HIV infection in primary human T

8 cells and a CB2 antagonist blocked this effect⁴⁵. Interference with the signal transduction of

9

10 chemokine receptor CXCR4 is thought to lead to reduced F-actin accumulation. This in turn

11

12 prevents movement of viral pre-integration complexes to the nucleus. It has also been postulated

13

14 that CB2 agonists may inhibit T cell activation induced by anti-CD3/anti-CD28⁴⁵. Along with

15

16 reduced HIV production, immunological effects of cannabinoids include induction of

17

18 immunosuppressive cytokines including IL-10, TGF- β and inhibition of IL-2 which stimulates

19

20 T-cell division and expansion. They have also been shown to decrease adhesion capacity of

21

22 leukocytes to extravasate into sites of inflammation⁴⁶⁻⁴⁸. Apoptosis and induction of Treg may

23

24 potentially add to the pathways of cannabinoid-mediated immunosuppression⁴⁹.

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45 **Animal studies**

46

47

48 Mice treated with Δ 9-THC experience persistent inhibition of IL-2 production even 7

49

50 days after treatment⁵⁰. Rhesus macaques who were administered Δ 9-THC for 28 days prior to

51

52 Simian Immunodeficiency virus (SIV) inoculation had reduced mortality and reduced SIV viral

53

54 load in the cerebrospinal fluid and plasma. In another study involving macaques infected with

55

56

57

58

59

60

SIV for 17 months of chronic THC administration⁵¹, Δ 9-THC resulted in trends towards a decreased viral load. The lack of statistical significance in some parameters can be attributed to the small number of animals in the vehicle vs. the Δ 9-THC groups (n=4)³⁰.

Human studies

To date, seven randomized controlled studies have examined the medicinal use of cannabis for reducing morbidity and mortality in people living with HIV⁵². Interventions included smoked marijuana or hashish, ingested marijuana or hashish or ingested THC (dronabinol or other synthetic cannabinoid). Studies ranged from 21-84 days. Primary outcomes included HIV-related mortality (all-cause), morbidity (including type and duration of episodes of opportunistic infections, malignancies, incidence of AIDS, and hospital admissions). Secondary outcomes included change in appetite, nausea, mood, pain, quality of life, body composition, hematological and nutritional markers, cognitive function, respiratory function and effect on pharmacokinetics of antiretrovirals and development of dependence or adverse sociological effects. The use of cannabis posed challenges for blinding due to the psychoactive effects⁵².

Only 1 study involving 62 patients examined the effects of cannabinoids on immune function and parameters associated with HIV infection. Participants in this study were assigned

to either a 3.95% THC marijuana cigarette, a 2.5 mg dronabinol capsule or a placebo capsule three times per day for 21 days. For the marijuana group, there was a statistically significant increase in CD4 counts from baseline vs. placebo and for the dronabinol group there was a trend towards statistical significance. Neither CD4 nor CD8 cell counts were adversely affected, and the pharmacokinetic component of the study did not reveal any clinically significant interactions that would require dose adjustments of protease inhibitors²². Adverse effects across studies included concentration difficulties, fatigue, sleepiness or sedation, increased duration of sleep, reduced salivation and thirst which improved upon discontinuation. The authors of a systematic review on the use of cannabis for reducing morbidity and mortality in HIV concluded that 1) evidence for substantial effects on morbidity and mortality is currently limited and 2) evidence for safety and efficacy of cannabis is lacking. Studies have been of short duration, in small numbers of patients and have focused on short-term measures of efficacy⁵². Furthermore, no study examined the effects of cannabinoids on inflammatory markers or HIV reservoir markers through a randomized trial.

More recently, Manuzak *et al.* published an observational study assessing the effect of cannabis use on peripheral immune cell frequency, activation, and function in 198 people living with HIV⁵³. Individuals were grouped into heavy, medium, or occasional cannabis users or noncannabis users as determined by the quantify cannabis metabolite 11-nor-carboxy-

tetrahydrocannabinol (THCCOOH) detected in plasma by mass spectrometry. They found that persons with heavy cannabis use had lower frequencies of HLA-DR+CD38+CD4+ and CD8+ T-cell frequencies compared to people living with HIV⁵³. Furthermore, heavy cannabis use was associated with decreased frequencies of pro-inflammatory intermediate (CD14++CD16+) and non-classical (CD14+CD16+) monocyte subsets⁵³. They also documented a reduction in antigen-producing cells secreting pro-inflammatory cytokines IL-23 and tumor necrosis factor (TNF)- α ⁵³. Rizzo *et al.* also demonstrated that levels of circulating CD16 monocytes and interferon-gamma-induced protein (IP)-10 from people living with HIV who either were or were not cannabis users⁵⁴. Lower levels of CD16+ monocytes and plasma IP-10 were found in cannabis users compared to non-cannabis users⁵⁴. However, this study did not quantify the level of cannabis exposure in the two groups. Although these studies demonstrated favorable associations between inflammation and cannabis use, it must be borne in mind that both of these studies were observational only. As these studies were not randomized controlled trials, it is possible that people living with HIV who used cannabis in these studies differed in other significant ways from PLWHIV who did not use cannabis.

Study rationale

Cannabis may hold many potential therapeutic benefits for people living with HIV due to its promising anti-inflammatory and anti-fibrotic effects. Before adequately-powered interventional studies can be designed to study cannabis as a therapy for specific conditions associated with chronic inflammation and fibrosis, a key first step will be to demonstrate that cannabinoid use in a clinical trial is feasible and that they have a favorable safety and tolerability profile. As such, we propose a proof-of-concept pilot study to examine the feasibility, safety and tolerability of cannabinoid oils consumed orally in people living with HIV on effective ART. As a secondary objective, we will examine the effect of cannabinoid oils on immune profiles, including levels inflammatory markers associated with HIV disease progression and frequencies of activated and senescent CD4 and CD8 T-cells. Frequencies of regulatory T cells and various subsets of Th17 will also be assessed. Finally, an exploratory objective will be to study the effect of cannabinoid oils on markers of HIV persistence and the composition of the gastrointestinal microbiome.

We propose to use combination therapy of THC:CBD oils in capsule format (TN-TC11LM and TN-TC19LM capsules) ingested orally to examine these outcomes. Although research to date involving HIV/SIV has examined THC, data from *in vitro*, animal and human studies suggests that CBD has favorable anti-inflammatory effects and the combination of CBD with THC increases tolerability⁵⁵⁻⁵⁹.

Methods/Design

Study design

This is a randomized, open-label, interventional study (the Canadian HIV Trials Network (CTN) number PT028) whereby capsules containing CBD:THC oils are consumed for 12 weeks to assess safety and tolerability. Their ability to reduce immune activation (as determined by percentage of activated CD8+CD38+HLA-DR+ T-cells), size of the peripheral HIV reservoir and change in gastrointestinal microbiome composition will also be examined. Participants will continue to take their ART treatments as prescribed throughout the study.

Setting

Recruitment of participants will occur at the Chronic Viral Illness Service (CVIS), Royal Victoria Hospital (Glen campus) of the McGill University Health Centre (MUHC), the largest academic HIV clinic in Canada.

Recruitment and enrollment

Study staff at the CVIS will conduct chart reviews of prospective people living with HIV ahead of their clinic visits to determine which persons have had suppressed viral load for at least 3 years on ART. The patient chart will be flagged, and if the treating HIV physician believes the person to be suitable for the study, the physician or study staff will approach potential trial participants at their clinic visit. The trial staff will inform the patients about the trial and invite him or her for eligibility screening and possible trial enrolment. Participant eligibility will be documented and written informed consent obtained for eligible patients by the study coordinator. The study coordinator will systematically document all individuals who have been approached for the study in addition to reasons for acceptance and refusal to participate in the study. Individuals who wish to discuss their participation in the study with their treating physician and or family and friends will be have the opportunity to do and may enroll a their next scheduled clinic visit. Following enrolment, participants will be followed during the study by the principal investigator and study coordinator at the CVIS.

Inclusion criteria

Eligible participants must meet the following criteria within 4 weeks prior to beginning the cannabinoid capsules to be considered eligible for study entry: 1) documented HIV infection

by Western blot, enzyme immunoassay or viral load assay; 2) aged 18 or older; 3) viral load <40 copies/mL for at least the last 3 years (maximum 2 blips <500 copies/ml allowed); 4) no cannabinoid use for at least 1 month prior to enrolment with negative baseline cannabinoid urine screen; 5) able to communicate in either English or French.

Exclusion criteria

Individuals who meet any of the following criteria will be ineligible to participate: 1) using cannabinoid-containing products outside of the study or within 4 weeks of study commencement; 2) pregnant, breastfeeding or planning to become pregnant during the course of the study; 3) enrolled in a separate study involving administration of medication, vitamin, supplement or herbal product; 4) active intravenous drug use; 5) active substance dependence; 6) prior history of hypersensitivity to cannabis or cannabis-containing products; 7) known or suspected allergy to sunflower lecithin oil; 8) active opportunistic infection or malignant condition; 9) unintentional weight loss of 10 % or more of body weight in the last 6 months; 10) unstable angina or acute cardiac event in the past year; 11) active psychiatric disorder or history of psychiatric depression (other than mild depression or anxiety); 12) on antipsychotic medication; 13) known or suspected family history of schizophrenia or severe personality

disorder; 14) serious cardiovascular disease such as ischemic heart disease, arrhythmias, poorly controlled hypertension, or severe heart failure; 15) anemia (Hemoglobin <100 g/L); 16) active liver disease or unexplained persistent elevations of serum transaminases; 17) Co-infection with Hepatitis B or C (positive HBsAg or positive anti-HBc antibodies with a detectable HBV DNA viral load or positive anti HCV antibodies with a detectable HCV RNA viral load); 18) alanine aminotransferase (ALT) or Aspartate aminotransferase (AST) or alkaline phosphatase >2.5 x upper limit of normal (ULN); 19) opportunistic infection in the last month as determined by the treating physician; 20) renal dysfunction; 21) unstable psychological or psychiatric condition as determined by the treating physician; 22) holding employment which requires operation of heavy machinery or which requires undergoing drug screening (ie, pilot or police officer); 23) concurrent use within the past 8 week of anabolic hormones, prednisone, IL-2 or other agents known to alter immune function.

Study intervention

The study medications are TN-TC11LM and TN-TC19LM capsules which contain THC:CBD in a ratio of 1:1 (2.5 mg/2.5 mg) and 1:9 (5 mg/45 mg), respectively. Participants will be advised to gradually increase the number of capsules they take based on the suggested titration scheme

presented in Tables 1 and 2, until a daily maximum is reached. These maximum amounts are comprised of 10 capsules of TN-TC11LM (25 mg THC/25 mg CBD total per day) or 3 capsules of TN-TC19LM (15 mg THC: 135 mg CBD for TN-CT19L) per day. These doses were selected as in a clinical trial for neuropathic pain, doses equivalent to 2.5 mg of THC were well-tolerated³⁴. More recently, among patients (ages 2-55 years) with the Lennox-Gastaut syndrome, cannabidiol at a dose of 10 or 20 mg per kilogram per day resulted in greater reductions in the frequency of drop seizures than placebo and was well-tolerated overall other than for an increase liver aminotransferase concentrations⁶⁰. Due to person-to-person variability in the ability to metabolize and tolerate cannabinoids⁶¹, we have opted for patients to titrate their dose of medication to a range where they are comfortable as the titration method of dosing has proven successful in other clinical trials involving cannabinoids³⁴.

Table 1: Recommended titration schedule for Group 1 – Low CBD dose TN-TC11LM oral capsules

Weeks	Daily Dose	Number of capsules
1	5 mg THC/ 5 mg CBD	1 capsule twice daily, taken orally
2	10 mg THC/ 10 mg CBD	2 capsules twice daily, taken orally (4 capsules per day)
3	15 mg THC/ 15 mg CBD	2 capsule three times daily, taken orally (6 capsules per day)
4	20 mg THC/ 20 mg CBD	2 capsules four times daily, taken orally (8 capsules per day)
5-12	25 mg THC/25 mg CBD	2 capsules 5 times daily, taken orally (10 capsules per day)

Group 1: Low CBD dose TN-TC11LM oral capsules (2.5 mg THC/2.5 mg CBD capsules). This group will be advised to start by taking 1 capsule twice daily for 1 week (5 mg THC/5 mg CBD) and increase the number of capsules as tolerated to a maximum of 10 capsules daily by weeks 5-12 (25 mg THC/25 mg CBD total per day). Participants will record the times and dates of all capsules consumed in a logbook.

Table 2: Suggested titration schedule for Group 2 – High CBD dose TN-TC19LM oral capsules

Weeks	Daily Dose	Number of capsules
1	5 mg THC/ 45 mg CBD	1 capsule daily, taken orally
2	10 mg THC/ 90 mg CBD	1 capsule twice daily, taken orally (2 capsules per day)
3	15 mg THC/ 135mg CBD	1 capsule three times daily, taken orally (3 capsules daily)
4	20 mg THC/ 180 mg CBD	1 capsule four times daily, taken orally (4 capsules)
5-12	25 mg THC/ 225 mg CBD	1 capsule five times daily, taken orally (5 capsules)

Group 2: High CBD dose TN-TC19LM (5 mg THC/45 mg CBD capsules). This group will be advised to start by taking 1 capsule once daily for 1 week (5 mg THC/45 mg CBD) and increase the number of capsules as tolerated to a maximum of 10 capsules daily by week 5(25 mg THC/225 mg CBD total). Participants will record the times and dates of all capsules consumed in a log book.

Randomization

After eligibility is confirmed and written informed consent obtained, participants will be randomized to either TN-TC11LM (group 1) or TN-TC19LM (group 2) capsules which contain THC:CBD in a ratio of 1:1 and 1:9, respectively. Prior to study commencement, a statistician unassociated with the study will develop a randomization scheme using SAS and input into a password-protected web-based randomization system. Variable block sizes of 2 and 4 will be used. Participants will be assigned to either Group 1 vs. Group 2 based on the pre-designated allocation code. As this is an unblinded study, participants and study staff will be aware of the group to which the participant has been randomized. A computerized audit trail will track date and time of allocation, patient study identification number and treatment allocation. The randomization group will be recorded in the study log, which will be accessible to the sponsor/medical manager and study coordinator.

Measurements

At the screening visit, clinical information will be collected from each participant including age, ethnicity, list of current medications, dosage, date of treatment initiation, psychiatric disorders, duration of HIV infection, current ART regimen and duration of ART

regimen months, ART history, CD4+ T cells count within the past 3 months, nadir CD4+ T cells count, CD4/CD8 ratio, duration of plasma viral load suppression and any preexisting medical conditions, signs or symptoms. Information will also be collected on whether the individual consumed cannabis in the past, the form(s) in which it was consumed, frequency of use and reasons for use.

Clinical parameters

Scheduled visits will occur to monitor safety and tolerability, as per the visit schedule depicted in Table 3. Visits will include physical exam with vital signs, weight, occurrence of adverse events (AEs and concomitant medications) and the presence of common symptoms associated with cannabinoids including dizziness, nausea, headaches, appetite or mood changes. At visits, blood for some or all of the following will be collected: CD4+ T cells count, CD8+ T cells, CD4/CD8 ratio, plasma viral load, Complete Blood Count (CBC), aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP), total bilirubin, urea, creatinine and blood glucose, T-cell activation and inflammatory markers and testing for syphilis if the participant tested positive during the 4 weeks prior to beginning consuming the study capsules. A stool sample for analysis of the bacterial and fungal

microbiome assessment will also be collected prior to beginning the study capsules. Participants will be enrolled in the study for up to 15 weeks but will consume capsules for a period of 12 weeks. Participants will undergo screening tests and eligibility assessment within 4 weeks prior to initiating study capsules. Participants will then undergo assessments after the first week of capsule consumption and every 2 weeks thereafter. A second stool sample for bacterial and fungal microbiome analysis will be collected during the final week of capsule consumption. The final visit will occur 2 weeks after study drug cessation.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47

Table 3. Schedule of Visits and Procedures

Visit Type	Screening	Visit 1 - Baseline 1	Visit 2 - Baseline 2	Visit 3	Visit 4	Visit 5	Visit 6	Visit 7	Visit 8	Visit 9 – End of Tx Visit	Visit 10 – Final Study Visit
Visit Window	-4 to -1 weeks	W -1	D1, W0	D1, W1 (± 4 days)	D1, W2 (± 4 days)	D1, W4 (± 4 days)	D1, W6 (± 4 days)	D1, W8 (± 4 days)	D1, W10 (± 4 days)	D1, W12 (± 4 days)	D1, W14 (± 4 days)
Procedures:											
Eligibility criteria assessment	X										
Informed Consent	X										
Medical History	X										
Pregnancy Test (urine) ¹	X										
Cannabinoids Screen (urine)	X										
Hepatitis B, C and syphilis	X ³										
Randomization			X								
Physical Exam			X	X	X	X	X	X	X	X	X
Hematology and chemistry profiles ²	X			X	X	X	X	X	X	X	
Viral load, CD4 and CD8	X				X		X			X	
Immune activation and inflammatory markers			X	X	X		X			X	X
HIV reservoir assays		X	X	X	X		X	X		X	X
Nasal swab and stool specimen for microbiome assessment		X								X	
WHOQOLHIV-BREF scale			X				X			X	
EQ-5D questionnaire			X				X			X	
POMS questionnaire			X				X ⁵			X	
Study medication dispensed			X	X	X	X	X	X	X		
Study Drug Compliance ⁴				X	X	X	X	X	X	X	
ART Compliance ⁴		X	X	X	X	X	X	X	X	X	X
Alcohol Intake		X	X	X	X	X	X	X	X	X	X
Adverse Events				X	X	X	X	X	X	X	X

Visit Type	Screening	Visit 1 - Baseline 1	Visit 2 - Baseline 2	Visit 3	Visit 4	Visit 5	Visit 6	Visit 7	Visit 8	Visit 9 - End of Tx Visit	Visit 10 - Final Study Visit
Visit Window	-4 to -1 weeks	W -1	D1, W0	D1, W1 (± 4 days)	D1, W2 (± 4 days)	D1, W4 (± 4 days)	D1, W6 (± 4 days)	D1, W8 (± 4 days)	D1, W10 (± 4 days)	D1, W12 (± 4 days)	D1, W14 (± 4 days)
Procedures:											
Concomitant Medications/Therapies	X	X	X	X	X	X	X	X	X	X	X

¹ If urine test is positive, perform serum pregnancy test.

² Complete blood count; AST, ALT, ALP, total bilirubin, urea, creatinine and blood glucose.

³ If participant tests positive for Hepatitis B and C, he/she will no longer be eligible for the study. If participant tests positive for syphilis, he or she will be treated for syphilis according to clinical care guidelines and will still be eligible to participate in the study. The need for syphilis treatment and follow-up testing (usually in 6 and 12 months) will be discussed between the Sponsor and the investigator at the CVIS. It will be up to the investigator to ensure proper follow-up and management of the syphilis, as this is part of standard of care.

⁴ Assessed by reviewing log book provided to each participant

⁵ Review individual POMS questionnaires completed at Visits 2 and 6 with each participant

1

2

3

4

5 **Medication adherence**

6

7

8 Participants will be asked to keep a log book in which they enter the number of TN-

9

10

11 TC11LM or TN-TC19LM capsules consumed, the time, as well any adverse effects they noted

12

13

14 and the timing of these adverse effects relative to capsule intake. Furthermore, individuals will

15

16

17 be asked to record whether or not they took their ART that day or whether any doses were

18

19

20 missed. Participants will be asked to bring their log books with them to study visits and the

21

22

23 coordinator will photocopy this information.

24

25

26

27

28

29

30 **Quality of life and mood assessment**

31

32

33

34 Questionnaires measuring quality of life (World Health Organization Quality of Life

35

36

37 HIV-BEF (WHOQOLHIV-BREF) and the EQ-5D) and mood (Profile of Mood States (POMS)

38

39

40 will be administered at baseline, midway through the study and at the end of the study, as

41

42

43 outlined in Table 3. WHOQOLHIV-BREF consists of 31 items which measure the following

44

45

46 domains: physical health, psychological health, social relationships and environment. It is a

47

48

49 shorter version of the original instrument (WHOQOL) and is more convenient for use in clinical

50

51

52 trials, taking approximately 10 minutes to complete. EQ-5D is a descriptive questionnaire

53

54

55 examining 5 dimensions: 1) mobility, 2) self-care, 3) usual activities, 4) pain/discomfort, and 5)

56

57

58

59

60

anxiety/depression. Each dimension has 5 levels: No problem, slight problems, moderate problems, severe problems and extreme problems. The participant indicates the state of his/her health by ticking the box most appropriate to the statement in each of the 5 dimensions. This decision results in a 1-digit number that expresses the level selected for that dimension. The digits for the 5 dimensions can be combined into a 5-digit number that describes the participant's health state. It takes about 10 minutes to complete. The Profile of Mood States (POMS) questionnaire measures the following 6 factors: 1) Tension-Anxiety, 2) Anger-Hostility, 3) Fatigue-Inertia, 4) Depression-Dejection, 5) Vigor-Activity, and 6) Confusion-Bewilderment. It is very sensitive to non-clinical changes in mood states and takes approximately 5 minutes to complete. These questionnaires will be administered by a trained research coordinator.

Research hypothesis

THC:CBD oils consumed orally – as TN-TC11LM and TN-TC19LM oral capsules – will be safe and well-tolerated in PLWHIV. They will also be associated with a reduction in markers of inflammation, reduction in frequency of activated T cells and reduction in HIV reservoir size.

Study outcome measures

The primary objective is to evaluate the safety and tolerability of TN-TC11LM and TN-TC19LM oral capsules in PLWHIV on effective ART. The primary between group comparison is the percentage of participants without any signs of significant toxicity; percentage of participants who are able to complete the study and scores on the WHOQOLHIV-BREF Scale, EQ-D5 and POMS questionnaires from week 0 to week 12 are secondary outcomes that will also be compared between groups. The secondary objective is to determine the effect of TN-TC11LM and TN-TC19LM oral capsules on frequency of activated T-cells and markers of inflammation association with HIV disease progression. Exploratory objectives are to determine the effect of TN-TC11LM and TN-TC19LM oral capsules on 1) the size of the peripheral HIV reservoir 2) the composition of the gastrointestinal bacterial and fungal microbiome.

Safety and tolerability

Safety will be assessed at regular intervals (Table 3) by vital signs and adverse effects (AE) monitoring, as reported by the participant and actively sought at each study visit by the coordinator or physician. Biological safety will be evaluated by hematology, biochemistry and other clinical, laboratory or other diagnostic tests done on participants during the course of the study. Lab results for all participants for assessed safety variables will be reviewed by the trial investigator. The Data safety monitor will also review safety information. Toxicity of TN-

TC11LM and TN-TC19LM will be assessed using the World Health Organization (WHO) toxicity scale. All AEs, regardless of the grade, will be documented and it will be noted whether or not these symptoms were already present at baseline. Any AEs that occur during the study will be evaluated by the trial investigators and grade 3 and 4 AEs will be recorded on the CRFs. If required, blood specimens will be collected for hematology and biochemistry tests. Participants having AEs will be monitored with relevant clinical assessments and laboratory tests, as determined by the trial investigator. The trial investigator will report ongoing AEs at the completion of the clinical study to the primary treating physician at the CVIS who will determine the need for and provide standard medical care. The trial investigator will ensure that the event is satisfactorily resolved or that no additional follow-up is needed. Any participant who discontinues the study for an unresolved clinically significant AE will be followed until satisfactory clinical resolution is achieved and the AE recorded on the case report form (CRF), regardless of severity grading. AEs that may be related to TN-TC11LM and TN-TC19LM will be managed by dose reduction of TN-TC11LM and TN-TC19LM. TN-TC11LM and TN-TC19LM will be discontinued permanently in the event of any life-threatening AEs.

1

2

3

4 **T-cell subsets and Immune activation**

5

6

7

8 The frequency of different CD4+ and CD8+ T cell subsets will be defined using multi-

9

10 parameter flow cytometry (BD Fortessa X-20) in peripheral blood. The expressions of CD3,

11

12 CD4 and/or CD8, CD45RA, CCR7 and CD27 will be used to measure the frequency of naïve

13

14 (CD45RA⁺CCR7⁺CD27⁺), central memory (CD45RA⁻CCR7⁺CD27⁺), transitional memory

15

16 (CD45RA⁻CCR7⁻CD27⁺), effector memory (CD45RA⁻CCR7⁻CD27⁻) and terminally

17

18 differentiated (CD45RA⁺CCR7⁻CD27⁻) cells. Regulatory T (Treg) cells will be defined as

19

20 CD3⁺CD4⁺CD25^{high}FoxP3^{high}CD127^{low} cells. Expression of the CD39 and CD73 ectoenzymes

21

22 involved in Treg-mediated immunosuppression and HIV disease progression (via the adenosine

23

24 pathway) will be also assessed ^{62 63}. Various subsets of Th cells (T helper) will be defined as

25

26 Th1 (CD45RA⁻CCR6⁻CCR4⁻CXCR3⁺), Th2 (CD45RA⁻CCR6⁻CCR4⁺CXCR3⁻), Th17 (CD45RA⁻

27

28 CCR6⁺CCR4⁺CXCR3⁻) and Th1/Th17 (CD45RA⁻CCR6⁺CCR4⁻CXCR3⁺). Levels of CD8+ and

29

30 CD4+ T cell immune activation (CD38/HLA-DR co-expression) and senescence (CD28⁻CD57⁺)

31

32 will be also assessed on all T cell subsets. These markers will be assessed at week 0, 1, 2, 6, 12

33

34 and 14.

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54 **Inflammatory markers assessments**

55

56

57

58

59

60

Plasma levels of various inflammatory markers including interferon- α , interleukin (IL)-1 β , IL-6, IL-10, IL-17, TGF- β , interferon-gamma-induced protein (IP)-10, will be assessed via Luminex (Millipore) and levels of d-dimer, C-reactive protein, and markers of microbial translocation lipopolysaccharide and sCD14 will be assessed by ELISA in batch from blood drawn at weeks 0, 1, 2, 6, 12 and 14.

Peripheral blood HIV reservoir size

Blood for HIV reservoir assessment will be collected at 2 time points prior to cannabis initiation (1 week prior to cannabinoid capsule initiation and immediately prior to cannabinoid capsule initiation) to account for normal fluctuations in baseline levels of HIV persistence markers. Subsequently, virological measures will be done at weeks 1, 2, 6, 8, 12, 14. CD4+ T cells isolated from PBMCs by magnetic negative selection. To capture all infected cells, the frequency of cells harbouring total and integrated HIV DNA will be measured using well-established assays on a total of 500,000 cells (sensitivity of 1 copy/reaction)^{17 64}. As most of the HIV genomes are defective, the recently developed “tat/rev induced limiting dilution assay” (TILDA), which provides a more functional measurement of the HIV reservoir⁶⁵, will be employed. To assess if residual levels of viral replication may occur, we will measure 2-LTR

circles, which are proposed to be a surrogate marker for ongoing HIV replication during ART⁶⁴
^{66 67}. Specifically, this combination of assays will indicate if cannabis has an impact on the size
of the total reservoir (DNA), the functional reservoir (TILDA) and ongoing viral replication (2-
LTR circles). Measurements will be performed in batch.

Gastrointestinal Microbiome Composition

A stool sample without preservative will be collected from each participant at the
beginning of the study prior to consuming the capsules and during the final week (week 12) of
capsule consumption. Specimens will be stored at -80°C until analyzed in batch, as previously
described⁶⁸. Bacterial DNA will be extracted with PCR amplified targeting of the 16S rRNA
gene using universal primers which flank the V3-V4 region of the 16S gene modified with the
addition of TruSeq Illumina adapters, also as previously outlined⁶⁹. Internal Transcribed Spacer
for Fungal DNA extraction will be used for fungi. PCR amplification, PCR amplicon
quantification and sequencing will be performed as previously described^{68 69}.

Sample size and statistical analyses

In this proof-of-concept study, we are exploring a phenomenon with little *in vivo* data and with a limited study budget. For this reason we have chosen a convenience sample of 26 participants, 13 per arm, without formal power calculations. This number of individuals will enable us to assess feasibility (willingness of patients to participate, attend study visits and complete questionnaires, numbers of drop-out participants) as well as safety and tolerability. The data obtained will help to guide future sample size calculations for future studies. Although the small number of participants may result in wide confidence intervals for adverse events, this number of participants should give us an idea about the degree of variability for continuous outcomes we are measuring.

For the primary endpoint, the proportions of participants without any signs of significant toxicity (Grades 0-2 scores on the WHO toxicity scale), proportions of participants who complete the study and scores on the WHOQOLHIV-BREF, EQ-5D and POMS questionnaires will be examined using descriptive statistics. We will also compare these proportions for Group 1 vs. Group 2 using a Fisher's exact test. For Quality of Life and mood measures, we will use analysis of covariance with 12 week score as outcome and baseline score as covariate and treatment as independent variable. With regards to the POMS questionnaire, we will consider only overall scores (and not sub-scores) due to the small sample size which would make comparisons of the sub-scale inappropriate.

Immune activation levels for Groups 1 vs. Group 2 at week 12 will be compared using analysis of co-variance with adjustment for the week 0 activation levels. The mean change and associated 95% confidence interval will be reported for each of the secondary endpoints described above. Furthermore, the study drug treatment period will be compared to the baseline period with each arm. If the treatment effect is similar within the two arms, then an analysis of the treatment effect will be pooled over the two arms, using analysis of covariance. The change in immune activation levels following discontinuation of study drug (i.e., from week 12 to week 14) will be reported as a mean with corresponding 95% confidence interval. A 50% reduction will be considered significant⁷⁰.

Analogous analyses will be conducted for reservoir assessments other outcome measures listed above as endpoints. A signed rank (non-parametric) test will be used to compare number of copies of total and integrated DNA at baseline 2 vs. at week 12. Group differences in the change of HIV reservoir size from baseline to 12 weeks will be assessed by the Mann-Whitney *U* test. Wilcoxon signed-rank test will be used to compare the HIV reservoir and inflammatory markers in blood samples of the same patient from baseline to 12 weeks. At least a 2-fold decrease in frequency of infected cells in both groups from baseline to 12 weeks of treatment will be considered significant⁷¹. Microbiome composition will be described with regards to the

frequencies of microorganisms families for the groups at baseline and then at 12 weeks of treatment. Due to the exploratory nature of this objective, no formal statistics will be applied.

Data management

All clinical data and electronic files will be stored in the secure environment of the CVIS of the MUHC. All data published will be anonymized. Only researchers affiliated with the study will have access to participant data. Study progress and safety will be evaluated in an ongoing fashion by the principal and co-investigators. The study will be monitored for safety and ongoing progress by a standing Data and Safety Monitoring Committee (DSMC) of clinicians and methodologists established by the Canadian HIV Trials Network. The committee meets every six months or as needed.

Storage of biological specimens

All biological specimens will be stored at the CVIS of the MUHC for analysis in the current trial and for future use in additional studies.

1

2

3

4 **Ethics and dissemination**

5

6

7

8 Written informed consent will be obtained from all study participants. The study

9

10 protocol and informed consent have been approved by the Research Ethics Board of the McGill

11

12 University Health Centre (MUHC-2018-4336) and is in the process of being reviewed by Health

13

14 Canada’s Therapeutic Product Directorate. The study will be conducted in accordance with the

15

16 application Health Canada regulations, International Conference on Harmonization guidelines on

17

18 current Good Clinical Practice and the Declaration of Helsinki. Patient enrollment for this trial is

19

20 anticipated to begin August 2018.

21

22

23

24

25

26

27

28

29 Regardless of outcome, trial results will be disseminated through scientific peer review

30

31 publication, international and national conferences and the CTN according the SPIRIT (Standard

32

33 Protocol Items: Recommendations for Interventional Trials) and CONSORT (Consolidated

34

35 Standards of Reporting Trials) guidelines for transparent reporting of trials^{72 73}. CTC will be

36

37 responsible for initially drafting the manuscripts and professional writers will not be used for any

38

39 of the publications. Authorship will be determined based on criteria defined by the International

40

41 Journal of Medical Editors⁷⁴. We aim to write the manuscript of the final results within 6 months

42

43 of completing the study. Participants who have been involved in the trial will be given the

44

45 option of having a summary of results sent to them.

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

Patient and Public Involvement

The CTN Community Advisory Committee (CAC) was involved in the peer review process of this study proposal, deemed that the research questions addressed were of very high priority to people living with HIV and voted for the funding of this study. The CAC's critiques of the initial proposal were taken into account in the revised proposal. Two members of the CAC (SM and EM) were involved in finalizing the study design, inclusion/exclusion criteria, outcome measures and monitoring plans and are formal study investigators and co-authors. Preliminary and final results of the study will be shared with community members, patient participant and the public at bi-annual CTN meetings, through the CTN newsletter and on the CTN website in addition to the annual Canadian Association of HIV Research meeting.

Discussion

Since the advent of ART, people living with HIV now have a longevity which approaches that of their HIV-uninfected counterparts but have a higher burden of non-communicable comorbidities including cardiovascular, pulmonary, renal and hepatic diseases^{1 2}. Heightened inflammation in people living with HIV despite ART is believed to be the driving force behind the increased rates of non-infectious comorbidities. Similarly, chronic immune activation fosters HIV persistence¹⁰⁻¹⁵. As cannabinoids possesses both anti-inflammatory and

anti-fibrotic properties¹⁹, cannabinoids may represent a feasible method to reduce immune activation and enhance immune profile. This, in turn, may hasten the progression of non-opportunistic complications associated with HIV.

In this pilot study, our primary objective is to assess the safety and tolerability of TN-TC11LM and TN-TC91LM taken by people living with HIV on suppressive ART. We hypothesize that these agents will be safe and well tolerated in people living with HIV, given that similar products are safe and well-tolerated in other populations. Sativex® is currently licensed as an adjunctive treatment for symptomatic relief of spasticity in adult patients with Multiple Sclerosis (MS) who have not responded adequately to other therapy⁷⁵⁻⁷⁷. It is an oromucosal spray containing CBD and THC in 1:1 ratio. Marinol®, which is a synthetic THC-containing capsule, is currently used for the treatment of anorexia associated with weight loss in persons with Acquired Immunodeficiency Syndrome (AIDS) and nausea and vomiting associated with cancer chemotherapy in patients with insufficient response to conventional antiemetics⁷⁸. It has been estimated that 5,472 patients have been exposed to Sativex® and there have been no safety concerns identified and the product remains well-tolerated. The primary safety concerns of both Sativex® and Marinol® are consistent with the known pharmacological activity of cannabinoids. The primary safety concerns associated with Sativex® included abuse potential, cardiovascular effects and central nervous system adverse effects⁷⁹. Although there is some evidence to suggest

that individuals can develop “cannabis use disorder”, individuals do not develop the same extremes of behavior as observed with other drugs of abuse⁸⁰. In two clinical trials, nabiximols such as Sativex® have been used in two clinical trials whereby treatment was abruptly ceased to study whether withdrawal symptoms would develop^{81 82}. In both studies, no withdrawal syndromes were observed⁸³. Cannabinoids have cardiovascular effects that include tachycardia and fluctuations in blood pressure, including episodes of postural hypotension. Therefore these agents should not be used in patients with serious cardiovascular disease, such as ischemic heart disease, arrhythmias, poorly controlled hypertension or severe heart failure. THC has complex effects on the CNS and should not be used in patients with a personal or strong family history of psychosis. Examples of such conditions include schizophrenia and affective psychosis since symptoms of these disorders may be aggravated by cannabinoids. In multiple sclerosis (MS) patients receiving Sativex® in clinical studies, psychiatric-related adverse effects included disorientation, depression including depressed mood, dissociation, euphoric mood, hallucination, hallucinations (auditory and visual), illusions, paranoia, suicidal ideation and delusional perception⁸⁴. Interestingly, there is some evidence to demonstrated that CBD may actually improve psychotic symptoms in persons suffering from schizophrenia⁸⁵.

For Marinol®, which contains only THC, the most frequency reported adverse effects experienced by patients with AIDS during placebo controlled clinical trials involved CNS and

were reported in 33% of patients receiving Marinol®. About 25% reported a CNS adverse event during the first 2 weeks and about 4% reported such an event each week for the next 6 weeks thereafter⁸⁶. By combining CBD with THC, we anticipate that tolerability will be greatly enhanced. When combined with THC, CBD reduces the risk for many adverse effects⁸⁵. Furthermore, individuals will be instructed to titrate up the dose based on their own tolerability and reduce the dose if they experience any undesirable effects. Furthermore, due to the extremely low levels of CB1 receptors in the brainstem⁸⁷, death due to overdosing on cannabis or cannabinoids alone has never been described.

The study medications and doses were chosen after a lengthy review of the existing literature and discussion with experts in the field of pain management. A high degree of inter-individual variability in metabolism following administration of cannabinoids is observed due to polymorphisms in cytochrome isoenzymes⁶¹. Given that the therapeutic doses of cannabinoids are highly variable between individuals, a dose titration schedules are usually recommended. When used to treat specific conditions, persons may be told to increase the dose until they achieve adequate symptom relief without adverse effects. This method was observed to work well when used in the first cohort study on the long-term safety of medicinal cannabis for non-cancer chronic pain in seven Canadian clinics³⁴.

Given this is a pilot study, a convenience sample of 26 participants was selected without formal power calculation. If this study demonstrated that TN-TC11LM and/or TN-TC91LM are safe and tolerable in people living with HIV and can reduce systemic inflammation, future studies will be performed to address the potential of these agents to ameliorate specific conditions in people living with HIV. Should future studies be conducted, data generated from this trial will assist with power calculations. Similar to a study conducted by members of our group on the ability of Niaspan® (extended-release niacin) to reduce immune activation, as determined by percentage of activated CD8+ CD38+ HLA-DR+ T-cells, we decided that a 50% reduction in activated CD8+ CD38+ HLA-DR+ T-cells would be considered significant⁷⁰. This is based on previous reports indicating that a 10-fold difference exists between uninfected healthy controls and treated aviremic HIV-infected individuals in level of activated CD8+ CD38+ HLA-DR+ T-cells^{88 89}. All of the inflammatory mediators we selected for this study are known to drive immune activation^{1 6 90}.

In addition, we decided to make our objective examining the ability of TN-TC11LM and TN-TC91LM capsules to reduce the HIV reservoir, through the reduction of systemic inflammation, an exploratory objective. It is unclear if 3 months of treatment will be long enough to produce any meaningful reduction in the size of the HIV reservoir. Furthermore, it is unclear what reduction in reservoir size is required to have a meaningful effect on clinical

outcomes. As mentioned earlier, we will consider a 50% decrease in the number of HIV-infected cells at baseline vs. week 12 to be a significant reduction in the reservoir, based on a study by *Hill et al*⁷¹. To our knowledge, there is no other randomized clinical trial examining the effect of cannabinoids on inflammation and HIV reservoir size in people living with HIV.

Our study is unique in being the first randomized trial in the world to examine the association between ingestion of precise quantities of cannabinoids and effect on inflammation and peripheral HIV reservoir size. It is also noteworthy that we chose not to have a placebo arm as the effects of psychoactive effects of THC would be difficult to camouflage. Furthermore, we chose to use oral formulations of cannabinoids so that we could precisely control the dose ingested by participants. When cannabis is smoked or vaped, there is variability in the methods and duration of inhalation used by participants which can influence dosage of cannabis ingested. The oral administration option also removes undesirable pulmonary effects such as symptoms wheezing or breathlessness in addition to inhalation of toxic chemicals⁹¹. Of special interest to our group is the recent discovery that administration of oral cannabis with lipids leads to high levels of cannabinoids in the intestinal lymphatic system and prominent immunomodulation, as demonstrated by *Zgair et al.*²⁰ This finding is especially important given the prominent role of the mesenteric lymph nodes and gut to HIV persistence⁷. If oral cannabinoids can modify gut

microbiome and the enteric immune system favorably, larger clinical trials could be conducted to examine this phenomena in further detail.

The Canadian government has declared that cannabis' regulatory status will change from being an illegal substance to that of a legal substance in July 2018. Cannabis' change in regulatory status will likely stimulate more discussion amongst patients and physicians and thus physicians need to be informed about the potential risks and benefits of cannabis use. The change in the regulatory landscape will likely also foster more research into cannabis' therapeutic potential. We hope that this study will be a stimulus towards more open discussion between patients and their physicians and that it will reduce stigma associated with cannabinoids use. We also hope that this study will be the cornerstone for future studies investigating the therapeutic benefits of cannabis in PLWHIV and its potential not only at the individual level but also at the population level in the form of harm reduction strategies.

1. Kalayjian RC, Machekano RN, Rizk N, et al. Pretreatment levels of soluble cellular receptors and interleukin-6 are associated with HIV disease progression in subjects treated with highly active antiretroviral therapy. *The Journal of infectious diseases* 2010;201(12):1796-805. doi: 10.1086/652750
2. Funderburg NT, Mayne E, Sieg SF, et al. Increased tissue factor expression on circulating monocytes in chronic HIV infection: relationship to in vivo coagulation and immune activation. *Blood* 2010;115(2):161-7. doi: 10.1182/blood-2009-03-210179
3. The definition of emphysema. Report of a National Heart, Lung, and Blood Institute, Division of Lung Diseases workshop. *The American review of respiratory disease* 1985;132(1):182-5. doi: 10.1164/arrd.1985.132.1.182 [published Online First: 1985/07/01]
4. Brenchley JM, Price DA, Schacker TW, et al. Microbial translocation is a cause of systemic immune activation in chronic HIV infection. *Nature medicine* 2006;12(12):1365-71. doi: 10.1038/nm1511 [published Online First: 2006/11/23]
5. Estes JD, Harris LD, Klatt NR, et al. Damaged intestinal epithelial integrity linked to microbial translocation in pathogenic simian immunodeficiency virus infections. *PLoS pathogens* 2010;6(8):e1001052. doi: 10.1371/journal.ppat.1001052 [published Online First: 2010/09/03]
6. Deeks SG, Tracy R, Douek DC. Systemic effects of inflammation on health during chronic HIV infection. *Immunity* 2013;39(4):633-45. doi: 10.1016/j.immuni.2013.10.001 [published Online First: 2013/10/22]
7. Costiniuk CT, Angel JB. Human immunodeficiency virus and the gastrointestinal immune system: does highly active antiretroviral therapy restore gut immunity? *Mucosal immunology* 2012;5(6):596-604. doi: 10.1038/mi.2012.82 [published Online First: 2012/08/30]
8. Dillon SM, Lee EJ, Donovan AM, et al. Enhancement of HIV-1 infection and intestinal CD4+ T cell depletion ex vivo by gut microbes altered during chronic HIV-1 infection. *Retrovirology* 2016;13:5. doi: 10.1186/s12977-016-0237-1
9. Vujkovic-Cvijin I, Dunham RM, Iwai S, et al. Dysbiosis of the gut microbiota is associated with HIV disease progression and tryptophan catabolism. *Science translational medicine* 2013;5(193):193ra91. doi: 10.1126/scitranslmed.3006438
10. El-Sadr WM, Lundgren J, Neaton JD, et al. CD4+ count-guided interruption of antiretroviral treatment. *The New England journal of medicine* 2006;355(22):2283-96. doi: 10.1056/NEJMoa062360 [published Online First: 2006/12/01]
11. Baker JV, Peng G, Rapkin J, et al. Poor initial CD4+ recovery with antiretroviral therapy prolongs immune depletion and increases risk for AIDS and non-AIDS diseases. *Journal of acquired immune deficiency syndromes* 2008;48(5):541-6. doi: 10.1097/QAI.0b013e31817bebb3

12. Baker JV, Peng G, Rapkin J, et al. CD4+ count and risk of non-AIDS diseases following initial treatment for HIV infection. *Aids* 2008;22(7):841-8. doi: 10.1097/QAD.0b013e3282f7cb76
13. Kim H, Perelson AS. Viral and latent reservoir persistence in HIV-1-infected patients on therapy. *PLoS Comput Biol* 2006;2(10):e135. doi: 10.1371/journal.pcbi.0020135
14. Lu W, Mehraj V, Vyboh K, et al. CD4:CD8 ratio as a frontier marker for clinical outcome, immune dysfunction and viral reservoir size in virologically suppressed HIV-positive patients. *Journal of the International AIDS Society* 2015;18:20052. doi: 10.7448/IAS.18.1.20052
15. Rouzioux C, Richman D. How to best measure HIV reservoirs? *Current opinion in HIV and AIDS* 2013;8(3):170-5. doi: 10.1097/COH.0b013e32835fc619
16. Chun TW, Carruth L, Finzi D, et al. Quantification of latent tissue reservoirs and total body viral load in HIV-1 infection. *Nature* 1997;387(6629):183-8. doi: 10.1038/387183a0 [published Online First: 1997/05/08]
17. Chomont N, El-Far M, Ancuta P, et al. HIV reservoir size and persistence are driven by T cell survival and homeostatic proliferation. *Nature medicine* 2009;15(8):893-900. doi: 10.1038/nm.1972 [published Online First: 2009/06/23]
18. Chomont N, DaFonseca S, Vandergeeten C, et al. Maintenance of CD4+ T-cell memory and HIV persistence: keeping memory, keeping HIV. *Current opinion in HIV and AIDS* 2011;6(1):30-6. doi: 10.1097/COH.0b013e3283413775
19. Zurier RB, Burstein SH. Cannabinoids, inflammation, and fibrosis. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology* 2016 doi: 10.1096/fj.201600646R [published Online First: 2016/07/21]
20. Zgair A, Lee JB, Wong JCM, et al. Oral administration of cannabis with lipids leads to high levels of cannabinoids in the intestinal lymphatic system and prominent immunomodulation. *Sci Rep* 2017;7(1):14542. doi: 10.1038/s41598-017-15026-z
21. Costiniuk CT, Brunet L, Rollet-Kurhajec KC, et al. Tobacco Smoking Is Not Associated With Accelerated Liver Disease in Human Immunodeficiency Virus-Hepatitis C Coinfection: A Longitudinal Cohort Analysis. *Open forum infectious diseases* 2016;3(2):ofw050. doi: 10.1093/ofid/ofw050 [published Online First: 2016/04/06]
22. Abrams DI, Hilton JF, Leiser RJ, et al. Short-term effects of cannabinoids in patients with HIV-1 infection: a randomized, placebo-controlled clinical trial. *Annals of internal medicine* 2003;139(4):258-66.
23. Lee MH, Hancox RJ. Effects of smoking cannabis on lung function. *Expert review of respiratory medicine* 2011;5(4):537-46; quiz 47. doi: 10.1586/ers.11.40 [published Online First: 2011/08/24]
24. Tahamtan A, Tavakoli-Yaraki M, Rygiel TP, et al. Effects of cannabinoids and their receptors on viral infections. *Journal of medical virology* 2016;88(1):1-12. doi: 10.1002/jmv.24292
25. Smith TH, Sim-Selley LJ, Selley DE. Cannabinoid CB1 receptor-interacting proteins: novel targets for central nervous system drug discovery? *Br J Pharmacol* 2010;160(3):454-66. doi: 10.1111/j.1476-5381.2010.00777.x

26. DiPatrizio NV. Endocannabinoids in the Gut. *Cannabis Cannabinoid Res* 2016;1(1):67-77. doi: 10.1089/can.2016.0001

27. Muccioli GG, Naslain D, Backhed F, et al. The endocannabinoid system links gut microbiota to adipogenesis. *Mol Syst Biol* 2010;6:392. doi: 10.1038/msb.2010.46

28. Rietschel ET, Kirikae T, Schade FU, et al. Bacterial endotoxin: molecular relationships of structure to activity and function. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology* 1994;8(2):217-25.

29. Yao B, Mackie K. Endocannabinoid receptor pharmacology. *Curr Top Behav Neurosci* 2009;1:37-63. doi: 10.1007/978-3-540-88955-7_2

30. Eisenstein TK, Meissler JJ. Effects of Cannabinoids on T-cell Function and Resistance to Infection. *Journal of neuroimmune pharmacology : the official journal of the Society on NeuroImmune Pharmacology* 2015;10(2):204-16. doi: 10.1007/s11481-015-9603-3

31. Hanus L, Breuer A, Tchilibon S, et al. HU-308: a specific agonist for CB(2), a peripheral cannabinoid receptor. *Proceedings of the National Academy of Sciences of the United States of America* 1999;96(25):14228-33.

32. Russo EB. Cannabidiol Claims and Misconceptions. *Trends Pharmacol Sci* 2017 doi: 10.1016/j.tips.2016.12.004

33. Laprairie RB, Bagher AM, Kelly ME, et al. Cannabidiol is a negative allosteric modulator of the cannabinoid CB1 receptor. *Br J Pharmacol* 2015;172(20):4790-805. doi: 10.1111/bph.13250

34. Ware MA, Wang T, Shapiro S, et al. Cannabis for the Management of Pain: Assessment of Safety Study (COMPASS). *The journal of pain : official journal of the American Pain Society* 2015;16(12):1233-42. doi: 10.1016/j.jpain.2015.07.014

35. Abrams DI, Jay CA, Shade SB, et al. Cannabis in painful HIV-associated sensory neuropathy: a randomized placebo-controlled trial. *Neurology* 2007;68(7):515-21. doi: 10.1212/01.wnl.0000253187.66183.9c

36. Wilsey B, Marcotte T, Tsodikov A, et al. A randomized, placebo-controlled, crossover trial of cannabis cigarettes in neuropathic pain. *The journal of pain : official journal of the American Pain Society* 2008;9(6):506-21. doi: 10.1016/j.jpain.2007.12.010

37. Steffens S, Veillard NR, Arnaud C, et al. Low dose oral cannabinoid therapy reduces progression of atherosclerosis in mice. *Nature* 2005;434(7034):782-6. doi: 10.1038/nature03389

38. Franz CA, Frishman WH. Marijuana Use and Cardiovascular Disease. *Cardiol Rev* 2016;24(4):158-62. doi: 10.1097/CRD.000000000000103

39. Mallat A, Lotersztajn S. Endocannabinoids and liver disease. I. Endocannabinoids and their receptors in the liver. *Am J Physiol Gastrointest Liver Physiol* 2008;294(1):G9-G12. doi: 10.1152/ajpgi.00467.2007

40. Mallat A, Teixeira-Clerc F, Deveaux V, et al. The endocannabinoid system as a key mediator during liver diseases: new insights and therapeutic openings. *Br J Pharmacol* 2011;163(7):1432-40. doi: 10.1111/j.1476-5381.2011.01397.x

Protected by copyright, including for uses related to text and data mining, AI training, and similar technologies.
Downloaded from <http://bmjopen.bmj.com/> on June 13, 2025 at Agence Bibliographique de l'Enseignement Supérieur (ABES).

41. Teixeira-Clerc F, Belot MP, Manin S, et al. Beneficial paracrine effects of cannabinoid receptor 2 on liver injury and regeneration. *Hepatology* 2010;52(3):1046-59. doi: 10.1002/hep.23779
42. Munoz-Luque J, Ros J, Fernandez-Varo G, et al. Regression of fibrosis after chronic stimulation of cannabinoid CB2 receptor in cirrhotic rats. *The Journal of pharmacology and experimental therapeutics* 2008;324(2):475-83. doi: 10.1124/jpet.107.131896
43. Tam J, Vemuri VK, Liu J, et al. Peripheral CB1 cannabinoid receptor blockade improves cardiometabolic risk in mouse models of obesity. *The Journal of clinical investigation* 2010;120(8):2953-66. doi: 10.1172/JCI42551
44. Cluny NL, Vemuri VK, Chambers AP, et al. A novel peripherally restricted cannabinoid receptor antagonist, AM6545, reduces food intake and body weight, but does not cause malaise, in rodents. *Br J Pharmacol* 2010;161(3):629-42. doi: 10.1111/j.1476-5381.2010.00908.x
45. Costantino CM, Gupta A, Yewdall AW, et al. Cannabinoid receptor 2-mediated attenuation of CXCR4-tropic HIV infection in primary CD4+ T cells. *PloS one* 2012;7(3):e33961. doi: 10.1371/journal.pone.0033961
46. Ramirez SH, Reichenbach NL, Fan S, et al. Attenuation of HIV-1 replication in macrophages by cannabinoid receptor 2 agonists. *Journal of leukocyte biology* 2013;93(5):801-10. doi: 10.1189/jlb.1012523
47. Xu H, Cheng CL, Chen M, et al. Anti-inflammatory property of the cannabinoid receptor-2-selective agonist JWH-133 in a rodent model of autoimmune uveoretinitis. *Journal of leukocyte biology* 2007;82(3):532-41. doi: 10.1189/jlb.0307159
48. Zhang M, Adler MW, Abood ME, et al. CB2 receptor activation attenuates microcirculatory dysfunction during cerebral ischemic/reperfusion injury. *Microvasc Res* 2009;78(1):86-94. doi: 10.1016/j.mvr.2009.03.005
49. Condie R, Herring A, Koh WS, et al. Cannabinoid inhibition of adenylate cyclase-mediated signal transduction and interleukin 2 (IL-2) expression in the murine T-cell line, EL4.IL-2. *The Journal of biological chemistry* 1996;271(22):13175-83.
50. Massi P, Sacerdote P, Ponti W, et al. Immune function alterations in mice tolerant to delta9-tetrahydrocannabinol: functional and biochemical parameters. *J Neuroimmunol* 1998;92(1-2):60-6.
51. Molina PE, Amedee AM, LeCapitaine NJ, et al. Modulation of gut-specific mechanisms by chronic delta(9)-tetrahydrocannabinol administration in male rhesus macaques infected with simian immunodeficiency virus: a systems biology analysis. *AIDS research and human retroviruses* 2014;30(6):567-78. doi: 10.1089/AID.2013.0182
52. Lutge EE, Gray A, Siegfried N. The medical use of cannabis for reducing morbidity and mortality in patients with HIV/AIDS. *Cochrane Database Syst Rev* 2013(4):CD005175. doi: 10.1002/14651858.CD005175.pub3
53. Manuzak JA, Gott TM, Kirkwood JS, et al. Heavy Cannabis Use Associated With Reduction in Activated and Inflammatory Immune Cell Frequencies in Antiretroviral Therapy-Treated Human Immunodeficiency Virus-Infected Individuals. *Clinical infectious diseases : an official publication of the Infectious Diseases Society of America* 2018 doi: 10.1093/cid/cix1116

54. Rizzo MD, Crawford RB, Henriquez JE, et al. HIV-infected cannabis users have lower circulating CD16+ monocytes and IFN-gamma-inducible protein 10 levels compared with nonusing HIV patients. *Aids* 2018;32(4):419-29. doi: 10.1097/QAD.0000000000001704

55. Burstein S. Cannabidiol (CBD) and its analogs: a review of their effects on inflammation. *Bioorg Med Chem* 2015;23(7):1377-85. doi: 10.1016/j.bmc.2015.01.059

56. Bergamaschi MM, Queiroz RH, Zuardi AW, et al. Safety and side effects of cannabidiol, a Cannabis sativa constituent. *Curr Drug Saf* 2011;6(4):237-49.

57. Ribeiro A, Ferraz-de-Paula V, Pinheiro ML, et al. Cannabidiol, a non-psychotropic plant-derived cannabinoid, decreases inflammation in a murine model of acute lung injury: role for the adenosine A(2A) receptor. *Eur J Pharmacol* 2012;678(1-3):78-85. doi: 10.1016/j.ejphar.2011.12.043

58. Burstein SH, Zurier RB. Cannabinoids, endocannabinoids, and related analogs in inflammation. *AAPS J* 2009;11(1):109-19. doi: 10.1208/s12248-009-9084-5

59. Iuvone T, Esposito G, De Filippis D, et al. Cannabidiol: a promising drug for neurodegenerative disorders? *CNS Neurosci Ther* 2009;15(1):65-75. doi: 10.1111/j.1755-5949.2008.00065.x

60. Devinsky O, Patel AD, Cross JH, et al. Effect of Cannabidiol on Drop Seizures in the Lennox-Gastaut Syndrome. *The New England journal of medicine* 2018;378(20):1888-97. doi: 10.1056/NEJMoa1714631

61. Sachse-Seeboth C, Pfeil J, Sehr D, et al. Interindividual variation in the pharmacokinetics of Delta9-tetrahydrocannabinol as related to genetic polymorphisms in CYP2C9. *Clinical pharmacology and therapeutics* 2009;85(3):273-6. doi: 10.1038/clpt.2008.213

62. Jenabian MA, Seddiki N, Yatim A, et al. Regulatory T cells negatively affect IL-2 production of effector T cells through CD39/adenosine pathway in HIV infection. *PLoS pathogens* 2013;9(4):e1003319. doi: 10.1371/journal.ppat.1003319 [published Online First: 2013/05/10]

63. Nikolova M, Carriere M, Jenabian MA, et al. CD39/adenosine pathway is involved in AIDS progression. *PLoS pathogens* 2011;7(7):e1002110. doi: 10.1371/journal.ppat.1002110

64. Vandergeeten C, Fromentin R, Merlini E, et al. Cross-Clade Ultrasensitive PCR-Based Assays To Measure HIV Persistence in Large-Cohort Studies. *Journal of virology* 2014;88(21):12385-96. doi: 10.1128/jvi.00609-14 [published Online First: 2014/08/15]

65. Procopio FA, Fromentin R, Kulpa DA, et al. A Novel Assay to Measure the Magnitude of the Inducible Viral Reservoir in HIV-infected Individuals. *EBioMedicine* 2015;2(8):872-81. doi: 10.1016/j.ebiom.2015.06.019 [published Online First: 2015/10/02]

66. Sharkey M, Triques K, Kuritzkes DR, et al. In vivo evidence for instability of episomal human immunodeficiency virus type 1 cDNA. *Journal of virology* 2005;79(8):5203-10. doi: 10.1128/jvi.79.8.5203-5210.2005 [published Online First: 2005/03/30]

67. Buzon MJ, Massanella M, Llibre JM, et al. HIV-1 replication and immune dynamics are affected by raltegravir intensification of HAART-suppressed subjects. *Nature*

- medicine* 2010;16(4):460-5. doi: 10.1038/nm.2111 [published Online First: 2010/03/17]
68. Nowak P, Trosheid M, Avershina E, et al. Gut microbiota diversity predicts immune status in HIV-1 infection. *Aids* 2015;29(18):2409-18. doi: 10.1097/QAD.0000000000000869
 69. Hoel H, Hove-Skovsgaard M, Hov JR, et al. Impact of HIV and Type 2 diabetes on Gut Microbiota Diversity, Tryptophan Catabolism and Endothelial Dysfunction. *Sci Rep* 2018;8(1):6725. doi: 10.1038/s41598-018-25168-3
 70. Lebouche B, Jenabian MA, Singer J, et al. The role of extended-release niacin on immune activation and neurocognition in HIV-infected patients treated with antiretroviral therapy - CTN PT006: study protocol for a randomized controlled trial. *Trials* 2014;15:390. doi: 10.1186/1745-6215-15-390
 71. Hill AL, Rosenbloom DI, Goldstein E, et al. Real-Time Predictions of Reservoir Size and Rebound Time during Antiretroviral Therapy Interruption Trials for HIV. *PLoS pathogens* 2016;12(4):e1005535. doi: 10.1371/journal.ppat.1005535
 72. 2018 C-PaFTAahwc-soAM.
 73. Chan AW, Tetzlaff JM, Altman DG, et al. SPIRIT 2013 statement: defining standard protocol items for clinical trials. *Annals of internal medicine* 2013;158(3):200-7. doi: 10.7326/0003-4819-158-3-201302050-00583
 74. [http://www.icmje.org/recommendations/archives/2008_urm.pdf] ICoMJEURfMStBJWaEfBP.
 75. Mechoulam R, Hanus LO, Pertwee R, et al. Early phytocannabinoid chemistry to endocannabinoids and beyond. *Nat Rev Neurosci* 2014;15(11):757-64. doi: 10.1038/nrn3811
 76. Portenoy RK, Ganae-Motan ED, Allende S, et al. Nabiximols for opioid-treated cancer patients with poorly-controlled chronic pain: a randomized, placebo-controlled, graded-dose trial. *The journal of pain : official journal of the American Pain Society* 2012;13(5):438-49. doi: 10.1016/j.jpain.2012.01.003
 77. Serpell M, Ratcliffe S, Hovorka J, et al. A double-blind, randomized, placebo-controlled, parallel group study of THC/CBD spray in peripheral neuropathic pain treatment. *Eur J Pain* 2014;18(7):999-1012. doi: 10.1002/j.1532-2149.2013.00445.x
 78. Pertwee RG. Emerging strategies for exploiting cannabinoid receptor agonists as medicines. *Br J Pharmacol* 2009;156(3):397-411. doi: 10.1111/j.1476-5381.2008.00048.x
 79. Tramer MR, Carroll D, Campbell FA, et al. Cannabinoids for control of chemotherapy induced nausea and vomiting: quantitative systematic review. *BMJ (Clinical research ed)* 2001;323(7303):16-21.
 80. Curran HV, Freeman TP, Mokrysz C, et al. Keep off the grass? Cannabis, cognition and addiction. *Nat Rev Neurosci* 2016;17(5):293-306. doi: 10.1038/nrn.2016.28
 81. Notcutt W, Langford R, Davies P, et al. A placebo-controlled, parallel-group, randomized withdrawal study of subjects with symptoms of spasticity due to multiple sclerosis who are receiving long-term Sativex(R) (nabiximols). *Mult Scler* 2012;18(2):219-28. doi: 10.1177/1352458511419700

82. Wade DT, Makela PM, House H, et al. Long-term use of a cannabis-based medicine in the treatment of spasticity and other symptoms in multiple sclerosis. *Mult Scler* 2006;12(5):639-45. doi: 10.1177/1352458505070618

83. Robson P. Abuse potential and psychoactive effects of delta-9-tetrahydrocannabinol and cannabidiol oromucosal spray (Sativex), a new cannabinoid medicine. *Expert Opin Drug Saf* 2011;10(5):675-85. doi: 10.1517/14740338.2011.575778

84. 2015 GPSPMM.

85. Zuardi AW, Crippa JA, Hallak JE, et al. Cannabidiol for the treatment of psychosis in Parkinson's disease. *J Psychopharmacol* 2009;23(8):979-83. doi: 10.1177/0269881108096519

86. 2017 AIMPLA.

87. Herkenham M, Lynn AB, Little MD, et al. Cannabinoid receptor localization in brain. *Proceedings of the National Academy of Sciences of the United States of America* 1990;87(5):1932-6.

88. Boulassel MR, Mercier F, Gilmore N, et al. Immunophenotypic patterns of CD8+ T cell subsets expressing CD8alphaalpha and IL-7Ralpha in viremic, aviremic and slow progressor HIV-1-infected subjects. *Clinical immunology* 2007;124(2):149-57. doi: 10.1016/j.clim.2007.05.005

89. Mercier F, Boulassel MR, Yassine-Diab B, et al. Persistent human immunodeficiency virus-1 antigenaemia affects the expression of interleukin-7Ralpha on central and effector memory CD4+ and CD8+ T cell subsets. *Clinical and experimental immunology* 2008;152(1):72-80. doi: 10.1111/j.1365-2249.2008.03610.x

90. Deeks SG, Phillips AN. HIV infection, antiretroviral treatment, ageing, and non-AIDS related morbidity. *BMJ (Clinical research ed)* 2009;338:a3172. doi: 10.1136/bmj.a3172 [published Online First: 2009/01/28]

91. Sparacino CM, Hyldborg PA, Hughes TJ. Chemical and biological analysis of marijuana smoke condensate. *NIDA research monograph* 1990;99:121-40. [published Online First: 1990/01/01]

Authors' contributions

CTC conceived and designed the study, drafted the grant and the protocol manuscript, will organize and supervise trial implementation and will be responsible for trial management. She will also be responsible for trial managements, staff training and supervision. MW and MAJ contributed to study design and participated in grant writing. BL, JPR, JC, MJB and MK will participate in study implementation. MJB provided input on questionnaires while CTC and MAJ provided immunological expertise and NC provided expertise related to the

HIV reservoir. SM and EM contributed to study design. CTC, MAJ and NC designed the experiments. JS contributed to the statistical analysis plan. CTC, MAJ and NC designed the experiments. All authors participated in refinement of the study methods, critical reviewed the manuscript drafts and approved the final manuscript. The CTN provides regulatory support. CTC and BL are Fonds de recherche du Québec-Santé (FRQ-S) chercheur-boursier-clinicien Junior 1. BL holds a rategy for Patient-Oriented Research (SPOR) Mentorship Chair in Innovative Clinical Trials. JPR holds the Louis Lowenstein Chair in Hematology and Oncology at McGill University. MAJ is holder of a Tier 2 Canada Research Chair in immunovirology.

Acknowledgements

This clinical trial was reviewed by both the CTN Scientific Advisory Committee and the Community Advisory Committee and is supported by a competitive grant (CTN PT028). We also wish to acknowledge Ms. Dana Nohyek, Ms. Judy Needham, Ms. Jacqueline Sas and the Community Advisory Committee of the CTN for ongoing support. In addition, we thank Ms Hansi Peiris and Jonathan Roger of the CVIS for logistical and administrative assistance. Furthermore, we wish to acknowledge Tilray and notably Philippe Lucas and Catherine Jacobson for interest and support of this study through the provision of cannabinoid capsules and for assistance with the application to Health Canada.

Funding statement: This work is supported by the CIHR Canadian HIV Trials Network.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Competing interests: Tilray Inc. is supplying the study medications free of charge. All elements of the study are being undertaken independently of Tilray Inc. The authors declare there are no conflicts of interests.

Data sharing: There are no data yet to share.

For peer review only



SPIRIT 2013 Checklist: Recommended items to address in a clinical trial protocol and related documents*

Section/item	Item No	Description	Addressed on page number
Administrative information			
Title	1	Descriptive title identifying the study design, population, interventions, and, if applicable, trial acronym	_____1_____
Trial registration	2a	Trial identifier and registry name. If not yet registered, name of intended registry	Abstract p 4_____
	2b	All items from the World Health Organization Trial Registration Data Set	_____p.3-4
Protocol version	3	Date and version identifier	Abstract /cover letter_
Funding	4	Sources and types of financial, material, and other support	_____50_____
Roles and responsibilities	5a	Names, affiliations, and roles of protocol contributors	_____1, 49,50_____
	5b	Name and contact information for the trial sponsor	_____1_____
	5c	Role of study sponsor and funders, if any, in study design; collection, management, analysis, and interpretation of data; writing of the report; and the decision to submit the report for publication, including whether they will have ultimate authority over any of these activities	_____49, 50_____

1				
2				
3		5d	Composition, roles, and responsibilities of the coordinating centre, steering committee, endpoint	_____4,24, 51,
4			adjudication committee, data management team, and other individuals or groups overseeing the trial, if	cover letter
5			applicable (see Item 21a for data monitoring committee)	
6				
7				
8				
9				
10				
11	Introduction			
12				
13	Background and	6a	Description of research question and justification for undertaking the trial, including summary of relevant	_13, 14_____
14	rationale		studies (published and unpublished) examining benefits and harms for each intervention	
15				
16		6b	Explanation for choice of comparators	18, 19_____
17	Objectives	7	Specific objectives or hypotheses	23,24_____
18				
19	Trial design	8	Description of trial design including type of trial (eg, parallel group, crossover, factorial, single group),	
20			allocation ratio, and framework (eg, superiority, equivalence, noninferiority, exploratory)	Abstract, 18, 19_
21				
22				
23	Methods: Participants, interventions, and outcomes			
24				
25	Study setting	9	Description of study settings (eg, community clinic, academic hospital) and list of countries where data will	15__
26			be collected. Reference to where list of study sites can be obtained	
27				
28	Eligibility criteria	10	Inclusion and exclusion criteria for participants. If applicable, eligibility criteria for study centres and	16,17_
29			individuals who will perform the interventions (eg, surgeons, psychotherapists)	
30				
31	Interventions	11a	Interventions for each group with sufficient detail to allow replication, including how and when they will be	18
32			administered	
33				
34		11b	Criteria for discontinuing or modifying allocated interventions for a given trial participant (eg, drug dose	25,26
35			change in response to harms, participant request, or improving/worsening disease)	
36				
37		11c	Strategies to improve adherence to intervention protocols, and any procedures for monitoring adherence	25,26
38			(eg, drug tablet return, laboratory tests)	
39				
40		11d	Relevant concomitant care and interventions that are permitted or prohibited during the trial	25,26
41				
42				

Outcomes	12	Primary, secondary, and other outcomes, including the specific measurement variable (eg, systolic blood pressure), analysis metric (eg, change from baseline, final value, time to event), method of aggregation (eg, median, proportion), and time point for each outcome. Explanation of the clinical relevance of chosen efficacy and harm outcomes is strongly recommended	24,25
Participant timeline	13	Time schedule of enrolment, interventions (including any run-ins and washouts), assessments, and visits for participants. A schematic diagram is highly recommended (see Figure)	Table 3
Sample size	14	Estimated number of participants needed to achieve study objectives and how it was determined, including clinical and statistical assumptions supporting any sample size calculations	29
Recruitment	15	Strategies for achieving adequate participant enrolment to reach target sample size	Cover letter, already advertised by Canadian HIV Trials Network newsletter and website, meetings involving community

Methods: Assignment of interventions (for controlled trials)

Allocation:

Sequence generation	16a	Method of generating the allocation sequence (eg, computer-generated random numbers), and list of any factors for stratification. To reduce predictability of a random sequence, details of any planned restriction (eg, blocking) should be provided in a separate document that is unavailable to those who enrol participants or assign interventions	19,20
Allocation concealment mechanism	16b	Mechanism of implementing the allocation sequence (eg, central telephone; sequentially numbered, opaque, sealed envelopes), describing any steps to conceal the sequence until interventions are assigned	19, 20
Implementation	16c	Who will generate the allocation sequence, who will enrol participants, and who will assign participants to interventions	19,20

1				
2				
3	Blinding (masking)	17a	Who will be blinded after assignment to interventions (eg, trial participants, care providers, outcome assessors, data analysts), and how	20,21 (non-blind)
4				
5		17b	If blinded, circumstances under which unblinding is permissible, and procedure for revealing a participant's allocated intervention during the trial	20, 21 (non-blind)
6				
7				
8				
9	Methods: Data collection, management, and analysis			
10				
11	Data collection methods	18a	Plans for assessment and collection of outcome, baseline, and other trial data, including any related processes to promote data quality (eg, duplicate measurements, training of assessors) and a description of study instruments (eg, questionnaires, laboratory tests) along with their reliability and validity, if known. Reference to where data collection forms can be found, if not in the protocol	20,21
12				
13		18b	Plans to promote participant retention and complete follow-up, including list of any outcome data to be collected for participants who discontinue or deviate from intervention protocols	22
14				
15	Data management	19	Plans for data entry, coding, security, and storage, including any related processes to promote data quality (eg, double data entry; range checks for data values). Reference to where details of data management procedures can be found, if not in the protocol	22, 31, 32
16				
17				
18				
19				
20				
21	Statistical methods	20a	Statistical methods for analysing primary and secondary outcomes. Reference to where other details of the statistical analysis plan can be found, if not in the protocol	29,30,31
22				
23		20b	Methods for any additional analyses (eg, subgroup and adjusted analyses)	29,30,31
24				
25		20c	Definition of analysis population relating to protocol non-adherence (eg, as randomised analysis), and any statistical methods to handle missing data (eg, multiple imputation)	N/A
26				
27				
28				
29				
30				
31				
32	Methods: Monitoring			
33				
34	Data monitoring	21a	Composition of data monitoring committee (DMC); summary of its role and reporting structure; statement of whether it is independent from the sponsor and competing interests; and reference to where further details about its charter can be found, if not in the protocol. Alternatively, an explanation of why a DMC is not needed	Cover letter, 25
35				
36				
37				
38				
39				
40				
41				
42				
43				
44				
45				
46				
47				

	21b	Description of any interim analyses and stopping guidelines, including who will have access to these interim results and make the final decision to terminate the trial	25, Health Canada, Canadian HIV Trials Network
Harms	22	Plans for collecting, assessing, reporting, and managing solicited and spontaneously reported adverse events and other unintended effects of trial interventions or trial conduct	25, 33, Health Canada, CTN
Auditing	23	Frequency and procedures for auditing trial conduct, if any, and whether the process will be independent from investigators and the sponsor	33
Ethics and dissemination			
Research ethics approval	24	Plans for seeking research ethics committee/institutional review board (REC/IRB) approval	32
Protocol amendments	25	Plans for communicating important protocol modifications (eg, changes to eligibility criteria, outcomes, analyses) to relevant parties (eg, investigators, REC/IRBs, trial participants, trial registries, journals, regulators)	32
Consent or assent	26a	Who will obtain informed consent or assent from potential trial participants or authorised surrogates, and how (see Item 32)	32
	26b	Additional consent provisions for collection and use of participant data and biological specimens in ancillary studies, if applicable	21 (study coordinator)
Confidentiality	27	How personal information about potential and enrolled participants will be collected, shared, and maintained in order to protect confidentiality before, during, and after the trial	N/A at this time
Declaration of interests	28	Financial and other competing interests for principal investigators for the overall trial and each study site	31, 32, 50
Access to data	29	Statement of who will have access to the final trial dataset, and disclosure of contractual agreements that limit such access for investigators	31, 32
Ancillary and post-trial care	30	Provisions, if any, for ancillary and post-trial care, and for compensation to those who suffer harm from trial participation	25

1				
2				
3	Dissemination policy	31a	Plans for investigators and sponsor to communicate trial results to participants, healthcare professionals, the public, and other relevant groups (eg, via publication, reporting in results databases, or other data sharing arrangements), including any publication restrictions	32
4				
5				
6				
7		31b	Authorship eligibility guidelines and any intended use of professional writers	32,33
8				
9		31c	Plans, if any, for granting public access to the full protocol, participant-level dataset, and statistical code	Not determined at this time
10				
11				
12	Appendices			
13				
14	Informed consent materials	32	Model consent form and other related documentation given to participants and authorised surrogates	__Available upon request__
15				
16				
17	Biological specimens	33	Plans for collection, laboratory evaluation, and storage of biological specimens for genetic or molecular analysis in the current trial and for future use in ancillary studies, if applicable	N/A (no plans for additional studies at this time)
18				
19				
20				

21 *It is strongly recommended that this checklist be read in conjunction with the SPIRIT 2013 Explanation & Elaboration for important clarification on the items.
22 Amendments to the protocol should be tracked and dated. The SPIRIT checklist is copyrighted by the SPIRIT Group under the Creative Commons
23 "[Attribution-NonCommercial-NoDerivs 3.0 Unported](#)" license.
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44

BMJ Open

Oral cannabinoids in people living with HIV on effective antiretroviral therapy—CTN PT028: Study protocol for a pilot randomized trial to assess safety, tolerability and effect on immune activation

Journal:	<i>BMJ Open</i>
Manuscript ID	bmjopen-2018-024793.R1
Article Type:	Protocol
Date Submitted by the Author:	17-Oct-2018
Complete List of Authors:	Costiniuk, C; Chronic Viral Illness Service, McGill University Health Centre; Research Institute, McGill University Health Centre Saneei, Zahra; Chronic Viral Illness Service, McGill University Health Centre Routy, JP; Chronic Viral Illness Service, McGill University Health Centre; Research Institute of the McGill University Health Centre Margolese, Shari; Canadian Institutes of Health Research Canadian HIV Trials Network Mandarino, Enrico; Canadian Institutes of Health Research Canadian HIV Trials Network; WILLL Cannabis Group Singer, Joel; Canadian Institutes of Health Research Canadian HIV Trials Network; School of Population and Public Health, University of British Columbia, Vancouver, BC, Canada Lebouché, Bertrand; Chronic Viral Illness Service, McGill University Health Centre; Department of Family Medicine, McGill University Cox, Joseph; Chronic Viral Illness Service, McGill University Health Centre; Department of Family Medicine, McGill University Szabo, Jason; Chronic Viral Illness Service, McGill University Health Centre; Department of Family Medicine, McGill University Brouillette, Marie-Josée ; Chronic Viral Illness Service, McGill University Health Centre; Department of Psychiatry, McGill University Health Centre Klein, Marina; Chronic Viral Illness Service, McGill University Health Centre; Research Institute, McGill University Health Centre Chomont, Nicolas; Centre de Recherche du CHUM and Department of microbiology, infectiology and immunology, Université de Montréal Jenabian, Mohammad-Ali; Université du Québec à Montréal, Biological Sciences
Primary Subject Heading:	HIV/AIDS
Secondary Subject Heading:	Immunology (including allergy), Complementary medicine, Patient-centred medicine
Keywords:	cannabinoids, cannabis, safety and tolerability, inflammation, immune activation, HIV reservoir



Oral cannabinoids in people living with HIV on effective antiretroviral therapy—CTN

PT028: Study protocol for a pilot randomized trial to assess safety, tolerability and effect on immune activation

Cecilia T. Costiniuk^{1,2}, Zahra Saneel¹, Jean-Pierre Routy^{1,2}, Shari Margolese³,

Enrico Mandarino⁴, Joel Singer⁵, Bertrand Lebouché^{1,2,6},

Joseph Cox^{1,2,6}, Jason Szabo^{1,2,6}, Marie-Josée Brouillette^{1,2,7}, Marina B. Klein^{1,2},

Nicolas Chomont⁸, Mohammad-Ali Jenabian⁹

¹Chronic Viral Illness Service, McGill University Health Centre, Montreal, QC, Canada

²Research Institute of McGill University Health Centre, Montreal, QC, Canada

³CIHR Canadian HIV Trials Network

⁴CIHR Canadian HIV Trials Network; WILL Cannabis Group

⁵Canadian Institute of Health Research Canadian HIV Trials Network, Vancouver, BC, Canada;

School of Population and Public Health, University of British Columbia, Vancouver, BC,

Canada

⁶Department of Family Medicine, McGill University Health Centre, Montreal, QC, Canada

⁷Department of Psychiatry, McGill University Health Centre, Montreal, QC, Canada

⁸Centre de Recherche du CHUM and Department of microbiology, infectiology and immunology, Université de Montréal, Montreal, QC, Canada

⁹Department of Biological Sciences and BioMed Research Centre, University of Quebec at Montreal (UQAM), Montreal, Quebec, Canada

Key words: cannabinoids; cannabis; HIV; inflammation; immune activation; HIV reservoir, microbiome

Short title: Cannabinoids and HIV

Word count: 7,640

Correspondence: Cecilia Costiniuk, MD, MSc, Division of Infectious Diseases and Chronic Viral Illness Service, McGill University Health Centre, 1001 boulevard Décarie, Montreal (QC) Canada, H4A 3J1 Tel: 1 (514) 843-2090; Fax: 1 (514) 843 2092; Email: cecilia.costiniuk@mcgill.ca

Abstract:

Introduction: Despite antiretroviral therapy(ART), people living with HIV have higher rates of non-infectious chronic diseases. These conditions are driven by relatively high levels of inflammation persisting on ART compared to uninfected individuals. Chronic inflammation also contributes to HIV persistence during ART. Cannabis when taken orally may represent a way to reduce inflammation and strengthen immune responses. Before planning large interventional studies, it is important to ensure that cannabis taken orally is safe and well-tolerated in people living with HIV. We propose to conduct a pilot randomized trial to examine the safety and tolerability of cannabis oils containing tetrahydrocannabinol(THC) and cannabidiol(CBD) consumed orally in people living with HIV. We will also measure inflammatory markers, markers of HIV persistence in peripheral blood cells and changes in the gastrointestinal microbiome.

Methods and Analysis: Twenty-six people living with HIV having undetectable viral load for at least three years will be randomized to receive TN-TC11LM(THC:CBD in 1:1 ratio) or TN-TC19LM(THC:CBD in 1:9 ratio) capsules daily for 12 weeks. Safety and tolerability of these capsules will be assessed through haematological, hepatic and renal blood tests, face-to-face interviews and questionnaires. Proportions of participants without any signs of significant

toxicity(Grades 0-2 scores on the WHO toxicity scale) and who complete the study, as well as scores on quality of life and mood will be examined using descriptive statistics. The effects on inflammatory markers, markers of peripheral blood reservoir size and effect on the composition of the gastrointestinal microbiome will be assessed before and after study completion.

Ethics and Dissemination: This study has been approved by the Research Institute of the McGill University Health Centre. A Data Safety Monitor will review safety information at regular intervals. The final manuscript will be submitted to an open-access journal within 6 months of study completion. This trial is registered with [clinicaltrials.gov](https://clinicaltrials.gov/ct2/show/study/NCT03550352)([NCT03550352](https://clinicaltrials.gov/ct2/show/study/NCT03550352)).

Strengths and limitations of this study

- Randomized clinical trial design involving oral consumption of capsules containing 2 different ratios of cannabinoids
- The capsules used will contain both $\Delta 9$ -tetrahydrocannabinol (THC) and cannabidiol (CBD), thus improving tolerability
- The use of oral capsules containing precise amounts of cannabinoids, rather than smoked cannabis, ensures more predictable dosage administration and avoids the harmful pulmonary effects associated with smoking
- Effects of the intervention on quality of life, cognition and mood in addition to biological outcomes are being examined
- The number of participants should provide insight into the degree of variability for continuous outcomes and should guide future sample size calculations for larger studies

1

2

3

4 **Background**

5

6

7

8 Despite antiretroviral therapy (ART), persistent immune activation is associated with

9

10 increased risk of non-opportunistic complications in people living with HIV such as

11

12 cardiovascular, pulmonary, renal and hepatic events^{1 2}. HIV pathogenesis and persistence appear

13

14 to be related to chronic inflammation and immune activation³, driven by microbial translocation

15

16 of bacterial products across the gut mucosa⁴⁻⁶. Even when ART is initiated in the primary or

17

18 early phase of HIV, gut integrity is not fully restored⁷. Furthermore, there is recent evidence to

19

20 suggest that inflammatory features of the enteric microbiota, and not just increased permeability

21

22 alone, is driving chronic inflammation in people living with HIV. Indeed, some studies have

23

24 shown correlations between specific enteric bacteria and immune activation markers in gut and

25

26 blood^{8 9}. Persistent immune activation may also contribute to the persistence of HIV during

27

28 ART¹⁰⁻¹⁵. HIV reservoirs are the reason why HIV remains an incurable infection. Although

29

30 HIV may also persist in myeloid cells, CD4+ T cells are the best-characterized and the most

31

32 abundant reservoirs¹⁶⁻¹⁸.

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47 Attenuation of immune activation and levels of inflammation may portend several

48

49 therapeutic benefits to people living with HIV and novel strategies are needed to achieve this

50

51 goal. Having both anti-inflammatory and anti-fibrotic properties¹⁹, cannabis may represent a

52

53

54

55

56

57

58

59

60

feasible method to reduce immune activation and enhance immune profile. When consumed orally, notably soluble in oil, it may also temper inflammation at the gastrointestinal mucosa²⁰. This, in turn, may hasten the progression of non-opportunistic complications associated with HIV. Furthermore, cannabis use, mainly through smoking, is already extremely common amongst PLWHIV. A study conducted by our team involving 1072 participants in the Canadian Co-infection Cohort Study (CTN 222), a multi-centered longitudinal study of HIV-Hepatitis C co-infected individuals, revealed that 53% of individuals had smoked marijuana in the past 6 months²¹. A clearer understanding of the safety, tolerability and feasibility of using cannabis in a clinical trial is an important first step before designing interventional studies to ameliorate specific conditions of importance to people living with HIV. Furthermore, as cannabis use will be legalized in Canada July 2018, understanding its safety profile is very important.

Pharmacology and medical properties of cannabis

Cannabinoids, found in the hemp plant *Cannabis sativa*, have been recognized for centuries for their analgesic, anticonvulsant, bronchodilatory, sedative, hypnotic and antispasmodic properties^{22 23}. Their biological activity is conferred by cannabinoid receptors CB1 and CB2 through activation of heterodimeric G-proteins that function as signaling and

regulatory proteins to operate or modulate intracellular signaling pathways^{24 25}. While CB1 receptors are expressed predominantly in the central nervous system, they are also found in the lung, liver and kidneys. The endocannabinoid system also plays a key role in the gastrointestinal tract's neural and molecular control mechanisms. Indeed, the endocannabinoid system plays a role in normal physiological functions of the gastrointestinal tract including motility, gut-brain-mediated fat intake, hunger signaling, inflammation and gut permeability²⁶. Furthermore, there is some evidence for and interactions between the endocannabinoid system and the gut microbiota²⁶. In lean mice who received 4 weeks of a cannabinoid-receptor agonist HU-210, plasma lipopolysaccharides (LPS) were significantly increased^{26 27}. When obese mice with disrupted gastrointestinal musoca were treated with rimonabant, an inverse CB1 agonist, for nearly 2 weeks, reductions in plasma LPS were observed^{26 28}. Furthermore, improvements in localization of tight junction proteins, occluding and zonula occludens-1 were measured, suggestive of improvement in endothelial barrier function^{26 28}.

The presence of cannabinoid receptors in the central nervous system accounts for the psychoactive effects of cannabis^{24 25}. In contrast, CB2 receptors are abundant on immune cells including T and B cells, natural killer cells, monocytes and neutrophils as well as the liver^{24 29}. Two primary active constituents found in hemp plants include Δ 9-tetrahydrocannabinol (THC) and cannabidiol (CBD). Δ 9-THC is a partial agonist at both CB1 and CB2^{3 30 31} while CBD is

thought to mediate its effects through a variety of signaling mechanisms and act as a negative allosteric modulator at CB1^{32 33}.

Cannabis is perhaps best known for its effect on stimulating appetite via CB1 receptor activity and was historically used for this purpose in AIDS wasting syndrome. A variety of randomized controlled trials have shown that smoked cannabis is efficacious in the management of chronic pain, including in painful HIV-associated sensory neuropathy³⁴⁻³⁶. In animal models, Δ^9 -THC prevents atherosclerosis by inhibiting macrophage migration into atheromas through CB2 activation^{37 38}. Endocannabinoids may also play a role in liver disease, where CB1 receptors are present in endothelial cells and hepatocytes and CB2 receptors are distributed in Kupffer cells^{39 40}. CB1 receptors have an important role in non-alcoholic fatty liver disease and alcoholic liver disease while anti-inflammatory action of CB2 receptors can be useful in inflammatory liver disease. CB2 receptors show antifibrinogenic properties and administration of its agonists in fibrotic rats resulted in improvement in liver fibrosis, decreased inflammation and increased apoptosis of hepatic myofibroblasts⁴⁰⁻⁴⁴.

Cannabis, the immune system and HIV

In vitro studies

Cannabinoids have been shown to inhibit productive HIV infection in primary human T cells and a CB2 antagonist blocked this effect⁴⁵. Interference with the signal transduction of chemokine receptor CXCR4 is thought to lead to reduced F-actin accumulation. This in turn prevents movement of viral pre-integration complexes to the nucleus. It has also been postulated that CB2 agonists may inhibit T cell activation induced by anti-CD3/anti-CD28⁴⁵. Along with reduced HIV production, immunological effects of cannabinoids include induction of immunosuppressive cytokines including IL-10, TGF- β and inhibition of IL-2 which stimulates T-cell division and expansion. They have also been shown to decrease adhesion capacity of leukocytes to extravasate into sites of inflammation⁴⁶⁻⁴⁸. Apoptosis and induction of Treg may potentially add to the pathways of cannabinoid-mediated immunosuppression⁴⁹.

Animal studies

Mice treated with Δ 9-THC experience persistent inhibition of IL-2 production even 7 days after treatment⁵⁰. Rhesus macaques who were administered Δ 9-THC for 28 days prior to Simian Immunodeficiency virus (SIV) inoculation had reduced mortality and reduced SIV viral load in the cerebrospinal fluid and plasma. In another study involving macaques infected with SIV for 17 months of chronic THC administration⁵¹, Δ 9-THC resulted in trends towards a

decreased viral load. The lack of statistical significance in some parameters can be attributed to the small number of animals in the vehicle vs. the Δ^9 -THC groups ($n=4$)³⁰.

Human studies

To date, seven randomized controlled studies have examined the medicinal use of cannabis for reducing morbidity and mortality in people living with HIV⁵². Interventions included smoked marijuana or hashish, ingested marijuana or hashish or ingested THC (dronabinol or other synthetic cannabinoid). Studies ranged from 21-84 days. Primary outcomes included HIV-related mortality (all-cause), morbidity (including type and duration of episodes of opportunistic infections, malignancies, incidence of AIDS, and hospital admissions). Secondary outcomes included change in appetite, nausea, mood, pain, quality of life, body composition, hematological and nutritional markers, cognitive function, respiratory function and effect on pharmacokinetics of antiretrovirals and development of dependence or adverse sociological effects. The use of cannabis posed challenges for blinding due to the psychoactive effects⁵².

Only 1 study involving 62 patients examined the effects of cannabinoids on immune function and parameters associated with HIV infection. Participants in this study were assigned to either a 3.95% THC marijuana cigarette, a 2.5 mg dronabinol capsule or a placebo capsule three

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

times per day for 21 days. For the marijuana group, there was a statistically significant increase in CD4 counts from baseline vs. placebo and for the dronabinol group there was a trend towards statistical significance. Neither CD4 nor CD8 cell counts were adversely affected, and the pharmacokinetic component of the study did not reveal any clinically significant interactions that would require dose adjustments of protease inhibitors²². Adverse effects across studies included concentration difficulties, fatigue, sleepiness or sedation, increased duration of sleep, reduced salivation and thirst which improved upon discontinuation. The authors of a systematic review on the use of cannabis for reducing morbidity and mortality in HIV concluded that 1) evidence for substantial effects on morbidity and mortality is currently limited and 2) evidence for safety and efficacy of cannabis is lacking. Studies have been of short duration, in small numbers of patients and have focused on short-term measures of efficacy⁵². Furthermore, no study examined the effects of cannabinoids on inflammatory markers or HIV reservoir markers through a randomized trial.

More recently, Manuzak *et al.* published an observational study assessing the effect of cannabis use on peripheral immune cell frequency, activation, and function in 198 people living with HIV⁵³. Individuals were grouped into heavy, medium, or occasional cannabis users or noncannabis users as determined by the quantify cannabis metabolite 11-nor-carboxy-tetrahydrocannabinol (THCCOOH) detected in plasma by mass spectrometry. They found that

persons with heavy cannabis use had lower frequencies of HLA-DR+CD38+CD4+ and CD8+ T-cell frequencies compared to people living with HIV⁵³. Furthermore, heavy cannabis use was associated with decreased frequencies of pro-inflammatory intermediate (CD14++CD16+) and non-classical (CD14+CD16+) monocyte subsets⁵³. They also documented a reduction in antigen-producing cells secreting pro-inflammatory cytokines IL-23 and tumor necrosis factor (TNF)- α ⁵³. Rizzo *et al.* also demonstrated that levels of circulating CD16 monocytes and interferon-gamma-induced protein (IP)-10 from people living with HIV who either were or were not cannabis users⁵⁴. Lower levels of CD16+ monocytes and plasma IP-10 were found in cannabis users compared to non-cannabis users⁵⁴. However, this study did not quantify the level of cannabis exposure in the two groups. Although these studies demonstrated favorable associations between inflammation and cannabis use, it must be borne in mind that both of these studies were observational only. As these studies were not randomized controlled trials, it is possible that people living with HIV who used cannabis in these studies differed in other significant ways from PLWHIV who did not use cannabis.

Study rationale

Cannabis may hold many potential therapeutic benefits for people living with HIV due to its promising anti-inflammatory and anti-fibrotic effects. Before adequately-powered interventional studies can be designed to study cannabis as a therapy for specific conditions associated with chronic inflammation and fibrosis, a key first step will be to demonstrate that cannabinoid use in a clinical trial is feasible and that they have a favorable safety and tolerability profile. As such, we propose a proof-of-concept pilot study to examine the feasibility, safety and tolerability of cannabinoid oils consumed orally in people living with HIV on effective ART. As a secondary objective, we will examine the effect of cannabinoid oils on immune profiles, including levels inflammatory markers associated with HIV disease progression and frequencies of activated and senescent CD4 and CD8 T-cells. Frequencies of regulatory T cells and various subsets of Th17 will also be assessed. Finally, an exploratory objective will be to study the effect of cannabinoid oils on markers of HIV persistence and the composition of the gastrointestinal microbiome.

We propose to use combination therapy of THC:CBD oils in capsule format (TN-TC11LM and TN-TC19LM capsules) ingested orally to examine these outcomes. Although research to date involving HIV/SIV has examined THC, data from *in vitro*, animal and human studies suggests that CBD has favorable anti-inflammatory effects and the combination of CBD with THC increases tolerability⁵⁵⁻⁵⁹.

Methods/Design

Study design

This is a randomized, open-label, interventional study (the Canadian HIV Trials Network (CTN) number PT028) whereby capsules containing CBD:THC oils are consumed for 12 weeks to assess safety and tolerability. Their ability to reduce immune activation (as determined by percentage of activated CD8+CD38+HLA-DR+ T-cells), size of the peripheral HIV reservoir and change in gastrointestinal microbiome composition will also be examined. Participants will continue to take their ART treatments as prescribed throughout the study.

Setting

Recruitment of participants will occur at the Chronic Viral Illness Service (CVIS), Royal Victoria Hospital (Glen campus) of the McGill University Health Centre (MUHC), the largest academic HIV clinic in Canada.

Recruitment and enrollment

Study staff at the CVIS will conduct chart reviews of prospective people living with HIV ahead of their clinic visits to determine which persons have had suppressed viral load for at least 3 years on ART. The patient chart will be flagged, and if the treating HIV physician believes the person to be suitable for the study, the physician or study staff will approach potential trial participants at their clinic visit. The trial staff will inform the patients about the trial and invite him or her for eligibility screening and possible trial enrolment. Participant eligibility will be documented and written informed consent obtained for eligible patients by the study coordinator. The study coordinator will systematically document all individuals who have been approached for the study in addition to reasons for acceptance and refusal to participate in the study. Individuals who wish to discuss their participation in the study with their treating physician and or family and friends will be have the opportunity to do and may enroll a their next scheduled clinic visit. Following enrolment, participants will be followed during the study by the principal investigator and study coordinator at the CVIS.

Inclusion criteria

Eligible participants must meet the following criteria within 4 weeks prior to beginning the cannabinoid capsules to be considered eligible for study entry: 1) documented HIV infection

by Western blot, enzyme immunoassay or viral load assay; 2) aged 18 or older; 3) viral load <40 copies/mL for at least the last 3 years (maximum 2 blips <500 copies/ml allowed); 4) no cannabinoid use for at least 1 month prior to enrolment with negative baseline cannabinoid urine screen; 5) able to communicate in either English or French. We will attempt to enrol equal proportions of men who have sex with men (MSM) and heterosexual individuals.

Exclusion criteria

Individuals who meet any of the following criteria will be ineligible to participate: 1) using cannabinoid-containing products outside of the study or within 4 weeks of study commencement; 2) pregnant, breastfeeding or planning to become pregnant during the course of the study; 3) enrolled in a separate study involving administration of medication, vitamin, supplement or herbal product; 4) active intravenous drug use; 5) active substance dependence; 6) prior history of hypersensitivity to cannabis or cannabis-containing products; 7) known or suspected allergy to sunflower lecithin oil; 8) active opportunistic infection or malignant condition; 9) unintentional weight loss of 10 % or more of body weight in the last 6 months; 10) unstable angina or acute cardiac event in the past year; 11) active psychiatric disorder or history of psychiatric depression (other than mild depression or anxiety); 12) on antipsychotic

medication; 13) known or suspected family history of schizophrenia or severe personality disorder; 14) serious cardiovascular disease such as ischemic heart disease, arrhythmias, poorly controlled hypertension, or severe heart failure; 15) anemia (Hemoglobin <100 g/L); 16) active liver disease or unexplained persistent elevations of serum transaminases; 17) Co-infection with Hepatitis B or C (positive HBsAg or positive anti-HBc antibodies with a detectable HBV DNA viral load or positive anti HCV antibodies with a detectable HCV RNA viral load); 18) alanine aminotransferase (ALT) or Aspartate aminotransferase (AST) or alkaline phosphatase >2.5 x upper limit of normal (ULN); 19) opportunistic infection in the last month as determined by the treating physician; 20) renal dysfunction; 21) unstable psychological or psychiatric condition as determined by the treating physician; 22) holding employment which requires operation of heavy machinery or which requires undergoing drug screening (ie, pilot or police officer); 23) concurrent use within the past 8 week of anabolic hormones, prednisone, IL-2 or other agents known to alter immune function.

The investigators will review potential participants' current medication lists at the screening visit. If any concomitant therapy interacts with the study medication, and if this therapy cannot be substituted, that participant will not be eligible to enroll in the study. Although some HIV antiretroviral drugs/pharmacokinetic "boosters" are metabolized by the CYP 1A2 and CYP3A4 (e.g., ritonavir and cobicistat-boosted protease inhibitors), suggesting that a drug

interaction may occur, in real practice at the Chronic Viral Illness Service no clinically significant drug interactions have been observed in patients reporting heavy cannabis use. Therefore, individuals will not be precluded to participate based on their specific antiretroviral regimen.

Study intervention

The study medications are TN-TC11LM and TN-TC19LM capsules which contain THC:CBD in a ratio of 1:1 (2.5 mg/2.5 mg) and 1:9 (5 mg/45 mg), respectively. These study drugs are being provided by Tilray and the active pharmaceutical ingredients are extracted from the cannabis plant and purified according to pharmaceutical standards (>98%). Participants will be advised to gradually increase the number of capsules they take based on the suggested titration scheme presented in Tables 1 and 2, until a daily maximum is reached. These maximum amounts are comprised of 10 capsules of TN-TC11LM (25 mg THC/25 mg CBD total per day) or 3 capsules of TN-TC19LM (15 mg THC: 135 mg CBD for TN-CT19L) per day. These doses were selected as in a clinical trial for neuropathic pain, doses equivalent to 2.5 mg of THC were well-tolerated³⁴. More recently, among patients (ages 2-55 years) with the Lennox-Gastaut syndrome, cannabidiol at a dose of 10 or 20 mg per kilogram per day resulted in greater reductions in the

frequency of drop seizures than placebo and was well-tolerated overall other than for an increase in liver aminotransferase concentrations⁶⁰. Due to person-to-person variability in the ability to metabolize and tolerate cannabinoids⁶¹, we have opted for patients to titrate their dose of medication to a range where they are comfortable as the titration method of dosing has proven successful in other clinical trials involving cannabinoids³⁴.

Table 1: Recommended titration schedule for Group 1 – Low CBD dose TN-TC11LM oral capsules

Weeks	Daily Dose	Number of capsules
1	5 mg THC/ 5 mg CBD	1 capsule twice daily, taken orally
2	10 mg THC/ 10 mg CBD	2 capsules twice daily, taken orally (4 capsules per day)
3	15 mg THC/ 15 mg CBD	2 capsule three times daily, taken orally (6 capsules per day)
4	20 mg THC/ 20 mg CBD	2 capsules four times daily, taken orally (8 capsules per day)
5-12	25 mg THC/25 mg CBD	2 capsules 5 times daily, taken orally (10 capsules per day)

Group 1: Low CBD dose TN-TC11LM oral capsules (2.5 mg THC/2.5 mg CBD capsules). This group will be advised to start by taking 1 capsule twice daily for 1 week (5 mg THC/5 mg CBD) and increase the number of capsules as tolerated to a maximum of 10 capsules daily by weeks 5-12 (25 mg THC/25 mg CBD total per day). Participants will record the times and dates of all capsules consumed in a logbook.

Table 2: Suggested titration schedule for Group 2 – High CBD dose TN-TC19LM oral capsules

Weeks	Daily Dose	Number of capsules
1	5 mg THC/ 45 mg CBD	1 capsule daily, taken orally
2	10 mg THC/ 90 mg CBD	1 capsule twice daily, taken orally (2 capsules per day)
3	15 mg THC/ 135mg CBD	1 capsule three times daily, taken orally (3 capsules daily)
4	20 mg THC/ 180 mg CBD	1 capsule four times daily, taken orally (4 capsules)
5-12	25 mg THC/ 225 mg CBD	1 capsule five times daily, taken orally (5 capsules)

Group 2: High CBD dose TN-TC19LM (5 mg THC/45 mg CBD capsules). This group will be advised to start by taking 1 capsule once daily for 1 week (5 mg THC/45 mg CBD) and increase the number of capsules as tolerated to a maximum of 10 capsules daily by week 5(25 mg THC/225 mg CBD total). Participants will record the times and dates of all capsules consumed in a log book.

1

2

3

4

5

6

7

8 **Randomization**

9

10

11 After eligibility is confirmed and written informed consent obtained, participants will be

12

13

14 randomized to either TN-TC11LM (group 1) or TN-TC19LM (group 2) capsules which contain

15

16

17 THC:CBD in a ratio of 1:1 and 1:9, respectively. Prior to study commencement, a statistician

18

19

20 unassociated with the study will develop a randomization scheme using SAS and input into a

21

22

23 password-protected web-based randomization system. Variable block sizes of 2 and 4 will be

24

25

26 used. Participants will be assigned to either Group 1 vs. Group 2 based on the pre-designated

27

28

29 allocation code. As this is an unblinded study, participants and study staff will be aware of the

30

31

32 group to which the participant has been randomized. A computerized audit trail will track date

33

34

35 and time of allocation, patient study identification number and treatment allocation. The

36

37

38 randomization group will be recorded in the study log, which will be accessible to the

39

40

41 sponsor/medical manager and study coordinator.

42

43

44

45

46

47

48 **Measurements**

49

50

51

52 At the screening visit, clinical information will be collected from each participant

53

54

55 including age, ethnicity, sexual orientation, list of current medications, dosage, date of treatment

56

57

58

59

60

initiation, any antimicrobials taken in the previous 6 months, psychiatric disorders, duration of HIV infection, current ART regimen and duration of ART regimen months, ART history, CD4+ T cells count within the past 3 months, nadir CD4+ T cells count, CD4/CD8 ratio, duration of plasma viral load suppression and any preexisting medical conditions, signs or symptoms. Information will also be collected on whether the individual consumed cannabis in the past, the form(s) in which it was consumed, frequency of use and reasons for use.

Clinical parameters

Scheduled visits will occur to monitor safety and tolerability, as per the visit schedule depicted in Table 3. Visits will include physical exam with vital signs, weight, occurrence of adverse events (AEs and concomitant medications) and the presence of common symptoms associated with cannabinoids including dizziness, nausea, headaches, appetite or mood changes. At visits, blood for some or all of the following will be collected: CD4+ T cells count, CD8+ T cells, CD4/CD8 ratio, plasma viral load, Complete Blood Count (CBC), aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP), total bilirubin, urea, creatinine and blood glucose, T-cell activation and inflammatory markers and testing for syphilis if the participant tested positive during the 4 weeks prior to beginning

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

consuming the study capsules. A stool sample for analysis of the bacterial and fungal microbiome assessment will also be collected prior to beginning the study capsules. Participants will be enrolled in the study for up to 15 weeks but will consume capsules for a period of 12 weeks. Participants will undergo screening tests and eligibility assessment within 4 weeks prior to initiating study capsules. Participants will then undergo assessments after the first week of capsule consumption and every 2 weeks thereafter. A second stool sample for bacterial and fungal microbiome analysis will be collected during the final week of capsule consumption. The final visit will occur 2 weeks after study drug cessation.

Table 3. Schedule of Visits and Procedures

Visit Type	Screening	Visit 1 - Baseline 1	Visit 2 - Baseline 2	Visit 3	Visit 4	Visit 5	Visit 6	Visit 7 - Superintendence (AES)	Visit 8	Visit 9 - End of Tx Visit	Visit 10 - Final Study Visit
Visit Window	-4 to -1 weeks	W -1	D1, W0	D1, W1 (± 4 days)	D1, W2 (± 4 days)	D1, W4 (± 4 days)	D1, W6 (± 4 days)	D1, W8 (± 4 days)	D1, W10 (± 4 days)	D1, W12 (± 4 days)	D1, W14 (± 4 days)
Procedures:											
Eligibility criteria assessment	X										
Informed Consent	X										
Medical History	X										
Pregnancy Test (urine) ¹	X										
Cannabinoids Screen (urine)	X										
Hepatitis B, C and syphilis	X ³										
Randomization			X								
Physical Exam			X	X	X	X	X	X	X	X	X
Hematology and chemistry profiles ²	X			X	X	X	X	X	X	X	
Viral load, CD4 and CD8	X				X		X			X	
Immune activation and inflammatory markers			X	X	X		X			X	X
HIV reservoir assays		X	X	X	X		X	X		X	X
Nasal swab and stool specimen for microbiome assessment		X								X	
WHOQOLHIV-BREF scale			X				X			X	
EQ-5D questionnaire			X				X			X	
POMS questionnaire			X				X ⁵			X	
Study medication dispensed			X	X	X	X	X	X	X		
Study Drug Compliance ⁴				X	X	X	X	X	X	X	
ART Compliance ⁴		X	X	X	X	X	X	X	X	X	X
Alcohol Intake		X	X	X	X	X	X	X	X	X	X
Adverse Events				X	X	X	X	X	X	X	X

Visit Type	Screening	Visit 1 - Baseline 1	Visit 2 - Baseline 2	Visit 3	Visit 4	Visit 5	Visit 6	Visit 7	Visit 8	Visit 9 - End of Tx Visit	Visit 10 - Final Study Visit
Visit Window	-4 to -1 weeks	W -1	D1, W0	D1, W1 (± 4 days)	D1, W2 (± 4 days)	D1, W4 (± 4 days)	D1, W6 (± 4 days)	D1, W8 (± 4 days)	D1, W10 (± 4 days)	D1, W12 (± 4 days)	D1, W14 (± 4 days)
Procedures:											
Concomitant Medications/Therapies	X	X	X	X	X	X	X	X	X	X	X

- ¹ If urine test is positive, perform serum pregnancy test.
- ² Complete blood count; AST, ALT, ALP, total bilirubin, urea, creatinine and blood glucose.
- ³ If participant tests positive for Hepatitis B and C, he/she will no longer be eligible for the study. If participant tests positive for syphilis, he or she will be treated for syphilis according to clinical care guidelines and will still be eligible to participate in the study. The need for syphilis treatment and follow-up testing (usually in 6 and 12 months) will be discussed between the Sponsor and the investigator at the CVIS. It will be up to the investigator to ensure proper follow-up and management of the syphilis, as this is part of standard of care.
- ⁴ Assessed by reviewing log book provided to each participant
- ⁵ Review individual POMS questionnaires completed at Visits 2 and 6 with each participant

Medication adherence

Participants will be asked to keep a log book in which they enter the number of TN-TC11LM or TN-TC19LM capsules consumed, the time, as well any adverse effects they noted and the timing of these adverse effects relative to capsule intake. Furthermore, individuals will be asked to record whether or not they took their ART that day or whether any doses were missed. Participants will be asked to bring their log books with them to study visits and the coordinator will photocopy this information.

Quality of life and mood assessment

Questionnaires measuring quality of life (World Health Organization Quality of Life HIV-BEF (WHOQOLHIV-BREF) and the EQ-5D) and mood (Profile of Mood States (POMS)) will be administered at baseline, midway through the study and at the end of the study, as outlined in Table 3. WHOQOLHIV-BREF consists of 31 items which measure the following domains: physical health, psychological health, social relationships and environment. It is a shorter version of the original instrument (WHOQOL) and is more convenient for use in clinical trials, taking approximately 10 minutes to complete. EQ-5D is a descriptive questionnaire examining 5 dimensions: 1) mobility, 2) self-care, 3) usual activities, 4) pain/discomfort, and 5)

anxiety/depression. Each dimension has 5 levels: No problem, slight problems, moderate problems, severe problems and extreme problems. The participant indicates the state of his/her health by ticking the box most appropriate to the statement in each of the 5 dimensions. This decision results in a 1-digit number that expresses the level selected for that dimension. The digits for the 5 dimensions can be combined into a 5-digit number that describes the participant's health state. It takes about 10 minutes to complete. The Profile of Mood States (POMS) questionnaire measures the following 6 factors: 1) Tension-Anxiety, 2) Anger-Hostility, 3) Fatigue-Inertia, 4) Depression-Dejection, 5) Vigor-Activity, and 6) Confusion-Bewilderment. It is very sensitive to non-clinical changes in mood states and takes approximately 5 minutes to complete. These questionnaires will be administered by a trained research coordinator.

Research hypothesis

THC:CBD oils consumed orally – as TN-TC11LM and TN-TC19LM oral capsules – will be safe and well-tolerated in PLWHIV. They will also be associated with a reduction in markers of inflammation, reduction in frequency of activated T cells and reduction in HIV reservoir size.

Study outcome measures

The primary objective is to evaluate the safety and tolerability of TN-TC11LM and TN-TC19LM oral capsules in PLWHIV on effective ART. The primary between group comparison is the percentage of participants without any signs of significant toxicity; percentage of participants who are able to complete the study and scores on the WHOQOLHIV-BREF Scale, EQ-D5 and POMS questionnaires from week 0 to week 12 are secondary outcomes that will also be compared between groups. The secondary objective is to determine the effect of TN-TC11LM and TN-TC19LM oral capsules on frequency of activated T-cells and markers of inflammation association with HIV disease progression. Exploratory objectives are to determine the effect of TN-TC11LM and TN-TC19LM oral capsules on 1) the size of the peripheral HIV reservoir 2) the composition of the gastrointestinal bacterial and fungal microbiome.

Safety and tolerability

Safety will be assessed at regular intervals (Table 3) by vital signs and adverse effects (AE) monitoring, as reported by the participant and actively sought at each study visit by the coordinator or physician. Biological safety will be evaluated by hematology, biochemistry and other clinical, laboratory or other diagnostic tests done on participants during the course of the study. Lab results for all participants for assessed safety variables will be reviewed by the trial

investigator. The Data safety monitor will also review safety information. Toxicity of TN-TC11LM and TN-TC19LM will be assessed using the World Health Organization (WHO) toxicity scale. All AEs, regardless of the grade, will be documented and it will be noted whether or not these symptoms were already present at baseline. Any AEs that occur during the study will be evaluated by the trial investigators and grade 3 and 4 AEs will be recorded on the CRFs. If required, blood specimens will be collected for hematology and biochemistry tests. Participants having AEs will be monitored with relevant clinical assessments and laboratory tests, as determined by the trial investigator. The trial investigator will report ongoing AEs at the completion of the clinical study to the primary treating physician at the CVIS who will determine the need for and provide standard medical care. The trial investigator will ensure that the event is satisfactorily resolved or that no additional follow-up is needed. Any participant who discontinues the study for an unresolved clinically significant AE will be followed until satisfactory clinical resolution is achieved and the AE recorded on the case report form (CRF), regardless of severity grading. AEs that may be related to TN-TC11LM and TN-TC19LM will be managed by dose reduction of TN-TC11LM and TN-TC19LM. TN-TC11LM and TN-TC19LM will be discontinued permanently in the event of any life-threatening AEs.

T-cell subsets and Immune activation

The frequency of different CD4⁺ and CD8⁺ T cell subsets will be defined using multi-parameter flow cytometry (BD Fortessa X-20) in peripheral blood. The expressions of CD3, CD4 and/or CD8, CD45RA, CCR7 and CD27 will be used to measure the frequency of naïve (CD45RA⁺CCR7⁺CD27⁺), central memory (CD45RA⁻CCR7⁺CD27⁺), transitional memory (CD45RA⁻CCR7⁻CD27⁺), effector memory (CD45RA⁻CCR7⁻CD27⁻) and terminally differentiated (CD45RA⁺CCR7⁻CD27⁻) cells. Regulatory T (Treg) cells will be defined as CD3⁺CD4⁺CD25^{high}FoxP3^{high}CD127^{low} cells. Expression of the CD39 and CD73 ectoenzymes involved in Treg-mediated immunosuppression and HIV disease progression (via the adenosine pathway) will be also assessed^{62 63}. Various subsets of Th cells (T helper) will be defined as Th1 (CD45RA⁻CCR6⁻CCR4⁻CXCR3⁺), Th2 (CD45RA⁻CCR6⁻CCR4⁺CXCR3⁻), Th17 (CD45RA⁻CCR6⁺CCR4⁺CXCR3⁻) and Th1/Th17 (CD45RA⁻CCR6⁺CCR4⁻CXCR3⁺). Levels of CD8⁺ and CD4⁺ T cell immune activation (CD38/HLA-DR co-expression) and senescence (CD28⁻CD57⁺) will be also assessed on all T cell subsets. These markers will be assessed at week 0, 1, 2, 6, 12 and 14.

Inflammatory markers assessments

Plasma levels of various inflammatory markers including interferon- α , interleukin (IL)-1 β , IL-6, IL-10, IL-17, TGF- β , interferon-gamma-induced protein (IP)-10, will be assessed via Luminex (Millipore) and levels of d-dimer, C-reactive protein, and markers of microbial translocation lipopolysaccharide and sCD14 will be assessed by ELISA in batch from blood drawn at weeks 0, 1, 2, 6, 12 and 14.

Peripheral blood HIV reservoir size

Blood for HIV reservoir assessment will be collected at 2 time points prior to cannabis initiation (1 week prior to cannabinoid capsule initiation and immediately prior to cannabinoid capsule initiation) to account for normal fluctuations in baseline levels of HIV persistence markers. Subsequently, virological measures will be done at weeks 1, 2, 6, 8, 12, 14. CD4+ T cells isolated from PBMCs by magnetic negative selection. To capture all infected cells, the frequency of cells harbouring total and integrated HIV DNA will be measured using well-established assays on a total of 500,000 cells (sensitivity of 1 copy/reaction)^{17 64}. As most of the HIV genomes are defective, the recently developed “tat/rev induced limiting dilution assay” (TILDA), which provides a more functional measurement of the HIV reservoir⁶⁵, will be

employed. To assess if residual levels of viral replication may occur, we will measure 2-LTR circles, which are proposed to be a surrogate marker for ongoing HIV replication during ART⁶⁴^{66 67}. Specifically, this combination of assays will indicate if cannabis has an impact on the size of the total reservoir (DNA), the functional reservoir (TILDA) and ongoing viral replication (2-LTR circles). Measurements will be performed in batch.

Gastrointestinal Microbiome Composition

A stool sample without preservative will be collected from each participant at the beginning of the study prior to consuming the capsules and during the final week (week 12) of capsule consumption. Specimens will be stored at -80°C until analyzed in batch, as previously described⁶⁸. Bacterial DNA will be extracted with PCR amplified targeting of the 16S rRNA gene using universal primers which flank the V3-V4 region of the 16S gene modified with the addition of TruSeq Illumina adapters, also as previously outlined⁶⁹. Internal Transcribed Spacer for Fungal DNA extraction will be used for fungi. PCR amplification, PCR amplicon quantification and sequencing will be performed as previously described^{68 69}.

Sample size and statistical analyses

In this proof-of-concept study, we are exploring a phenomenon with little *in vivo* data and with a limited study budget. For this reason we have chosen a convenience sample of 26 participants, 13 per arm, without formal power calculations. This number of individuals will enable us to assess feasibility (willingness of patients to participate, attend study visits and complete questionnaires, numbers of drop-out participants) as well as safety and tolerability. The data obtained will help to guide future sample size calculations for future studies. Although the small number of participants may result in wide confidence intervals for adverse events, this number of participants should give us an idea about the degree of variability for continuous outcomes we are measuring.

For the primary endpoint, the proportions of participants without any signs of significant toxicity (Grades 0-2 scores on the WHO toxicity scale), proportions of participants who complete the study and scores on the WHOQOLHIV-BREF, EQ-5D and POMS questionnaires will be examined using descriptive statistics. We will also compare these proportions for Group 1 vs. Group 2 using a Fisher's exact test. For Quality of Life and mood measures, we will use analysis of covariance with 12 week score as outcome and baseline score as covariate and treatment as independent variable. With regards to the POMS questionnaire, we will consider only overall scores (and not sub-scores) due to the small sample size which would make comparisons of the sub-scale inappropriate.

Immune activation levels for Groups 1 vs. Group 2 at week 12 will be compared using analysis of co-variance with adjustment for the week 0 activation levels. The mean change and associated 95% confidence interval will be reported for each of the secondary endpoints described above. Furthermore, the study drug treatment period will be compared to the baseline period with each arm. If the treatment effect is similar within the two arms, then an analysis of the treatment effect will be pooled over the two arms, using analysis of covariance. The change in immune activation levels following discontinuation of study drug (i.e., from week 12 to week 14) will be reported as a mean with corresponding 95% confidence interval. A 50% reduction will be considered significant⁷⁰.

Analogous analyses will be conducted for reservoir assessments other outcome measures listed above as endpoints. A signed rank (non-parametric) test will be used to compare number of copies of total and integrated DNA at baseline 2 vs. at week 12. Group differences in the change of HIV reservoir size from baseline to 12 weeks will be assessed by the Mann-Whitney *U* test. Wilcoxon signed-rank test will be used to compare the HIV reservoir and inflammatory markers in blood samples of the same patient from baseline to 12 weeks. At least a 2-fold decrease in frequency of infected cells in both groups from baseline to 12 weeks of treatment will be considered significant⁷¹. Microbiome composition will be described with regards to the

frequencies of microorganisms families for the groups at baseline and then at 12 weeks of treatment. Due to the exploratory nature of this objective, no formal statistics will be applied.

Patient and Public Involvement

The CTN Community Advisory Committee (CAC) was involved in the peer review process of this study proposal, deemed that the research questions addressed were of very high priority to people living with HIV and voted for the funding of this study. The CAC’s critiques of the initial proposal were taken into account in the revised proposal. Two members of the CAC (SM and EM) were involved in finalizing the study design, inclusion/exclusion criteria, outcome measures and monitoring plans and are formal study investigators and co-authors. Preliminary and final results of the study will be shared with community members, patient participant and the public at bi-annual CTN meetings, through the CTN newsletter and on the CTN website in addition to the annual Canadian Association of HIV Research meeting.

Data management

All clinical data and electronic files will be stored in the secure environment of the CVIS of the MUHC. All data published will be anonymized. Only researchers affiliated with the

study will have access to participant data. Study progress and safety will be evaluated in an ongoing fashion by the principal and co-investigators. The study will be monitored for safety and ongoing progress by a standing Data and Safety Monitoring Committee (DSMC) of clinicians and methodologists established by the Canadian HIV Trials Network. The committee meets every six months or as needed.

Storage of biological specimens

. Within 1 hour of being drawn, blood from the CVIS will be transferred to the laboratory at the RI-MUHC (in the adjacent, connected building) and the plasma will be separated from peripheral blood mononuclear cells (PBMCs) by Ficoll density centrifugation by an experienced laboratory personnel. PBMCs and plasma will be stored in liquid Nitrogen tanks at the RI-MUHC laboratory until time for analysis. Patients will be contacted the day before their clinic visit to remind them when a stool specimen is due. They will be instructed to record the time of the provision of the sample on the paper bag containing the sterile container and store it in a refrigerator (-4C) until brought to the clinic. Once at the CVIS, the stool specimen will be placed in a large fridge designated specifically for the storage of stool specimens.

1

2

3

4 **Ethics and dissemination**

5

6

7

8 Written informed consent will be obtained from all study participants. The study

9

10

11 protocol and informed consent have been approved by the Research Ethics Board of the McGill

12

13 University Health Centre (MUHC-2018-4336) and is in the process of being reviewed by Health

14

15 Canada’s Therapeutic Product Directorate. The study will be conducted in accordance with the

16

17 application Health Canada regulations, International Conference on Harmonization guidelines on

18

19 current Good Clinical Practice and the Declaration of Helsinki. Patient enrollment for this trial is

20

21 anticipated to begin August 2018.

22

23

24

25

26

27

28

29 Regardless of outcome, trial results will be disseminated through scientific peer review

30

31 publication, international and national conferences and the CTN according the SPIRIT (Standard

32

33 Protocol Items: Recommendations for Interventional Trials) and CONSORT (Consolidated

34

35 Standards of Reporting Trials) guidelines for transparent reporting of trials^{72 73}. CTC will be

36

37 responsible for initially drafting the manuscripts and professional writers will not be used for any

38

39 of the publications. Authorship will be determined based on criteria defined by the International

40

41 Journal of Medical Editors⁷⁴. We aim to write the manuscript of the final results within 6 months

42

43 of completing the study. Participants who have been involved in the trial will be given the

44

45 option of having a summary of results sent to them.

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

Discussion

Since the advent of ART, people living with HIV now have a longevity which approaches that of their HIV-uninfected counterparts but have a higher burden of non-communicable comorbidities including cardiovascular, pulmonary, renal and hepatic diseases^{1 2}. Heightened inflammation in people living with HIV despite ART is believed to be the driving force behind the increased rates of non-infectious comorbidities. Similarly, chronic immune activation fosters HIV persistence¹⁰⁻¹⁵. As cannabinoids possess both anti-inflammatory and anti-fibrotic properties¹⁹, cannabinoids may represent a feasible method to reduce immune activation and enhance immune profile. This, in turn, may hasten the progression of non-opportunistic complications associated with HIV. Although some studies have examined whether there are beneficial effects on inflammation resulting from treatment with integrase inhibitors compared to protease inhibitors (PIs), between PIs and non-nucleoside reverse transcriptase inhibitors (NNRTIs), between specific nucleoside reverse transcriptase inhibitors, or with maraviroc in ART-naïve patients, to date insufficient to conclude that any class of antiretrovirals is superior to other classes of antiretrovirals with regards to effects on inflammation⁷⁵. Furthermore, cannabis may induce cytochrome P450 (CYP) 1A2 via activation of the aromatic hydrocarbon receptor⁷⁶. CYP3A4 inducers and inhibitors alter the pharmacokinetics of Δ^9 -THC and CBD when administered as Δ^9 -THC/CBD oromucosal spray. To date only one study has

ever examined the effects of cannabinoids on the pharmacokinetics of antiretrovirals. Kosel et al. studied the pharmacokinetics of smoked marijuana and dronabinol in people living with HIV receiving either indinavir and nelfinavir (2 protease inhibitors no longer used due to their toxicity and adverse effect profiles)⁷⁷. Individuals on stable regimens of indinavir 800 mg every 8 hours or nelfinavir 750 mg three times daily were randomized to one of three treatment arms: 1) 3.95% THC marijuana cigarettes 2) dronabinol 2.5 mg capsules or 3) placebo capsules given three times daily. Although there were statistically significant decreases in maximum concentration (C_{max}) of indinavir in the smoked marijuana arm, the size of the changes in the pharmacokinetic parameters of both indinavir and nelfinavir were sufficiently small not to impose any short-term clinical consequence⁷⁷. Furthermore, the investigators concluded that use of marijuana or dronabinol is unlikely to impact antiretroviral therapy⁷⁷. For these reasons and based on clinical experience at our clinic, we have not precluded individuals on any particular antiretroviral regimens from participating in this study.

In this pilot study, our primary objective is to assess the safety and tolerability of TN-TC11LM and TN-TC91LM taken by people living with HIV on suppressive ART. We hypothesize that these agents will be safe and well tolerated in people living with HIV, given that similar products are safe and well-tolerated in other populations. Sativex® is currently licensed as an adjunctive treatment for symptomatic relief of spasticity in adult patients with Multiple

1
2
3
4 Sclerosis (MS) who have not responded adequately to other therapy⁷⁸⁻⁸⁰. It is an oromucosal
5
6
7 spray containing CBD and THC in 1:1 ratio. Marinol®, which is a synthetic THC-containing
8
9
10 capsule, is currently used for the treatment of anorexia associated with weight loss in persons
11
12
13 with Acquired Immunodeficiency Syndrome (AIDS) and nausea and vomiting associated with
14
15
16 cancer chemotherapy in patients with insufficient response to conventional antiemetics⁸¹. It has
17
18
19 been estimated that 5,472 patients have been exposed to Sativex® and there have been no safety
20
21
22 concerns identified and the product remains well-tolerated. The primary safety concerns of both
23
24
25 Sativex® and Marinol® are consistent with the known pharmacological activity of cannabinoids.
26
27
28 The primary safety concerns associated with Sativex® included abuse potential, cardiovascular
29
30
31 effects and central nervous system adverse effects⁸². Although there is some evidence to suggest
32
33
34 that individuals can develop “cannabis use disorder”, individuals do not develop the same
35
36
37 extremes of behavior as observed with other drugs of abuse⁸³. In two clinical trials, nabiximols
38
39
40 such as Sativex® have been used in two clinical trials whereby treatment was abruptly ceased to
41
42
43 study whether withdrawal symptoms would develop^{84 85}. In both studies, no withdrawal
44
45
46 syndromes were observed⁸⁶. Cannabinoids have cardiovascular effects that include tachycardia
47
48
49 and fluctuations in blood pressure, including episodes of postural hypotension. Therefore these
50
51
52 agents should not be used in patients with serious cardiovascular disease, such as ischemic heart
53
54
55 disease, arrhythmias, poorly controlled hypertension or severe heart failure. THC has complex
56
57
58
59
60

effects on the CNS and should not be used in patients with a personal or strong family history of psychosis. Examples of such conditions include schizophrenia and affective psychosis since symptoms of these disorders may be aggravated by cannabinoids. In multiple sclerosis (MS) patients receiving Sativex® in clinical studies, psychiatric-related adverse effects included disorientation, depression including depressed mood, dissociation, euphoric mood, hallucination, hallucinations (auditory and visual), illusions, paranoia, suicidal ideation and delusional perception⁸⁷. Interestingly, there is some evidence to demonstrated that CBD may actually improve psychotic symptoms in persons suffering from schizophrenia⁸⁸.

For Marinol®, which contains only THC, the most frequency reported adverse effects experienced by patients with AIDS during placebo controlled clinical trials involved CNS and were reported in 33% of patients receiving Marinol®. About 25% reported a CNS adverse event during the first 2 weeks and about 4% reported such an event each week for the next 6 weeks thereafter⁸⁹. By combining CBD with THC, we anticipate that tolerability will be greatly enhanced. When combined with THC, CBD reduces the risk for many adverse effects ⁸⁸. Furthermore, individuals will be instructed to titrate up the dose based on their own tolerability and reduce the dose if they experience any undesirable effects. Furthermore, due to the extremely low levels of CB1 receptors in the brainstem⁹⁰, death due to overdosing on cannabis or cannabinoids alone has never been described.

The study medications and doses were chosen after a lengthy review of the existing literature and discussion with experts in the field of pain management. A high degree of inter-individual variability in metabolism following administration of cannabinoids is observed due to polymorphisms in cytochrome isoenzymes⁶¹. Given that the therapeutic doses of cannabinoids are highly variable between individuals, a dose titration schedules are usually recommended. When used to treat specific conditions, persons may be told to increase the dose until they achieve adequate symptom relief without adverse effects. This method was observed to work well when used in the first cohort study on the long-term safety of medicinal cannabis for non-cancer chronic pain in seven Canadian clinics³⁴.

Given this is a pilot study and given our budgetary restrictions, a convenience sample of 26 participants was selected without formal power calculation. If this study demonstrated that TN-TC11LM and/or TN-TC91LM are safe and tolerable in people living with HIV and can reduce systemic inflammation, future studies will be performed to address the potential of these agents to ameliorate specific conditions in people living with HIV. Should future studies be conducted, data generated from this trial will assist with power calculations. Similar to a study conducted by members of our group on the ability of Niaspan® (extended-release niacin) to reduce immune activation, as determined by percentage of activated CD8+ CD38+ HLA-DR+ T-cells, we decided that a 50% reduction in activated CD8+ CD38+ HLA-DR+ T-cells would be

considered significant⁷⁰. This is based on previous reports indicating that a 10-fold difference exists between uninfected healthy controls and treated aviremic HIV-infected individuals in level of activated CD8+ CD38+ HLA-DR+ T-cells^{91 92}. All of the inflammatory mediators we selected for this study are known to drive immune activation^{1 6 93}.

In addition, we decided to make our objective examining the ability of TN-TC11LM and TN-TC91LM capsules to reduce the HIV reservoir, through the reduction of systemic inflammation, an exploratory objective. It is unclear if 3 months of treatment will be long enough to produce any meaningful reduction in the size of the HIV reservoir. Furthermore, it is unclear what reduction in reservoir size is required to have a meaningful effect on clinical outcomes. As mentioned earlier, we will consider a 50% decrease in the number of HIV-infected cells at baseline vs. week 12 to be a significant reduction in the reservoir, based on a study by *Hill et al*⁷¹. To our knowledge, there is no other randomized clinical trial examining the effect of cannabinoids on inflammation and HIV reservoir size in people living with HIV.

Our study is unique in being the first randomized trial in the world to examine the association between ingestion of precise quantities of cannabinoids and effect on inflammation and peripheral HIV reservoir size. It is also noteworthy that we chose not to have a placebo arm as the effects of psychoactive effects of THC would be difficult to camouflage. Furthermore, we

chose to use oral formulations of cannabinoids so that we could precisely control the dose ingested by participants. The active pharmaceutical ingredients are extracted from the cannabis plant and purified according to pharmaceutical standards (>98%). This level of purity will enable investigators to know that the dosing provided is accurate and quantifiable, and will also enable us to draw conclusions about the efficacy of the active ingredients being studying. When cannabis is smoked or vaped, there is variability in the methods and duration of inhalation used by participants which can influence dosage of cannabis ingested. The oral administration option also removes undesirable pulmonary effects such as symptoms wheezing or breathlessness in addition to inhalation of toxic chemicals⁹⁴. Of special interest to our group is the recent discovery that administration of oral cannabis with lipids leads to high levels of cannabinoids in the intestinal lymphatic system and prominent immunomodulation, as demonstrated by *Zgair et al.*²⁰ This finding is especially important given the prominent role of the mesenteric lymph nodes and gut to HIV persistence⁷. If oral cannabinoids can modify gut microbiome and the enteric immune system favorably, larger clinical trials could be conducted to examine this phenomena in further detail. As gut microbiota differs by sexual orientation, we are attempting to enrol approximately equal numbers of MSM as well as heterosexual individuals⁹⁵ and will describe the demographics of the individuals enrolled in our study in detail. Similarly, we will report participants' current and nadir CD4 T counts as in some studies the enteric bacterial microbiome

of patients with lower CD4 T counts exhibited reduced phylogenetic diversity and richness⁹⁶. There were increases in Enterobacteriaceae, which have been associated with inflammation. Therefore, immunodeficiency in progressive HIV infection is associated with alterations in the enteric virome and bacterial microbiome⁹⁶.

The Canadian government has declared that cannabis' regulatory status will change from being an illegal substance to that of a legal substance in October 2018. Cannabis' change in regulatory status will likely stimulate more discussion amongst patients and physicians and thus physicians need to be informed about the potential risks and benefits of cannabis use. The change in the regulatory landscape will likely also foster more research into cannabis' therapeutic potential. We hope that this study will be a stimulus towards more open discussion between patients and their physicians and that it will reduce stigma associated with cannabinoids use. We also hope that this study will be the cornerstone for future studies investigating the therapeutic benefits of cannabis in PLWHIV and its potential not only at the individual level but also at the population level in the form of harm reduction strategies.

1. Kalayjian RC, Machekano RN, Rizk N, et al. Pretreatment levels of soluble cellular receptors and interleukin-6 are associated with HIV disease progression in subjects treated with highly active antiretroviral therapy. *The Journal of infectious diseases* 2010;201(12):1796-805. doi: 10.1086/652750
2. Funderburg NT, Mayne E, Sieg SF, et al. Increased tissue factor expression on circulating monocytes in chronic HIV infection: relationship to in vivo coagulation and immune activation. *Blood* 2010;115(2):161-7. doi: 10.1182/blood-2009-03-210179
3. The definition of emphysema. Report of a National Heart, Lung, and Blood Institute, Division of Lung Diseases workshop. *The American review of respiratory disease* 1985;132(1):182-5. doi: 10.1164/arrd.1985.132.1.182 [published Online First: 1985/07/01]
4. Brenchley JM, Price DA, Schacker TW, et al. Microbial translocation is a cause of systemic immune activation in chronic HIV infection. *Nature medicine* 2006;12(12):1365-71. doi: 10.1038/nm1511 [published Online First: 2006/11/23]
5. Estes JD, Harris LD, Klatt NR, et al. Damaged intestinal epithelial integrity linked to microbial translocation in pathogenic simian immunodeficiency virus infections. *PLoS pathogens* 2010;6(8):e1001052. doi: 10.1371/journal.ppat.1001052 [published Online First: 2010/09/03]
6. Deeks SG, Tracy R, Douek DC. Systemic effects of inflammation on health during chronic HIV infection. *Immunity* 2013;39(4):633-45. doi: 10.1016/j.immuni.2013.10.001 [published Online First: 2013/10/22]
7. Costiniuk CT, Angel JB. Human immunodeficiency virus and the gastrointestinal immune system: does highly active antiretroviral therapy restore gut immunity? *Mucosal immunology* 2012;5(6):596-604. doi: 10.1038/mi.2012.82 [published Online First: 2012/08/30]
8. Dillon SM, Lee EJ, Donovan AM, et al. Enhancement of HIV-1 infection and intestinal CD4+ T cell depletion ex vivo by gut microbes altered during chronic HIV-1 infection. *Retrovirology* 2016;13:5. doi: 10.1186/s12977-016-0237-1
9. Vujkovic-Cvijin I, Dunham RM, Iwai S, et al. Dysbiosis of the gut microbiota is associated with HIV disease progression and tryptophan catabolism. *Science translational medicine* 2013;5(193):193ra91. doi: 10.1126/scitranslmed.3006438
10. El-Sadr WM, Lundgren J, Neaton JD, et al. CD4+ count-guided interruption of antiretroviral treatment. *The New England journal of medicine* 2006;355(22):2283-96. doi: 10.1056/NEJMoa062360 [published Online First: 2006/12/01]
11. Baker JV, Peng G, Rapkin J, et al. Poor initial CD4+ recovery with antiretroviral therapy prolongs immune depletion and increases risk for AIDS and non-AIDS diseases. *Journal of acquired immune deficiency syndromes* 2008;48(5):541-6. doi: 10.1097/QAI.0b013e31817bebb3

12. Baker JV, Peng G, Rapkin J, et al. CD4+ count and risk of non-AIDS diseases following initial treatment for HIV infection. *Aids* 2008;22(7):841-8. doi: 10.1097/QAD.0b013e3282f7cb76

13. Kim H, Perelson AS. Viral and latent reservoir persistence in HIV-1-infected patients on therapy. *PLoS Comput Biol* 2006;2(10):e135. doi: 10.1371/journal.pcbi.0020135

14. Lu W, Mehraj V, Vyboh K, et al. CD4:CD8 ratio as a frontier marker for clinical outcome, immune dysfunction and viral reservoir size in virologically suppressed HIV-positive patients. *Journal of the International AIDS Society* 2015;18:20052. doi: 10.7448/IAS.18.1.20052

15. Rouzioux C, Richman D. How to best measure HIV reservoirs? *Current opinion in HIV and AIDS* 2013;8(3):170-5. doi: 10.1097/COH.0b013e32835fc619

16. Chun TW, Carruth L, Finzi D, et al. Quantification of latent tissue reservoirs and total body viral load in HIV-1 infection. *Nature* 1997;387(6629):183-8. doi: 10.1038/387183a0 [published Online First: 1997/05/08]

17. Chomont N, El-Far M, Ancuta P, et al. HIV reservoir size and persistence are driven by T cell survival and homeostatic proliferation. *Nature medicine* 2009;15(8):893-900. doi: 10.1038/nm.1972 [published Online First: 2009/06/23]

18. Chomont N, DaFonseca S, Vandergeeten C, et al. Maintenance of CD4+ T-cell memory and HIV persistence: keeping memory, keeping HIV. *Current opinion in HIV and AIDS* 2011;6(1):30-6. doi: 10.1097/COH.0b013e3283413775

19. Zurier RB, Burstein SH. Cannabinoids, inflammation, and fibrosis. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology* 2016 doi: 10.1096/fj.201600646R [published Online First: 2016/07/21]

20. Zgair A, Lee JB, Wong JCM, et al. Oral administration of cannabis with lipids leads to high levels of cannabinoids in the intestinal lymphatic system and prominent immunomodulation. *Sci Rep* 2017;7(1):14542. doi: 10.1038/s41598-017-15026-z

21. Costiniuk CT, Brunet L, Rollet-Kurhajec KC, et al. Tobacco Smoking Is Not Associated With Accelerated Liver Disease in Human Immunodeficiency Virus-Hepatitis C Coinfection: A Longitudinal Cohort Analysis. *Open forum infectious diseases* 2016;3(2):ofw050. doi: 10.1093/ofid/ofw050 [published Online First: 2016/04/06]

22. Abrams DI, Hilton JF, Leiser RJ, et al. Short-term effects of cannabinoids in patients with HIV-1 infection: a randomized, placebo-controlled clinical trial. *Annals of internal medicine* 2003;139(4):258-66.

23. Lee MH, Hancox RJ. Effects of smoking cannabis on lung function. *Expert review of respiratory medicine* 2011;5(4):537-46; quiz 47. doi: 10.1586/ers.11.40 [published Online First: 2011/08/24]

24. Tahamtan A, Tavakoli-Yaraki M, Rygiel TP, et al. Effects of cannabinoids and their receptors on viral infections. *Journal of medical virology* 2016;88(1):1-12. doi: 10.1002/jmv.24292

25. Smith TH, Sim-Selley LJ, Selley DE. Cannabinoid CB1 receptor-interacting proteins: novel targets for central nervous system drug discovery? *Br J Pharmacol* 2010;160(3):454-66. doi: 10.1111/j.1476-5381.2010.00777.x

Protected by copyright, including for uses related to text and data mining, AI training, and similar technologies.
Downloaded from <http://bmjopen.bmj.com/> on June 13, 2025 at Agence Bibliographique de l'Enseignement Supérieur (ABES)

26. DiPatrizio NV. Endocannabinoids in the Gut. *Cannabis Cannabinoid Res* 2016;1(1):67-77. doi: 10.1089/can.2016.0001
27. Muccioli GG, Naslain D, Backhed F, et al. The endocannabinoid system links gut microbiota to adipogenesis. *Mol Syst Biol* 2010;6:392. doi: 10.1038/msb.2010.46
28. Rietschel ET, Kirikae T, Schade FU, et al. Bacterial endotoxin: molecular relationships of structure to activity and function. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology* 1994;8(2):217-25.
29. Yao B, Mackie K. Endocannabinoid receptor pharmacology. *Curr Top Behav Neurosci* 2009;1:37-63. doi: 10.1007/978-3-540-88955-7_2
30. Eisenstein TK, Meissler JJ. Effects of Cannabinoids on T-cell Function and Resistance to Infection. *Journal of neuroimmune pharmacology : the official journal of the Society on NeuroImmune Pharmacology* 2015;10(2):204-16. doi: 10.1007/s11481-015-9603-3
31. Hanus L, Breuer A, Tchilibon S, et al. HU-308: a specific agonist for CB(2), a peripheral cannabinoid receptor. *Proceedings of the National Academy of Sciences of the United States of America* 1999;96(25):14228-33.
32. Russo EB. Cannabidiol Claims and Misconceptions. *Trends Pharmacol Sci* 2017 doi: 10.1016/j.tips.2016.12.004
33. Laprairie RB, Bagher AM, Kelly ME, et al. Cannabidiol is a negative allosteric modulator of the cannabinoid CB1 receptor. *Br J Pharmacol* 2015;172(20):4790-805. doi: 10.1111/bph.13250
34. Ware MA, Wang T, Shapiro S, et al. Cannabis for the Management of Pain: Assessment of Safety Study (COMPASS). *The journal of pain : official journal of the American Pain Society* 2015;16(12):1233-42. doi: 10.1016/j.jpain.2015.07.014
35. Abrams DI, Jay CA, Shade SB, et al. Cannabis in painful HIV-associated sensory neuropathy: a randomized placebo-controlled trial. *Neurology* 2007;68(7):515-21. doi: 10.1212/01.wnl.0000253187.66183.9c
36. Wilsey B, Marcotte T, Tsodikov A, et al. A randomized, placebo-controlled, crossover trial of cannabis cigarettes in neuropathic pain. *The journal of pain : official journal of the American Pain Society* 2008;9(6):506-21. doi: 10.1016/j.jpain.2007.12.010
37. Steffens S, Veillard NR, Arnaud C, et al. Low dose oral cannabinoid therapy reduces progression of atherosclerosis in mice. *Nature* 2005;434(7034):782-6. doi: 10.1038/nature03389
38. Franz CA, Frishman WH. Marijuana Use and Cardiovascular Disease. *Cardiol Rev* 2016;24(4):158-62. doi: 10.1097/CRD.000000000000103
39. Mallat A, Lotersztajn S. Endocannabinoids and liver disease. I. Endocannabinoids and their receptors in the liver. *Am J Physiol Gastrointest Liver Physiol* 2008;294(1):G9-G12. doi: 10.1152/ajpgi.00467.2007
40. Mallat A, Teixeira-Clerc F, Deveaux V, et al. The endocannabinoid system as a key mediator during liver diseases: new insights and therapeutic openings. *Br J Pharmacol* 2011;163(7):1432-40. doi: 10.1111/j.1476-5381.2011.01397.x

41. Teixeira-Clerc F, Belot MP, Manin S, et al. Beneficial paracrine effects of cannabinoid receptor 2 on liver injury and regeneration. *Hepatology* 2010;52(3):1046-59. doi: 10.1002/hep.23779

42. Munoz-Luque J, Ros J, Fernandez-Varo G, et al. Regression of fibrosis after chronic stimulation of cannabinoid CB2 receptor in cirrhotic rats. *The Journal of pharmacology and experimental therapeutics* 2008;324(2):475-83. doi: 10.1124/jpet.107.131896

43. Tam J, Vemuri VK, Liu J, et al. Peripheral CB1 cannabinoid receptor blockade improves cardiometabolic risk in mouse models of obesity. *The Journal of clinical investigation* 2010;120(8):2953-66. doi: 10.1172/JCI42551

44. Cluny NL, Vemuri VK, Chambers AP, et al. A novel peripherally restricted cannabinoid receptor antagonist, AM6545, reduces food intake and body weight, but does not cause malaise, in rodents. *Br J Pharmacol* 2010;161(3):629-42. doi: 10.1111/j.1476-5381.2010.00908.x

45. Costantino CM, Gupta A, Yewdall AW, et al. Cannabinoid receptor 2-mediated attenuation of CXCR4-tropic HIV infection in primary CD4+ T cells. *PloS one* 2012;7(3):e33961. doi: 10.1371/journal.pone.0033961

46. Ramirez SH, Reichenbach NL, Fan S, et al. Attenuation of HIV-1 replication in macrophages by cannabinoid receptor 2 agonists. *Journal of leukocyte biology* 2013;93(5):801-10. doi: 10.1189/jlb.1012523

47. Xu H, Cheng CL, Chen M, et al. Anti-inflammatory property of the cannabinoid receptor-2-selective agonist JWH-133 in a rodent model of autoimmune uveoretinitis. *Journal of leukocyte biology* 2007;82(3):532-41. doi: 10.1189/jlb.0307159

48. Zhang M, Adler MW, Abood ME, et al. CB2 receptor activation attenuates microcirculatory dysfunction during cerebral ischemic/reperfusion injury. *Microvasc Res* 2009;78(1):86-94. doi: 10.1016/j.mvr.2009.03.005

49. Condie R, Herring A, Koh WS, et al. Cannabinoid inhibition of adenylate cyclase-mediated signal transduction and interleukin 2 (IL-2) expression in the murine T-cell line, EL4.IL-2. *The Journal of biological chemistry* 1996;271(22):13175-83.

50. Massi P, Sacerdote P, Ponti W, et al. Immune function alterations in mice tolerant to delta9-tetrahydrocannabinol: functional and biochemical parameters. *J Neuroimmunol* 1998;92(1-2):60-6.

51. Molina PE, Amedee AM, LeCapitaine NJ, et al. Modulation of gut-specific mechanisms by chronic delta(9)-tetrahydrocannabinol administration in male rhesus macaques infected with simian immunodeficiency virus: a systems biology analysis. *AIDS research and human retroviruses* 2014;30(6):567-78. doi: 10.1089/AID.2013.0182

52. Lutge EE, Gray A, Siegfried N. The medical use of cannabis for reducing morbidity and mortality in patients with HIV/AIDS. *Cochrane Database Syst Rev* 2013(4):CD005175. doi: 10.1002/14651858.CD005175.pub3

53. Manuzak JA, Gott TM, Kirkwood JS, et al. Heavy Cannabis Use Associated With Reduction in Activated and Inflammatory Immune Cell Frequencies in Antiretroviral Therapy-Treated Human Immunodeficiency Virus-Infected Individuals. *Clinical infectious diseases : an official publication of the Infectious Diseases Society of America* 2018 doi: 10.1093/cid/cix1116

54. Rizzo MD, Crawford RB, Henriquez JE, et al. HIV-infected cannabis users have lower circulating CD16+ monocytes and IFN-gamma-inducible protein 10 levels compared with nonusing HIV patients. *Aids* 2018;32(4):419-29. doi: 10.1097/QAD.0000000000001704
55. Burstein S. Cannabidiol (CBD) and its analogs: a review of their effects on inflammation. *Bioorg Med Chem* 2015;23(7):1377-85. doi: 10.1016/j.bmc.2015.01.059
56. Bergamaschi MM, Queiroz RH, Zuardi AW, et al. Safety and side effects of cannabidiol, a Cannabis sativa constituent. *Curr Drug Saf* 2011;6(4):237-49.
57. Ribeiro A, Ferraz-de-Paula V, Pinheiro ML, et al. Cannabidiol, a non-psychotropic plant-derived cannabinoid, decreases inflammation in a murine model of acute lung injury: role for the adenosine A(2A) receptor. *Eur J Pharmacol* 2012;678(1-3):78-85. doi: 10.1016/j.ejphar.2011.12.043
58. Burstein SH, Zurier RB. Cannabinoids, endocannabinoids, and related analogs in inflammation. *AAPS J* 2009;11(1):109-19. doi: 10.1208/s12248-009-9084-5
59. Iuvone T, Esposito G, De Filippis D, et al. Cannabidiol: a promising drug for neurodegenerative disorders? *CNS Neurosci Ther* 2009;15(1):65-75. doi: 10.1111/j.1755-5949.2008.00065.x
60. Devinsky O, Patel AD, Cross JH, et al. Effect of Cannabidiol on Drop Seizures in the Lennox-Gastaut Syndrome. *The New England journal of medicine* 2018;378(20):1888-97. doi: 10.1056/NEJMoa1714631
61. Sachse-Seeboth C, Pfeil J, Sehr D, et al. Interindividual variation in the pharmacokinetics of Delta9-tetrahydrocannabinol as related to genetic polymorphisms in CYP2C9. *Clinical pharmacology and therapeutics* 2009;85(3):273-6. doi: 10.1038/clpt.2008.213
62. Jenabian MA, Seddiki N, Yatim A, et al. Regulatory T cells negatively affect IL-2 production of effector T cells through CD39/adenosine pathway in HIV infection. *PLoS pathogens* 2013;9(4):e1003319. doi: 10.1371/journal.ppat.1003319 [published Online First: 2013/05/10]
63. Nikolova M, Carriere M, Jenabian MA, et al. CD39/adenosine pathway is involved in AIDS progression. *PLoS pathogens* 2011;7(7):e1002110. doi: 10.1371/journal.ppat.1002110
64. Vandergeeten C, Fromentin R, Merlini E, et al. Cross-Clade Ultrasensitive PCR-Based Assays To Measure HIV Persistence in Large-Cohort Studies. *Journal of virology* 2014;88(21):12385-96. doi: 10.1128/jvi.00609-14 [published Online First: 2014/08/15]
65. Procopio FA, Fromentin R, Kulpa DA, et al. A Novel Assay to Measure the Magnitude of the Inducible Viral Reservoir in HIV-infected Individuals. *EBioMedicine* 2015;2(8):872-81. doi: 10.1016/j.ebiom.2015.06.019 [published Online First: 2015/10/02]
66. Sharkey M, Triques K, Kuritzkes DR, et al. In vivo evidence for instability of episomal human immunodeficiency virus type 1 cDNA. *Journal of virology* 2005;79(8):5203-10. doi: 10.1128/jvi.79.8.5203-5210.2005 [published Online First: 2005/03/30]
67. Buzon MJ, Massanella M, Llibre JM, et al. HIV-1 replication and immune dynamics are affected by raltegravir intensification of HAART-suppressed subjects. *Nature*

- medicine* 2010;16(4):460-5. doi: 10.1038/nm.2111 [published Online First: 2010/03/17]
68. Nowak P, Trosheid M, Avershina E, et al. Gut microbiota diversity predicts immune status in HIV-1 infection. *Aids* 2015;29(18):2409-18. doi: 10.1097/QAD.0000000000000869
69. Hoel H, Hove-Skovsgaard M, Hov JR, et al. Impact of HIV and Type 2 diabetes on Gut Microbiota Diversity, Tryptophan Catabolism and Endothelial Dysfunction. *Sci Rep* 2018;8(1):6725. doi: 10.1038/s41598-018-25168-3
70. Lebouche B, Jenabian MA, Singer J, et al. The role of extended-release niacin on immune activation and neurocognition in HIV-infected patients treated with antiretroviral therapy - CTN PT006: study protocol for a randomized controlled trial. *Trials* 2014;15:390. doi: 10.1186/1745-6215-15-390
71. Hill AL, Rosenbloom DI, Goldstein E, et al. Real-Time Predictions of Reservoir Size and Rebound Time during Antiretroviral Therapy Interruption Trials for HIV. *PLoS pathogens* 2016;12(4):e1005535. doi: 10.1371/journal.ppat.1005535
72. CONSORT -Pilot and Feasibility Trials. Available at: <http://www.consort-statement.org> Accessed 19 May 2018
73. Chan AW, Tetzlaff JM, Altman DG, et al. SPIRIT 2013 statement: defining standard protocol items for clinical trials. *Annals of internal medicine* 2013;158(3):200-7. doi: 10.7326/0003-4819-158-3-201302050-00583
74. International Committee of Medical Journal Editors: Uniform Requirements for Manuscripts Submitted to Biomedical Journals: Writing and Editing for Biomedical Publication. [\[http://www.icmje.org/recommendations/archives/2008_urm.pdf\]](http://www.icmje.org/recommendations/archives/2008_urm.pdf)
75. Hileman CO, Funderburg NT. Inflammation, Immune Activation, and Antiretroviral Therapy in HIV. *Current HIV/AIDS reports* 2017;14(3):93-100. doi: 10.1007/s11904-017-0356-x
76. Anderson GD, Chan LN. Pharmacokinetic Drug Interactions with Tobacco, Cannabinoids and Smoking Cessation Products. *Clin Pharmacokinet* 2016;55(11):1353-68. doi: 10.1007/s40262-016-0400-9
77. Kosel BW, Aweeka FT, Benowitz NL, et al. The effects of cannabinoids on the pharmacokinetics of indinavir and nelfinavir. *Aids* 2002;16(4):543-50.
78. Mechoulam R, Hanus LO, Pertwee R, et al. Early phytocannabinoid chemistry to endocannabinoids and beyond. *Nat Rev Neurosci* 2014;15(11):757-64. doi: 10.1038/nrn3811
79. Portenoy RK, Ganae-Motan ED, Allende S, et al. Nabiximols for opioid-treated cancer patients with poorly-controlled chronic pain: a randomized, placebo-controlled, graded-dose trial. *The journal of pain : official journal of the American Pain Society* 2012;13(5):438-49. doi: 10.1016/j.jpain.2012.01.003
80. Serpell M, Ratcliffe S, Hovorka J, et al. A double-blind, randomized, placebo-controlled, parallel group study of THC/CBD spray in peripheral neuropathic pain treatment. *Eur J Pain* 2014;18(7):999-1012. doi: 10.1002/j.1532-2149.2013.00445.x
81. Pertwee RG. Emerging strategies for exploiting cannabinoid receptor agonists as medicines. *Br J Pharmacol* 2009;156(3):397-411. doi: 10.1111/j.1476-5381.2008.00048.x

82. Tramer MR, Carroll D, Campbell FA, et al. Cannabinoids for control of chemotherapy induced nausea and vomiting: quantitative systematic review. *BMJ (Clinical research ed)* 2001;323(7303):16-21.
83. Curran HV, Freeman TP, Mokrysz C, et al. Keep off the grass? Cannabis, cognition and addiction. *Nat Rev Neurosci* 2016;17(5):293-306. doi: 10.1038/nrn.2016.28
84. Notcutt W, Langford R, Davies P, et al. A placebo-controlled, parallel-group, randomized withdrawal study of subjects with symptoms of spasticity due to multiple sclerosis who are receiving long-term Sativex(R) (nabiximols). *Mult Scler* 2012;18(2):219-28. doi: 10.1177/1352458511419700
85. Wade DT, Makela PM, House H, et al. Long-term use of a cannabis-based medicine in the treatment of spasticity and other symptoms in multiple sclerosis. *Mult Scler* 2006;12(5):639-45. doi: 10.1177/1352458505070618
86. Robson P. Abuse potential and psychoactive effects of delta-9-tetrahydrocannabinol and cannabidiol oromucosal spray (Sativex), a new cannabinoid medicine. *Expert Opin Drug Saf* 2011;10(5):675-85. doi: 10.1517/14740338.2011.575778
87. GW Pharma. SATIVEX—Product Monograph March 31 2015
88. Zuardi AW, Crippa JA, Hallak JE, et al. Cannabidiol for the treatment of psychosis in Parkinson's disease. *J Psychopharmacol* 2009;23(8):979-83. doi: 10.1177/0269881108096519
89. Abbvie Inc. MARINOL—Product Label August 2017
90. Herkenham M, Lynn AB, Little MD, et al. Cannabinoid receptor localization in brain. *Proceedings of the National Academy of Sciences of the United States of America* 1990;87(5):1932-6.
91. Boulassel MR, Mercier F, Gilmore N, et al. Immunophenotypic patterns of CD8+ T cell subsets expressing CD8alphaalpha and IL-7Ralpha in viremic, aviremic and slow progressor HIV-1-infected subjects. *Clinical immunology* 2007;124(2):149-57. doi: 10.1016/j.clim.2007.05.005
92. Mercier F, Boulassel MR, Yassine-Diab B, et al. Persistent human immunodeficiency virus-1 antigenaemia affects the expression of interleukin-7Ralpha on central and effector memory CD4+ and CD8+ T cell subsets. *Clinical and experimental immunology* 2008;152(1):72-80. doi: 10.1111/j.1365-2249.2008.03610.x
93. Deeks SG, Phillips AN. HIV infection, antiretroviral treatment, ageing, and non-AIDS related morbidity. *BMJ (Clinical research ed)* 2009;338:a3172. doi: 10.1136/bmj.a3172 [published Online First: 2009/01/28]
94. Sparacino CM, Hyldborg PA, Hughes TJ. Chemical and biological analysis of marijuana smoke condensate. *NIDA research monograph* 1990;99:121-40. [published Online First: 1990/01/01]
95. Noguera-Julian M, Rocafort M, Guillen Y, et al. Gut Microbiota Linked to Sexual Preference and HIV Infection. *EBioMedicine* 2016;5:135-46. doi: 10.1016/j.ebiom.2016.01.032
96. Monaco CL, Gootenberg DB, Zhao G, et al. Altered Virome and Bacterial Microbiome in Human Immunodeficiency Virus-Associated Acquired Immunodeficiency Syndrome. *Cell Host Microbe* 2016;19(3):311-22. doi: 10.1016/j.chom.2016.02.011

Authors' contributions

CTC conceived and designed the study, drafted the grant and the protocol manuscript, will organize and supervise trial implementation and will be responsible for trial management. She will also be responsible for trial managements, staff training and supervision. MAJ contributed to study design and participated in grant writing. ZS, BL, JPR, JC, J Szabo, MJB and MK will participate in study implementation. MJB provided input on questionnaires while CTC and MAJ provided immunological expertise and NC provided expertise related to the HIV reservoir. SM and EM contributed to study design. CTC, MAJ and NC designed the experiments. J Singer contributed to the statistical analysis plan. CTC, MAJ and NC designed the experiments. All authors participated in refinement of the study methods, critical reviewed the manuscript drafts and approved the final manuscript. The CTN provides regulatory support. CTC and BL are Fonds de recherche du Québec-Santé (FRQ-S) chercheur-boursier-clinicien Junior 1. BL holds a Strategy for Patient-Oriented Research (SPOR) Mentorship Chair in Innovative Clinical Trials. JPR holds the Louis Lowenstein Chair in Hematology and Oncology at McGill University. MAJ is holder of a Tier 2 Canada Research Chair in immunovirology.

Acknowledgements

This clinical trial was reviewed by both the CTN Scientific Advisory Committee and the Community Advisory Committee and is supported by a competitive grant (CTN PT028). We also wish to acknowledge Ms. Dana Nohyek, Ms. Judy Needham, Ms. Jacqueline Sas and

the Community Advisory Committee of the CTN for ongoing support. In addition, we thank Ms Hansi Peiris and Jonathan Roger of the CVIS for logistical and administrative assistance. Furthermore, we wish to acknowledge Tilray and notably Philippe Lucas and Catherine Jacobson for interest and support of this study through the provision of cannabinoid capsules and for assistance with the application to Health Canada.

Funding statement: This work is supported by the CIHR Canadian HIV Trials Network.

Competing interests: Tilray Inc. is supplying the study medications free of charge. All elements of the study are being undertaken independently of Tilray Inc. The authors declare there are no conflicts of interests.

Data sharing: There are no data yet to share.



SPIRIT 2013 Checklist: Recommended items to address in a clinical trial protocol and related documents*

Section/item	Item No	Description	Addressed on page number
Administrative information			
Title	1	Descriptive title identifying the study design, population, interventions, and, if applicable, trial acronym	1
Trial registration	2a	Trial identifier and registry name. If not yet registered, name of intended registry	Abstract, 4
	2b	All items from the World Health Organization Trial Registration Data Set	3,4
Protocol version	3	Date and version identifier	Abstract /cover letter
Funding	4	Sources and types of financial, material, and other support	54,55
Roles and responsibilities	5a	Names, affiliations, and roles of protocol contributors	1, 54,55
	5b	Name and contact information for the trial sponsor	1
	5c	Role of study sponsor and funders, if any, in study design; collection, management, analysis, and interpretation of data; writing of the report; and the decision to submit the report for publication, including whether they will have ultimate authority over any of these activities	38
	5d	Composition, roles, and responsibilities of the coordinating centre, steering committee, endpoint adjudication committee, data management team, and other individuals or groups overseeing the trial, if applicable (see Item 21a for data monitoring committee)	30,36,37,cover letter

Introduction

Background and rationale	6a	Description of research question and justification for undertaking the trial, including summary of relevant studies (published and unpublished) examining benefits and harms for each intervention	6,7,11-14
	6b	Explanation for choice of comparators	14,19,20
Objectives	7	Specific objectives or hypotheses	28,29
Trial design	8	Description of trial design including type of trial (eg, parallel group, crossover, factorial, single group), allocation ratio, and framework (eg, superiority, equivalence, noninferiority, exploratory)	Abstract, 15

Methods: Participants, interventions, and outcomes

Study setting	9	Description of study settings (eg, community clinic, academic hospital) and list of countries where data will be collected. Reference to where list of study sites can be obtained	15
Eligibility criteria	10	Inclusion and exclusion criteria for participants. If applicable, eligibility criteria for study centres and individuals who will perform the interventions (eg, surgeons, psychotherapists)	16,17
Interventions	11a	Interventions for each group with sufficient detail to allow replication, including how and when they will be administered	19
	11b	Criteria for discontinuing or modifying allocated interventions for a given trial participant (eg, drug dose change in response to harms, participant request, or improving/worsening disease)	30
	11c	Strategies to improve adherence to intervention protocols, and any procedures for monitoring adherence (eg, drug tablet return, laboratory tests)	25,26
	11d	Relevant concomitant care and interventions that are permitted or prohibited during the trial	15,17-19
Outcomes	12	Primary, secondary, and other outcomes, including the specific measurement variables (eg, systolic blood pressure), analysis metric (eg, change from baseline, final value, time to event), method of aggregation (eg, median, proportion), and time point for each outcome. Explanation of the clinical relevance of chosen efficacy and harm outcomes is strongly recommended	34,35
Participant timeline	13	Time schedule of enrolment, interventions (including any run-ins and washouts), assessments, and visits for participants. A schematic diagram is highly recommended (see Figure)	Table 3

1	Sample size	14	Estimated number of participants needed to achieve study objectives and how it was determined, including clinical and statistical assumptions supporting any sample size calculation	34,35
2				
3				
4	Recruitment	15	Strategies for achieving adequate participant enrolment to reach target sample size	Cover letter, already advertised by Canadian HIV Trials Network newsletter and website, meetings involving community; 16
5				
6				
7				
8				
9				
10				
11				
12				
13				
14				
15				
16				
17				

18 **Methods: Assignment of interventions (for controlled trials)**

19 Allocation:

21	Sequence generation	16a	Method of generating the allocation sequence (eg, computer-generated random numbers), and list of any factors for stratification. To reduce predictability of a random sequence, details of any planned restriction (eg, blocking) should be provided in a separate document that is unavailable to those who enrol participants or assign interventions	22
22				
23				
24				
25				
26				
27	Allocation concealment mechanism	16b	Mechanism of implementing the allocation sequence (eg, central telephone; sequentially numbered, opaque, sealed envelopes), describing any steps to conceal the sequence until interventions are assigned	22
28				
29				
30				
31				
32	Implementation	16c	Who will generate the allocation sequence, who will enrol participants, and who will assign participants to interventions	22
33				
34				
35	Blinding (masking)	17a	Who will be blinded after assignment to interventions (eg, trial participants, care providers, outcome assessors, data analysts), and how	22 (non-blind)
36				
37				
38		17b	If blinded, circumstances under which unblinding is permissible, and procedure for revealing a participant's allocated intervention during the trial	22 (non-blind)
39				
40				
41				

42 **Methods: Data collection, management, and analysis**

1	Data collection methods	18a	Plans for assessment and collection of outcome, baseline, and other trial data, including any related processes to promote data quality (eg, duplicate measurements, training of assessors) and a description of study instruments (eg, questionnaires, laboratory tests) along with their reliability and validity, if known. Reference to where data collection forms can be found, if not in the protocol	22-24
6		18b	Plans to promote participant retention and complete follow-up, including list of any outcome data to be collected for participants who discontinue or deviate from intervention protocols	22-24
10	Data management	19	Plans for data entry, coding, security, and storage, including any related processes to promote data quality (eg, double data entry; range checks for data values). Reference to where details of data management procedures can be found, if not in the protocol	36,37
14	Statistical methods	20a	Statistical methods for analysing primary and secondary outcomes. Reference to where other details of the statistical analysis plan can be found, if not in the protocol	34,35
17		20b	Methods for any additional analyses (eg, subgroup and adjusted analyses)	34,35
19		20c	Definition of analysis population relating to protocol non-adherence (eg, as randomised analysis), and any statistical methods to handle missing data (eg, multiple imputation)	N/A
23	Methods: Monitoring			
25	Data monitoring	21a	Composition of data monitoring committee (DMC); summary of its role and reporting structure; statement of whether it is independent from the sponsor and competing interests; and reference to where further details about its charter can be found, if not in the protocol. Alternatively, an explanation of why a DMC is not needed	Cover letter, 29,30,36,37
31		21b	Description of any interim analyses and stopping guidelines, including who will have access to these interim results and make the final decision to terminate the trial	Health Canada, Canadian HIV Trials Network; 29, 30, 36
38	Harms	22	Plans for collecting, assessing, reporting, and managing solicited and spontaneous reported adverse events and other unintended effects of trial interventions or trial conduct	Health Canada, CTN; 29, 30, 36
41	Auditing	23	Frequency and procedures for auditing trial conduct, if any, and whether the process will be independent from investigators and the sponsor	36

1 Ethics and dissemination				
2				
3	Research ethics	24	Plans for seeking research ethics committee/institutional review board (REC/IRB) approval	38
4	approval			
5				
6	Protocol	25	Plans for communicating important protocol modifications (eg, changes to eligibility criteria,	38
7	amendments		outcomes, analyses) to relevant parties (eg, investigators, REC/IRBs, trial participants, trial	
8			registries, journals, regulators)	
9				
10				
11	Consent or assent	26a	Who will obtain informed consent or assent from potential trial participants or authorised surrogates,	38
12			and how (see Item 32)	
13				
14		26b	Additional consent provisions for collection and use of participant data and biological specimens in	N/A at this time
15			ancillary studies, if applicable	
16				
17	Confidentiality	27	How personal information about potential and enrolled participants will be collected, shared, and	36,37
18			maintained in order to protect confidentiality before, during, and after the trial	
19				
20	Declaration of	28	Financial and other competing interests for principal investigators for the overall trial and each study	57
21	interests		site	
22				
23	Access to data	29	Statement of who will have access to the final trial dataset, and disclosure of contractual agreements	35,36
24			that limit such access for investigators	
25				
26				
27	Ancillary and post-	30	Provisions, if any, for ancillary and post-trial care, and for compensation to those who suffer harm	30
28	trial care		from trial participation	
29				
30	Dissemination	31a	Plans for investigators and sponsor to communicate trial results to participants, healthcare	38
31	policy		professionals, the public, and other relevant groups (eg, via publication, reporting in results	
32			databases, or other data sharing arrangements), including any publication restrictions	
33				
34		31b	Authorship eligibility guidelines and any intended use of professional writers	38
35				
36		31c	Plans, if any, for granting public access to the full protocol, participant-level dataset, and statistical	Not determined
37			code	at this time
38				

40 **Appendices**

Informed consent materials	32	Model consent form and other related documentation given to participants and authorised surrogates	Available upon request
Biological specimens	33	Plans for collection, laboratory evaluation, and storage of biological specimens for genetic or molecular analysis in the current trial and for future use in ancillary studies, if applicable	37

*It is strongly recommended that this checklist be read in conjunction with the SPIRIT 2013 Explanation & Elaboration for important clarification on the items. Amendments to the protocol should be tracked and dated. The SPIRIT checklist is copyrighted by the SPIRIT Group under the Creative Commons "[Attribution-NonCommercial-NoDerivs 3.0 Unported](http://creativecommons.org/licenses/by-nc-nd/3.0/)" license.