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Possible role of IL-6 and TIE2 gene polymorphisms in predicting the initial high transport status in peritoneal dialysis patients: an observational study Li Ding^{1,#}, Xinghua Shao^{1,#}, Liou Cao¹, Wei Fang¹, Hao Yan¹, Jiaying Huang¹, Aiping Gu¹, Zanzhe Yu¹, Chaojun Qi¹, Xinbei Chang¹, Zhaohui Ni^{1,*} ¹Department of Nephrology, Renji Hospital, School of Medicine, Shanghai Jiao Tong University, Shanghai 200127, China [#]The authors contributed equally to this work

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Running title: Role of IL-6 and TIE2 Gene Polymorphisms

Abstract

Objectives: The aim of this study was to investigate the effect of IL-6 and TIE2 gene polymorphisms on baseline peritoneal transport property.

Design: An observational study

Setting: A hospital.

Participants: This study included 220 continuous ambulatory peritoneal dialysis (PD) patients.

Outcome measures: Patients were divided into two groups based on the results of an initial peritoneal equilibration test performed within 3 months of starting PD therapy: group 1 consisted of low/low average transporters (n = 123), and group 2 consisted of high/high average transporters (n = 97). We genotyped TIE2 and IL-6 polymorphisms and analysed their effects on baseline transport status.

Results: The genotype AT in IL-6 Rs13306435 and the genotype CC in TIE2 Rs639225 were both negatively associated with a higher initial peritoneal transport status (IL-6 Rs13306435: OR = 0.408, 95% CI, 0.227~0.736; TIE2 Rs639225: OR = 0.188, 95% CI, 0.044~0.806).

Conclusions: IL-6 and TIE2 polymorphisms are associated with baseline peritoneal transport property.

Keywords: peritoneal dialysis, high transport status, interleukin-6, TIE2, gene polymorphism

Article summary Strengths of this study This study was the first study exploring the possible association between TIE2 gene polymorphisms and the characteristics of peritoneal transport. This study confirmed the association between IL-6 polymorphism and baseline peritoneal transport among the Chinese Han population. This study may provide a convenient and non-invasive method to determine transport status before starting dialysis. This study may be helpful in improving the prognosis of ESRD patients. This study may help us elucidate the mechanisms underlying high transport status. Limitations of this study The TA genotype of rs13306435 presents in only 7% of the total population; therefore, it is not the main determinant of peritoneal transport in most patients. This study was a single center research. This study needs to be improved by increasing the sample size.

We did not examine dialysate IL-6/TIE2 concentration to investigate its relationship with SNPs.

We found no statistically significant difference in the data of D/P creatinine and D/D0 glucose among different genotypes.

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Peritoneal dialysis (PD) is an effective renal replacement therapy for end stage renal disease (ESRD) patients¹. Patients undergoing peritoneal dialysis have significantly different small solute transport rates. The standard peritoneal equilibration test (PET) proposed by Twardowski in 1987 is the most widely used method to assess the peritoneal small solute transport rate². Patients can be divided into four types: high (H), high-average (HA), low-average (LA), and low (L) based on PET results. Studies have shown an association between high transport status and poor outcome³⁻⁶. Results of meta-analysis⁷ have indicated that for every 0.1 increase in the dialysate over plasma ratio for creatinine (D/P Cr), the relative risks for mortality and technique failure increase by 1.15 and 1.18, respectively. Compared with that of the low transport group, the mortality of the low-average, high-average, and high transport groups increased by 21.9%, 45.7%, and 77.3%, respectively. As technology advances, new peritoneal dialysate (icodextrin)⁸⁻¹¹ and automated peritoneal dialysis (APD)¹²⁻¹⁴ have been shown to improve the prognosis of high transporters. However, in developing countries, icodextrin and APD cannot be widely used for PD patients. Initial high transport is still an important factor that influences the outcome of these patients without icodextrin or APD. Therefore, it is important to know the baseline peritoneal transport property before starting PD therapy. We can advise probable high transporter patients to choose haemodialysis (HD) or renal transplantation for renal replacement therapy. Researchers have attempted to find non-invasive biomarkers to predict the baseline peritoneal membrane function before starting dialysis. Previous studies found that age, gender, and complications such as hypertension, diabetes, and malnutrition might influence transport characteristics¹. However, they are not sufficient to predict high transport status.

In recent years, many studies have shown that genetic variants may play an important role in mechanisms contributing to the baseline variability in peritoneal transport¹⁵⁻²⁰. It has been

suggested that chronic inflammation mediated by various inflammatory cytokines may have an effect on peritoneal transport²¹. Studies have shown that the IL-6 level in peritoneal dialysates is associated with the peritoneal solute transport rate in dialysis patients²²⁻²⁴. Polymorphism of IL-6 is reported to correlate with baseline peritoneal transport status in Caucasian and Korean patients^{15, 19}. An increase in the effective solute exchange area caused by peritoneal vascular proliferation is also an important factor for high peritoneal transport status²⁵. TIE2 is the receptor of angiogenin (Ang) 1 and 2. Ang/Tie2 has been confirmed to play an important role in angiogenesis in the peritoneum²⁶. The angiogenesis of the peritoneum induced by PD can be inhibited using sTie2/Fc in a uremic rat model²⁷. Therefore, it is possible that the genetic polymorphisms of Il-6 and TIE2 might be involved in the mechanism of high peritoneal transport status. This study aimed to determine whether TIE2 and IL-6 gene polymorphisms have an effect on the baseline peritoneal transport property and explore its possible role in predicting initial high transport status.

Materials and Methods

Patient selection

All PD patients having an initial PET performed within 3 months of starting PD therapy were included. Those who switched from failed renal allograft or maintenance haemodialysis were excluded. Two hundred and twenty continuous ambulatory PD patients in the Peritoneal Dialysis Center, School of Medicine, in Shanghai Jiaotong University were enrolled in the study. The study was approved by the ethical committee of Renji Hospital, School of Medicine, Shanghai Jiao Tong University, Shanghai 200127, China. Written informed consent was obtained from each patient.

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Study of peritoneal transport

A standard PET was performed for each of the enrolled patients. Dialysate as well as plasma

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creatinine and glucose levels were measured at 4 h using 2 L of 2.5% glucose dialysis fluid. Creatinine dialysate to plasma ratios at 4 h (D/P Cr) were calculated. Patients were classified into four types based on the D/P Cr value: high (D/P Cr > 0.8), high average (D/P Cr 0.66 to 0.8), low average (D/P Cr 0.5 to 0.65), and low transporters (D/P Cr < 0.5). Then, they were divided into two groups: group 1 consisted of low/low average transporters, and group 2 consisted of high/high average transporters. The residual urine volume was assessed after 24 h of urine collection. Weekly peritoneal Kt/V (peritoneal Kt/V) and residual urine Kt/V (urine Kt/V) were calculated and presented as total weekly Kt/V (total Kt/V).

DNA extraction and genotyping

DNA was extracted from whole blood using a DNA purification kit (Promega, USA). The SNPs of IL-6 and TIE2 were genotyped by a single base primer extension assay. The genomic DNA flanking the SNP was amplified by polymerase chain reaction (PCR) using forward and reverse primer pairs (Tables 1 and 2), and standard PCR reagents in a 10-µL reaction volume containing 20 ng DNA sample, 0.4 µmol of each primer, 10X PCR buffer, 0.4 µmol dNTPs (Generay Biotech, China), 10 mmol MgCl₂, and 0.25 units HotStarTag DNA Polymerase (QIAGEN, Germany). After 40 cycles of PCR (MJ Research PT-100), the products were purified by 2 U SAP and 2 U Exonuclease I (Epicentre, USA). The purified amplification products (2 µL) were added into a SNaPShot Multiplex Ready reaction mixture (Applied Biosystems, USA) containing 1 μ L of genotyping primer for the primer extension reaction (Table 1, Table 2). The primer extension reaction was carried out with 25 cycles of 96°C for 10 s, 50°C for 5 s, and 60°C for 30 s. The reaction products were purified by 0.5 U SAP. The final reaction samples (0.5 μ L) were added into 9.25 μ L Hi-Di formamide (Applied Biosystems) and 0.25 µL GS-120 LIZ (Applied Biosystems). The mixture was incubated at 95°C for 5 min and then analysed by electrophoresis using the ABI Prism 3730x1 DNA analyser (Applied Biosystems). Results were analysed using GeneScan analysis software

(Applied Biosystems).

Statistical analysis

Statistical analysis was conducted using SPSS version 17.0. All categorical data were presented as absolute counts or percentages, mean and standard deviation (SD) were provided for continuous data. To compare the differences between two baseline transport groups, categorical data were analysed by Fisher's exact test and continuous variables were analysed by an unpaired t-test. Logistic regression analysis was applied to determine whether polymorphism of IL-6 and TIE2 affected the baseline peritoneal transport status. P-values of < 0.05 were considered statistically significant.

Results

Clinical parameters between different transport groups

In total, 220 patients were enrolled in this study. The average age of the patients was 52.54 ± 14.56 years; the male to female ratio was 118:102 and the average body mass index was 21.83 ± 3.47 kg/m². Residual renal function was 3.98 ± 3.47 mL/min. The causes of ESRD were as follows: chronic glomerulonephritis (n = 71; 32.3%), diabetic nephropathy (n = 32; 14.5%), hypertensive nephropathy (n = 9; 4.1%), and other/unknown (n = 107; 48.6%). Based on the first PET results, there were 97 patients (44.1%) in the H/HA group, and 123 patients (55.9%) in the L/LA group. Comparisons of clinical characteristics between the two groups are shown in Table 3.

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Distribution of IL-6 and TIE2 polymorphisms in different transport groups

The distribution of IL-6 and TIE2 genotypes in the peritoneal transport groups are summarized in Tables 4 and 5. Distributions of the 24 alleles (12 polymorphisms) were within the Hardy–Weinberg equilibrium. For the IL-6 polymorphism, there was a statistically

significant correlation between AT genotype of rs13306435 and peritoneal transport group (p = 0.023). For the TIE2 polymorphism, the distribution of rs10967789 and rs639225 genotypes differed significantly between the two groups (p = 0.039 and 0.047, respectively).

Parameters for peritoneal solute transport rate for different genotypes

We further compared the peritoneal solute transport rate among groups with the three SNPs (rs13306435, rs10967789, and rs639225). We found no statistically significant difference in the data of D/P creatinine and D/D0 glucose among different genotypes (Table 6).

Roles of IL-6 and TIE2 gene polymorphisms in predicting initial peritoneal high transport status

With possible clinical factors controlled in a multiple logistic regression model, the genotype AT in IL-6 Rs13306435 and CC in TIE2 Rs639225 were both negatively associated with a higher initial peritoneal transport status (IL-6 Rs13306435: OR=0.408, 95% CI, 0.227~0.736; TIE2 Rs639225: OR = 0.188, 95% CI, 0.044~0.806) (Table 7).

Discussion

The relationship between gene polymorphisms and disease had been gaining greater attention by researchers. It has been shown that SNPs of IL6 may influence the development of cardiovascular disease²⁸, cancer^{29,30}, fractures³¹, and autoimmune diseases³²⁻³⁴. In contrast, there is less available research regarding TIE2 gene polymorphisms, except for a study on the relationship between rs638203/ rs639225 and vascular malformations³⁵.

In this study, we investigated the effect of genetic polymorphisms of IL-6 and TIE2 on the baseline peritoneal transport property. Results showed that IL-6 and TIE2 gene

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polymorphisms were both negatively associated with initial high transport status. The genotypes of rs13306435 and rs639225 were shown to be independent predictors of initial high transport status in PD patients.

Initial transport status can determine the patients' dialysis prescription, which may influence the outcome for PD patients. Previous studies have shown an association between initial high transport status and poor outcome. Although icodextrin and APD have been shown to improve the prognosis of high transporters, most of the PD patients in developing countries are unable to use them. In China, for instance, icodextrin has not been approved for sale yet, and few patients can afford the cost of APD therapy. Therefore, predicting the baseline transport status is important to select a better treatment strategy. Our study provided a potential solution to predict initial high transport status before beginning PD.

There have been several genetic studies of peritoneal solute transport rate in PD patients. Polymorphisms of endothelial nitric oxide synthase¹⁶, receptor of advanced glycation end products¹⁷, and transforming growth factor $-\beta l^{18}$ were reported to be involved in baseline transport status. In 2005, Gillerot et al. showed that the SNP of IL-6 (rs1800795) influenced baseline peritoneal permeability in Caucasian PD patients¹⁵. Additionally, Hwang et al. reported that the rs1800795 polymorphism was associated with dialysate IL-6 concentration and baseline peritoneal transport status in Korean PD patients¹⁹. However, for the Chinese Han population, the minor allele frequency (MAF) of rs1800795 was reported to be very low (MAF = 0.02). Thus, it might not be appropriate to directly apply these results to this population. Our finding similar to those of the aforementioned studies. This supports the role of the IL-6 polymorphism in predicting baseline peritoneal transport.

Angiogenesis in the peritoneum could directly increase the effective solute exchange area and influence the transport characteristics. Previous studies have found that polymorphisms of vascular endothelial growth factor were not associated with initial peritoneal transport type^{15,}

¹⁷. Ang/Tie2 has recently been confirmed to play an important role in angiogenesis in the peritoneum²⁶. Whether the polymorphisms of TIE2 are involved in initial peritoneal transport status remains unknown. In this study, we carefully selected nine SNPs with MAF > 0.05 in the Chinese Han population to investigate their possible role in predicting high transport status. Results showed that TIE2 could be a new factor in predicting the baseline transport type. This study is the first to explore the possible association between TIE2 gene polymorphisms and characteristics of peritoneal transport. Rs639225 is located in exon 13 of TIE2. A study reported its association with venous malformation and presumed that this polymorphism might cause abnormal splicing of *TIE2* into a defective protein²⁸. Functional validation of this polymorphism is warranted in the future.

There are some limitations to our study. The TA genotype of rs13306435 presents in only 7% of the total population; therefore, it is not the main determinant of peritoneal transport in most patients. Additionally, this study was a single centre research and the number of cases was limited; increasing the sample size would improve this study. Research has shown that the IL-6/TIE2 concentration was associated with baseline transport status. We hypothesized that the SNPs may participate in the formation of high transport status by influencing the dialysate IL-6/TIE2 concentration. We did not examine the dialysate IL-6/TIE2 concentration in this study. As shown in Table 6, we did not find any statistically significant difference in the data of D/P creatinine and D/D0 glucose between the different genotypes. We believe this may be due to the limited sample size.

In conclusion, IL-6 and TIE2 polymorphisms are associated with baseline peritoneal transport property. Functional study of the polymorphisms is required in the future.

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Author contributions statement

Li Ding, Xinghua Shao and Zhaohui Ni contributed to conception and design, Li Ding, Xinghua Shao, Liou Cao1, Wei Fang, Hao Yan, Jiaying Huang, Aiping Gu, Zanzhe Yu, Chaojun Qi, Xinbei Chang and Zhaohui Ni contributed to acquisition of data, or analysis and interpretation of data; Li Ding, Xinghua Shao, Liou Cao1, Wei Fang, Hao Yan, Jiaying Huang, Aiping Gu, Zanzhe Yu, Chaojun Qi, Xinbei Chang and Zhaohui Ni contributed to drafting the manuscript or revising it critically for important intellectual content. All authors reviewed the manuscript.

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Competing interests

The authors declare that they have no competing interests.

Data sharing statement

Technical appendix, statistical code and dataset are available from the corresponding author.

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Tables

Table 1 Primer Sequences of IL6 for Genotyping Using SNapShot Assay

SNP	PCR-L	PCR-R	Target
rs18007			TCCCCCTAGTTGTGTCT
95	AACCTCCTCTAAGT	GGTGGGGGCTGATTG	TGC
rs18007 96	GGGCTG	GAAAC	CCAGGCAGTTCTACAA CAGCC
rs13306 435	GAAGGGTCCTACTC AGAGCA	GTTGGGTCAGGGGT GGTTAT	TTCCTTCAGGCAAAGA ATCTAGA
435	AGAGCA	GGTTAT	ATCTAGA



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Table 2 Primer Sequences of TIE2 for	Genotyping Using SNapShot Assay
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TS1096 GAGGAGTATAATGATT 7717 TTTCCTCTGGTGGG GGGCTACTGGGATCT TCCTCAGGC 78578 TAGGAA CTGAC CCACATGGTTGAATTG 67 GCAGA GGTCT ATCACAGC 78592 TTCTTCCTCCTCAA TCACATCAACGTGCT CAATATTGTCCAAGAA 25 CCAGAAA GGTCT ATCACAGC 785429 ACGGGTGGGTCTG GAGGCTTGCCTAAG CATTCTCCTTTGCACAT 13 TTTCTC GAAAT TGC rs3737 GATTGTCCCGAGGT TTTCCCAGGGCACAC TTGTCCCGAGGTCAAG 18 CAAGAG AGTAT AGGCACACCTACTGC 719 TGGCACTGTTTGTC ACCGGCTGACATTC GGCA 718 TTCCAG TAGAG AGTCTGTAGCCTGGG 718 TTCCAG AACCGTACTATCAG AATGCTATTAAATGATT 719 TGGGCATGAAATC ACCGGCTGACAATG GGCA 718 TGGGCTGAAAATC AACCTGTACTATCAG GATGCAGCCTGGGT 718 TGGGGTGACATTG CACTGTGTGT GCAAGA 817 GGAGAC CGTGA GATGAGACGTGAGATAG 817 GGAGAC CGTGA GCAAGA	SNP	PCR-L	PCR-R	Target
rs6578TAGGAACTGACCCACATGGTTTGAATTG GGA67TTCTTCCTCCTCAATCACATCAACGTGCTCAATATTGTCCAAGAA25CCAGAAAGGTCTCAATATTGTCCAAGAA25ACGGGTGGGTCTGGAGGCTTGCCTAAGGCATTCTCCTTTGCACAT13TTTCTCGAAATTTGC13TTGTCCCGAGGTTTTCCCAGGGCACACTTGTCCCGAGGTCAAG18CAAGAGAGTATAGGCACACCCTACTGC719TGGCACTGTTTGTCACCGGCTGACTTTGCGG718TTCCAGAACCTGTACTATCAGAATGCTATTAAATGTTT7789AGAATGCGGTCATTGCACTCCTGGATGAGAGATGAGACGTGAGTAG				
67GGArs6392TTCTTCCTCCTCAATCACATCAACGTGCTCAATATTGTCCAAGAA25CCAGAAAGGTCTCAATATTGTCCAAGAA25ACGGGTGGGTCTGGAGGCTTGCCTAAGGCATTCTCCTTTGCACAT13TTTCTCGAAATTTGCrs3737GATTGTCCCGAGGTTTTCCCAGGGCACACTTGTCCCGAGGTCAAG188CAAGAGAGTATAGGCACACCCTACTGC719TGGCACTGTTTGTCACCGGCTGACTTTGCGGrs2273TTCCAGTAGAGAGTCTGTAGCCCTGGG718rs1096ATGGGCTGAAATCAACCTGTACTATCAGAATGCTATTAAATGTTT7789AGAATGCGGTCATTGCACTCCTGGATGAGAGATGAGACGTGAGTAG	7717			
rs6392TTCTTCCTCCTCAA CCAGAAATCACATCAACGTGCT GGTCTCAATATTGTCCAAGAA ATCACAGC25ACGGGTGGGTCTG TTTCTCGAGGCTTGCCTAAGG GAAATCATTCTCCTTTGCACAT TTGC13TTTCTCGAAGGCAAATTTGCC13TTTCCCGAGGT CAAGAGTTTCCCAGGGCACAC AGTATTTGTCCCGAGGTCAAG AGGTGTA18CAAGAGATTAGGCACACCCTACTGC719TGGCACTGTTTGTC TCCAGACCGGCTGACTTTGC TAGAGGG GCA718TTCCAGAACCTGTACTATCAG GGTCATTGAATGCTATTAAATGTTT TCCTGTGT7789AGAATGCGGTCATTGCACTCCTGGATGAGA78987CTGGGTGACATTGCACTCCTGGATGAGAGATGAGACGTGAGTAG		TAGGAA	CTGAC	CCACATGGTTTGAATTG
25CCAGAAAGGTCTATCACAGCrs5429ACGGGTGGGTCTG TTTCTCGAGGCTTGCCTAAGG GAAATCATTCTCCTTTGCACAT TTGCrs3737GATTGTCCCGAGGT CAAGAGTTTCCCAGGGCACAC AGTATTTGTCCCGAGGTCAAG AGGTGTArs2273TGGCACTGTTTGTC TS2273ACCGGCTGACTTTGC TAGAGGG GGrs2273TTCCAGACCGGCTGACTTTGC TAGAGGG GCArs1096ATGGGCTGAAATC AGAATGCAACCTGTACTATCAG GGTCATTGAATGCTATTAAATGTTT TCCTGTGTrs9987CTGGGTGACATTTGCACTCCTGGATGAGAGATGAGACGTGAGTAG				
rs5429 ACGGGTGGGTCTG GAGGCTTGCCTAAGG CATTCTCCTTTGCACAT 13 TTTCTC GAAAT TTGC rs3737 GATTGTCCCGAGGT TTTCCCAGGGCACAC AGTAGTA 188 CAAGAG AGTAT AGGCACACCAC 19 TGGCACTGTTTGTC ACCGGCTGACTTTGC 719 TGGCACTGTTTGTC ACCGGCTGACTTTGC 719 TGGCACTGTTTGTC TAGAG AGGCCTGACTTTGC 718 TTCCAG AGGCTGAAATC ACCCGGCTGACTTTCG 718 AGAATGC AACCTGTACTATCAG AATGCTATTAAATGTTT 7789 AGAATGC CACTCTGGATGAGA GATGAGACGTGAGTAG				
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rs3737 GATTGTCCCGAGGT TTTCCCAGGGCACAC AGGTGTA 188 CAAGAG AGTAT AGGTGTA rs2273 TGGCACTGTTTGTC ACCGGCTGACTTTGC GG 719 TGGCACTGTTTGTC ACCGGCTGACTTTGC GG rs2273 TTCCAG AGTGAGAC AGGTCTGCCTGGG 718 ATGGGCTGAAATC AACCTGTACTATCAG AGTCTGTAGCCCTGGG 718 GGACTGTAGCCCTGGG 718 CTGGGTGACATTG CACTCTGGATGAGA GATGAGACGTGAGTAG				
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rs2273 719 TGGCACTGTTTGTC ACCGGCTGACTTTGC GG 718 TTCCAG TAGAG ACCGGCTGACTTTGC TAGAGCCCTGGG 718 ATGGGCTGAAATC AACCTGTACTATCAG AATGCTATTAAATGTTT 7789 AGAATGC GGTCATTG AACCTGTACTATCAG 759987 CTGGGTGACATTG CACTCCTGGATGAGA GATGAGACGTGAGTAG				
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Table 3 Comparison of Clinical Characteristics and Peritoneal Parameters between the

 PeritonealTransport Groups of 220 Peritoneal Dialysis Patients

L/LA (n=123)	H/HA (n=97)	Р
52.85±14.94	52.27±14.42	0.761
68(55.3)	60(61.8)	0.291
21.82±3.58	21.85±3.03	0.939
22(17.8)	23(23.7)	0.266
94(68.1)	75(70.1)	0.740
48(39,69)	47(35,68)	0.924
104.60±22.67	99.13±21.29	0.056
36.37±4.7	34.8±5.33	0.017
2.65(0.71,3.83)	2.58(1,3.86)	0.657
64(52)	53(54.6)	0.624
0.44 ± 0.08	0.34 ± 0.08	< 0.01
0.55 ± 0.08	$0.74{\pm}0.07$	< 0.01
4±2.92	3.95±2.63	0.901
1.05±0.6	0.96±0.26	
160(-200,520)	80(-45,500)	0.004
1000(560,1400)	1000(500,1500)	0.217
2.23±0.58	2.20±0.56	0.647
	52.85 ± 14.94 $68(55.3)$ 21.82 ± 3.58 $22(17.8)$ $94(68.1)$ $48(39,69)$ 104.60 ± 22.67 36.37 ± 4.7 $2.65(0.71,3.83)$ $64(52)$ 0.44 ± 0.08 0.55 ± 0.08 4 ± 2.92 1.05 ± 0.6 $160(-200,520)$ $1000(560,1400)$	52.85 ± 14.94 52.27 ± 14.42 $68(55.3)$ $60(61.8)$ 21.82 ± 3.58 21.85 ± 3.03 $22(17.8)$ $23(23.7)$ $94(68.1)$ $75(70.1)$ $48(39,69)$ $47(35,68)$ 104.60 ± 22.67 99.13 ± 21.29 36.37 ± 4.7 34.8 ± 5.33 $2.65(0.71,3.83)$ $2.58(1,3.86)$ $64(52)$ $53(54.6)$ 0.4 ± 0.08 0.74 ± 0.07 4 ± 2.92 3.95 ± 2.63 1.05 ± 0.6 0.96 ± 0.26 $160(-200,520)$ $80(-45,500)$ $1000(560,1400)$ $1000(500,1500)$

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Rs13306435	L/LA (n=123)	H/HA (n=97)	Р
AT	14	3	0.023
TT	109	94	
Rs1800796			
CC	77	61	0.986
CG	37	28	
GG	9	8	
Rs1800795			
GG	121	96	0.506
GC	2	1	

Table 4 II -6 Gene Polymorphisms in Two (

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 Table 5 TIE2 Gene Polymorphisms in Two Groups

	L/LA (n=123)	H/HA (n=97)	Р
Rs10967717			
AA	15	16	0.652
AG	56	47	
GG	52	34	
Rs10967789			
CC	90	83	0.039
CG	33	14	
Rs2273718			
AG	14	4	0.081
GG	109	93	
Rs2273719			
AA	4	2	0.868
AG	36	27	
GG	83	68	
Rs3737188			
CC	6	1	0.373
СТ	44	31	
ГТ	73	65	
Rs542913			
CC	100	80	0.403
СТ	22	17	
ГТ	1	0	
Rs639225			
CC	40	18	0.047
СТ	53	46	
ГТ	30	33	
Rs657867			
AA	112	87	0.732
AG	11	10	
Rs9987817			
CC	97	74	0.730
СТ	22	22	
ГТ	4	1	

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IL-6 Rs13306435	AT	TT	Р
D/P creatinine	0.61 ± 0.99	0.63 ± 0.12	0.357
D/D0 glucose	0.41 ± 0.092	0.39 ± 0.085	0.424
ГІЕ2 Rs639225	CC	CT/TT	Р
D/P creatinine	0.61 ± 0.11	0.64 ± 0.11	0.056
D/D0 glucose	0.40 ± 0.09	0.39 ± 0.08	0.518
ГІЕ2 Rs10967789	CC	CG	Р
D/P creatinine	0.64 ± 0.12	0.61 ± 0.09	0.187
D/D0 glucose	0.39 ± 0.09	0.40 ± 0.06	0.839

Table 6 The parameters for peritoneal solute transport rate according to different genotypes

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Table 7 Multivariate Logistic Regression Model to Identify Factors Associated withHigh/High-Average Transport Status

	OR	95%CI	Р
TIE2 Rs639225 (CC vs. CT/TT)	0.188	0.044~0.806	0.024
IL-6 Rs13306435 (AT vs. TT)	0.408	0.227~0.736	0.043
Age	0.966	0.930~1.004	0.081
Male	1.401	0.519~3.788	0.506
DM	3.28	0.952~11.360	0.060
Periods between operation and initial PET(d)	0.996	0.987~1.005	0.401
hs-CRP(mg/L)	1.081	0.964~1.212	0.182
Serum albumin (g/L)	0.898	0.796~1.014	0.083
TIE2 Rs10967789 (CC vs. CG)	1.061	0.371~1.632	0.197
Hemoglobin (g/L)	0.984	0.796~1.014	0.192

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Section & Topic	No	Item	Reported on pa
TITLE OR ABSTRACT			
	1	Identification as a study of diagnostic accuracy using at least one measure of accuracy	2
		(such as sensitivity, specificity, predictive values, or AUC)	
ABSTRACT			
	2	Structured summary of study design, methods, results, and conclusions	2
		(for specific guidance, see STARD for Abstracts)	
INTRODUCTION			
	3	Scientific and clinical background, including the intended use and clinical role of the index test	4-5
	4	Study objectives and hypotheses	4-5
METHODS	-		
Study design	5	Whether data collection was planned before the index test and reference standard	5
		were performed (prospective study) or after (retrospective study)	
Participants	6	Eligibility criteria	5
	7	On what basis potentially eligible participants were identified	5
		(such as symptoms, results from previous tests, inclusion in registry)	
	8	Where and when potentially eligible participants were identified (setting, location and dates)	5
	9	Whether participants formed a consecutive, random or convenience series	5
Test methods	10a	Index test, in sufficient detail to allow replication	6
	10b	Reference standard, in sufficient detail to allow replication	6
	11	Rationale for choosing the reference standard (if alternatives exist)	6
	12a	Definition of and rationale for test positivity cut-offs or result categories	6
		of the index test, distinguishing pre-specified from exploratory	
	12b	Definition of and rationale for test positivity cut-offs or result categories	6
		of the reference standard, distinguishing pre-specified from exploratory	
	13a	Whether clinical information and reference standard results were available	6
		to the performers/readers of the index test	
	13b	Whether clinical information and index test results were available	6
		to the assessors of the reference standard	
Analysis	14	Methods for estimating or comparing measures of diagnostic accuracy	7
	15	How indeterminate index test or reference standard results were handled	7
	16	How missing data on the index test and reference standard were handled	6
	17	Any analyses of variability in diagnostic accuracy, distinguishing pre-specified from exploratory	7
	18	Intended sample size and how it was determined	6
RESULTS			
Participants	19	Flow of participants, using a diagram	
	20	Baseline demographic and clinical characteristics of participants	7,18
	21a	Distribution of severity of disease in those with the target condition	7,18
	21b	Distribution of alternative diagnoses in those without the target condition	7,18
	22	Time interval and any clinical interventions between index test and reference standard	18
Test results	23	Cross tabulation of the index test results (or their distribution)	19,20
		by the results of the reference standard	
	24	Estimates of diagnostic accuracy and their precision (such as 95% confidence intervals)	22
	25	Any adverse events from performing the index test or the reference standard	8
DISCUSSION			-
	26	Study limitations, including sources of potential bias, statistical uncertainty, and generalisability	3
	27	Implications for practice, including the intended use and clinical role of the index test	5
OTHER			
INFORMATION			_
	28	Registration number and name of registry	-
	29	Where the full study protocol can be accessed	
	30	Sources of funding and other support; role of funders	11

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Possible role of IL-6 and TIE2 gene polymorphisms in predicting the initial high transport status in peritoneal dialysis patients: an observational study

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Primary Subject Heading :	Urology
Secondary Subject Heading:	Urology, Renal medicine
Keywords:	peritoneal dialysis, high transport status, interleukin-6, TIE2, gene polymorphism

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Possible role of IL-6 and TIE2 gene polymorphisms in predicting the initial high transport status in peritoneal dialysis patients: an observational study Li Ding^{1,#}, Xinghua Shao^{1,#}, Liou Cao¹, Wei Fang¹, Hao Yan¹, Jiaying Huang¹, Aiping Gu¹, Zanzhe Yu¹, Chaojun Qi¹, Xinbei Chang¹, Zhaohui Ni^{1,*} ¹Department of Nephrology, Renji Hospital, School of Medicine, Shanghai Jiao Tong University, Shanghai 200127, China [#]The authors contributed equally to this work

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Running title: Role of IL-6 and TIE2 Gene Polymorphisms

Abstract

Objectives: The aim of this study was to investigate the effect of IL-6 and TIE2 gene polymorphisms on baseline peritoneal transport property.

Design: An observational study

Setting: Renji Hospital in Shanghai, China

Participants: This study included 220 continuous ambulatory peritoneal dialysis (PD) patients.

Outcome measures: Patients were divided into two groups based on the results of an initial peritoneal equilibration test performed within 3 months of starting PD therapy: group 1 consisted of low/low average transporters (n = 123), and group 2 consisted of high/high average transporters (n = 97). We genotyped TIE2 and IL-6 polymorphisms and analysed their effects on baseline transport status.

Results: The genotype AT in IL-6 Rs13306435 and the genotype CC in TIE2 Rs639225 were both negatively associated with a higher initial peritoneal transport status (IL-6 Rs13306435: OR = 0.408, 95% CI, 0.227~0.736; TIE2 Rs639225: OR = 0.188, 95% CI, 0.044~0.806).

Conclusions: IL-6 and TIE2 polymorphisms are associated with baseline peritoneal transport property.

Keywords: peritoneal dialysis, high transport status, interleukin-6, TIE2, gene polymorphism

Article summary

Strengths and limitations of this study

This study was the first study exploring the possible association between TIE2 gene polymorphisms and the characteristics of peritoneal transport.

This study was also the first study confirming the association between IL-6 polymorphism and baseline peritoneal transport among the Chinese Han population.

We used a convenient and non-invasive method to study the initial high transport status in PD patients.

The TA genotype of rs13306435 presents in only 7% of the total population, therefore, it is not the main determinant of peritoneal transport in most patients.

We did not examine dialysate IL-6/TIE2 concentration to investigate its relationship with SNPs.

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Introduction

Peritoneal dialysis (PD) is an effective renal replacement therapy for end stage renal disease (ESRD) patients¹. Patients undergoing peritoneal dialysis have significantly different small solute transport rates. The standard peritoneal equilibration test (PET) proposed by Twardowski in 1987 is the most widely used method to assess the peritoneal small solute transport rate². Patients can be divided into four types: high (H), high-average (HA), low-average (LA), and low (L) based on PET results. Studies have shown an association between high transport status and poor outcome³⁻⁶. The results of a meta-analysis⁷ have indicated that for every 0.1 increase in the dialysate over plasma ratio for creatinine (D/P Cr), the relative risks for mortality and technique failure increase by 1.15 and 1.18, respectively. Compared with that of the low transport group, the mortality of the low-average, high-average, and high transport groups increased by 21.9%, 45.7%, and 77.3%, respectively. As technology advances, new peritoneal dialysate (icodextrin)⁸⁻¹¹ and automated peritoneal dialysis (APD)¹²⁻¹⁴ have been shown to improve the prognosis of high transporters. However, in developing countries, icodextrin and APD cannot be widely used for PD patients. Initial high transport is still an important factor that influences the outcome of these patients without icodextrin or APD. Therefore, it is important to know the baseline peritoneal transport property before starting PD therapy. We can advise probable high transporter patients to choose haemodialysis (HD) or renal transplantation for renal replacement therapy. Researchers have attempted to find non-invasive biomarkers to predict the baseline peritoneal membrane function before starting dialysis. Previous studies found that age, gender, and complications such as hypertension, diabetes, and malnutrition might influence transport characteristics¹. However, they are not sufficient to predict high transport status.

In recent years, many studies have shown that genetic variants may play an important role in mechanisms contributing to the baseline variability in peritoneal transport¹⁵⁻²⁰. It has been

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suggested that chronic inflammation mediated by various inflammatory cytokines may have an effect on peritoneal transport²¹. Studies have shown that the IL-6 level in peritoneal dialysates is associated with the peritoneal solute transport rate in dialysis patients²²⁻²⁴. A polymorphism of IL-6 (Rs13306435) is reported to correlate with baseline peritoneal transport status in Caucasian and Korean patients^{15, 19}. An increase in the effective solute exchange area caused by peritoneal vascular proliferation is also an important factor for high peritoneal transport status²⁵. TIE2 is the receptor of angiogenin (Ang) 1 and 2. Ang/Tie2 has been confirmed to play an important role in angiogenesis in the peritoneum²⁶. The angiogenesis of the peritoneum induced by PD can be inhibited using sTie2/Fc in a uremic rat model²⁷. Therefore, it is possible that the genetic polymorphisms of Il-6 and TIE2 might be involved in the mechanism of high peritoneal transport status. This study aimed to determine whether TIE2 and IL-6 gene polymorphisms have an effect on the baseline peritoneal transport property and explore its possible role in predicting initial high transport status.

Materials and Methods

Patient selection

All PD patients having an initial PET performed within 3 months of starting PD therapy were included. Those who switched from failed renal allograft or maintenance haemodialysis were excluded. Two hundred and twenty continuous ambulatory PD patients in the Peritoneal Dialysis Center, School of Medicine, in Shanghai Jiaotong University were enrolled in the study. The study was approved by the ethical committee of Renji Hospital, School of Medicine, Shanghai Jiao Tong University, Shanghai 200127, China. Written informed consent was obtained from each patient.

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Study of peritoneal transport

A standard PET was performed for each of the enrolled patients. Dialysate as well as plasma

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creatinine and glucose levels were measured at 4 h using 2 L of 2.5% glucose dialysis fluid. Creatinine dialysate to plasma ratios at 4 h (D/P Cr) were calculated. Patients were classified into four types based on the D/P Cr value: high (D/P Cr > 0.8), high average (D/P Cr 0.66 to 0.8), low average (D/P Cr 0.5 to 0.65), and low transporters (D/P Cr < 0.5). Then, they were divided into two groups: group 1 consisted of low/low average transporters, and group 2 consisted of high/high average transporters. The residual urine volume was assessed after 24 h of urine collection. Weekly peritoneal Kt/V (peritoneal Kt/V) and residual urine Kt/V (urine Kt/V) were calculated and presented as total weekly Kt/V (total Kt/V).

DNA extraction and genotyping

DNA was extracted from whole blood using a DNA purification kit (Promega, USA). The SNPs of IL-6 and TIE2 were genotyped by a single base primer extension assay. The genomic DNA flanking the SNP was amplified by polymerase chain reaction (PCR) using forward and reverse primer pairs (Tables 1 and 2), and standard PCR reagents in a 10-µL reaction volume containing 20 ng DNA sample, 0.4 µmol of each primer, 10X PCR buffer, 0.4 µmol dNTPs (Generay Biotech, China), 10 mmol MgCl₂, and 0.25 units HotStarTaq DNA Polymerase (QIAGEN, Germany). After 40 cycles of PCR (MJ Research PT-100), the products were purified by 2 U SAP and 2 U Exonuclease I (Epicentre, USA). The purified amplification products (2 µL) were added into a SNaPShot Multiplex Ready reaction mixture (Applied Biosystems, USA) containing 1 μ L of genotyping primer for the primer extension reaction (Table 1, Table 2). The primer extension reaction was carried out with 25 cycles of 96°C for 10 s, 50°C for 5 s, and 60°C for 30 s. The reaction products were purified by 0.5 U SAP. The final reaction samples (0.5 μ L) were added into 9.25 μ L Hi-Di formamide (Applied Biosystems) and 0.25 µL GS-120 LIZ (Applied Biosystems). The mixture was incubated at 95°C for 5 min and then analysed by electrophoresis using the ABI Prism 3730x1 DNA analyser (Applied Biosystems). Results were analysed using GeneScan analysis software

(Applied Biosystems).

Statistical analysis

Statistical analysis was conducted using SPSS version 17.0. All categorical data were presented as absolute counts or percentages, mean and standard deviation (SD) were provided for continuous data. To compare the differences between two baseline transport groups, categorical data were analysed by Fisher's exact test and continuous variables were analysed by an unpaired t-test. Logistic regression analysis was applied to determine whether polymorphism of IL-6 and TIE2 affected the baseline peritoneal transport status. P-values of < 0.05 were considered statistically significant.

Results

Clinical parameters between different transport groups

In total, 220 patients were enrolled in this study. The average age of the patients was 52.54 ± 14.56 years; the male to female ratio was 118:102 and the average body mass index was 21.83 ± 3.47 kg/m². Residual renal function was 3.98 ± 3.47 mL/min. The causes of ESRD were as follows: chronic glomerulonephritis (n = 71; 32.3%), diabetic nephropathy (n = 32; 14.5%), hypertensive nephropathy (n = 9; 4.1%), and other/unknown (n = 107; 48.6%). Based on the first PET results, there were 97 patients (44.1%) in the H/HA group, and 123 patients (55.9%) in the L/LA group. Comparisons of clinical characteristics between the two groups are shown in Table 3.

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Distribution of IL-6 and TIE2 polymorphisms in different transport groups

The distribution of IL-6 and TIE2 genotypes in the peritoneal transport groups are summarized in Tables 4 and 5. Distributions of the 24 alleles (12 polymorphisms) were within the Hardy–Weinberg equilibrium. For the IL-6 polymorphism, there was a statistically

significant correlation between AT genotype of rs13306435 and peritoneal transport group (p = 0.023). For the TIE2 polymorphism, the distribution of rs10967789 and rs639225 genotypes differed significantly between the two groups (p = 0.039 and 0.047, respectively).

Parameters for peritoneal solute transport rate for different genotypes

We further compared the peritoneal solute transport rate among groups with the three SNPs (rs13306435, rs10967789, and rs639225). We found no statistically significant difference in the data of D/P creatinine and D/D0 glucose among different genotypes (Table 6).

Roles of IL-6 and TIE2 gene polymorphisms in predicting initial peritoneal high transport status

With possible clinical factors controlled in a multiple logistic regression model, the genotype AT in IL-6 Rs13306435 and CC in TIE2 Rs639225 were both negatively associated with a higher initial peritoneal transport status (IL-6 Rs13306435: OR=0.408, 95% CI, 0.227~0.736; TIE2 Rs639225: OR = 0.188, 95% CI, 0.044~0.806) (Table 7).

Discussion

The relationship between gene polymorphisms and disease has been gaining greater attention by researchers. It has been shown that SNPs of IL6 may influence the development of cardiovascular disease²⁸, cancer^{29,30}, fractures³¹, and autoimmune diseases³²⁻³⁴. In contrast, there is less available research regarding TIE2 gene polymorphisms, except for a study on the relationship between rs638203/ rs639225 and vascular malformations³⁵.

In this study, we investigated the effect of genetic polymorphisms of IL-6 and TIE2 on the baseline peritoneal transport property. Results showed that IL-6 and TIE2 gene

polymorphisms were both negatively associated with initial high transport status. The genotypes of rs13306435 and rs639225 were shown to be independent predictors of initial high transport status in PD patients.

Initial transport status can determine the patients' dialysis prescription, which may influence the outcome for PD patients. Previous studies have shown an association between initial high transport status and poor outcome. Although icodextrin and APD have been shown to improve the prognosis of high transporters, most of the PD patients in developing countries are unable to use them. In China, for instance, icodextrin has not been approved for sale yet, and few patients can afford the cost of APD therapy. Therefore, predicting the baseline transport status is important to select a better treatment strategy. Our study provided a potential solution to predict initial high transport status before beginning PD.

There have been several genetic studies of peritoneal solute transport rate in PD patients. Polymorphisms of endothelial nitric oxide synthase¹⁶, receptor of advanced glycation end products¹⁷, and transforming growth factor $-\beta l^{18}$ were reported to be involved in baseline transport status. In 2005, Gillerot et al. showed that the SNP of IL-6 (rs1800795) influenced baseline peritoneal permeability in Caucasian PD patients¹⁵. Additionally, Hwang et al. reported that the rs1800795 polymorphism was associated with dialysate IL-6 concentration and baseline peritoneal transport status in Korean PD patients¹⁹. However, for the Chinese Han population, the minor allele frequency (MAF) of rs1800795 was reported to be very low (MAF = 0.02). Thus, it might not be appropriate to directly apply these results to this population. Our finding similar to those of the aforementioned studies. This supports the role of the IL-6 polymorphism in predicting baseline peritoneal transport.

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Angiogenesis in the peritoneum could directly increase the effective solute exchange area and influence the transport characteristics. Previous studies have found that polymorphisms of vascular endothelial growth factor were not associated with initial peritoneal transport type^{15,}

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¹⁷. Ang/Tie2 has recently been confirmed to play an important role in angiogenesis in the peritoneum²⁶. Whether the polymorphisms of TIE2 are involved in initial peritoneal transport status remains unknown. In this study, we carefully selected nine SNPs with MAF > 0.05 in the Chinese Han population to investigate their possible role in predicting high transport status. Results showed that TIE2 could be a new factor in predicting the baseline transport type. This study is the first to explore the possible association between TIE2 gene polymorphisms and characteristics of peritoneal transport. Rs639225 is located in exon 13 of TIE2. A study reported its association with venous malformation and presumed that this polymorphism might cause abnormal splicing of *TIE2* into a defective protein²⁸. Functional validation of this polymorphism is warranted in the future. There are some limitations to our study. The TA genotype of rs13306435 presents in only 7% of the total population; therefore, it is not the main determinant of peritoneal transport in most patients. Additionally, this study was conducted at a single centre and the number of cases was limited; increasing the sample size would improve this study. Research has shown that the IL-6/TIE2 concentration was associated with baseline transport status. We hypothesized that the SNPs may participate in the formation of high transport status by influencing the dialysate IL-6/TIE2 concentration. We did not examine the dialysate IL-6/TIE2 concentration in this study. As shown in Table 6, we did not find any statistically significant difference in the data of D/P creatinine and D/D0 glucose between the different genotypes. We believe this may be due to the limited sample size.

> In conclusion, IL-6 and TIE2 polymorphisms are associated with baseline peritoneal transport property. Functional study of the polymorphisms is required in the future.

Author contributions statement

Li Ding, Xinghua Shao and Zhaohui Ni contributed to conception and design, Li Ding, Xinghua Shao, Liou Cao1, Wei Fang, Hao Yan, Jiaying Huang, Aiping Gu, Zanzhe Yu, Chaojun Qi, Xinbei Chang and Zhaohui Ni contributed to acquisition of data, or analysis and interpretation of data; Li Ding, Xinghua Shao, Liou Cao1, Wei Fang, Hao Yan, Jiaying Huang, Aiping Gu, Zanzhe Yu, Chaojun Qi, Xinbei Chang and Zhaohui Ni contributed to drafting the manuscript or revising it critically for important intellectual content. All authors reviewed the manuscript.

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Competing interests

The authors declare that they have no competing interests.

Data sharing statement

Technical appendix, statistical code and dataset are available from the corresponding author.

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Tables

Table 1 Primer Sequences of IL6 for Genotyping Using SNapShot Assay

SNP	PCR-L	PCR-R	Target
rs18007 95 rs18007 96	AACCTCCTCTAAGT GGGCTG	GGTGGGGGCTGATTG GAAAC	TCCCCCTAGTTGTGTCT TGC CCAGGCAGTTCTACAA CAGCC
rs13306 435	GAAGGGTCCTACTC AGAGCA	GTTGGGTCAGGGGT GGTTAT	TTCCTTCAGGCAAAGA ATCTAGA



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Table 2 Primer Sequences of TIE2 for	Genotyping Using SNapShot Assay
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SNP	PCR-L	PCR-R	Target
rs1096			GAGGAGTATAATGATT
7717	TTTCCTCTGGTGGG	GGGCTACTGGGATCT	TCCTCAGGC
rs6578	TAGGAA	CTGAC	CCACATGGTTTGAATTG
67			GGA
rs6392	TTCTTCCTCCTCAA	TCACATCAACGTGCT	CAATATTGTCCAAGAA
25	CCAGAAA	GGTCT	ATCACAGC
rs5429	ACGGGTGGGTCTG	GAGGCTTGCCTAAGG	CATTCTCCTTTGCACAT
13	TTTCTC	GAAAT	TTGC
rs3737	GATTGTCCCGAGGT	TTTCCCAGGGCACAC	TTGTCCCGAGGTCAAG
188	CAAGAG	AGTAT	AGGTGTA
rs2273			AGGCACACCCTACTGC
719	TGGCACTGTTTGTC	ACCGGCTGACTTTGC	GG
rs2273	TTCCAG	TAGAG	AGTCTGTAGCCCTGGG
718			GCA
rs1096	ATGGGCTGAAATC	AACCTGTACTATCAG	AATGCTATTAAATGTTT
7789	AGAATGC	GGTCATTG	TCCTGTGT
rs9987	CTGGGTGACATTTG	CACTCCTGGATGAGA	GATGAGACGTGAGTAG
817	GGAGAC	CGTGA	GCAAGA



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Table 3 Comparison of Clinical Characteristics and Peritoneal Parameters between thePeritonealTransport Groups of 220 Peritoneal Dialysis Patients

	L/LA (n=123)	H/HA (n=97)	Р
Age (y)	52.85±14.94	52.27±14.42	0.761
Male (%)	68(55.3)	60(61.8)	0.291
BMI (kg/m²)	21.82±3.58	21.85±3.03	0.939
DM (%)	22(17.8)	23(23.7)	0.266
Hypertension (%)	94(68.1)	75(70.1)	0.740
Periods between operation and initial PET(d)	48(39,69)	47(35,68)	0.924
Hemoglobin (g/L)	104.60±22.67	99.13±21.29	0.056
Serum albumin (g/L)	36.37±4.7	34.8±5.33	0.017
hs-CRP (mg/L)	2.65(0.71,3.83)	2.58(1,3.86)	0.657
ACEI/ARB(%)	64(52)	53(54.6)	0.624
D4/D0 (glu)	0.44 ± 0.08	$0.34{\pm}0.08$	< 0.01
D/P Cr	0.55 ± 0.08	$0.74{\pm}0.07$	< 0.01
RRF(ml/min)	4±2.92	3.95±2.63	0.901
nPCR[g/(kg.d)]	1.05±0.6	0.96±0.26	
UF(ml)	160(-200,520)	80(-45,500)	0.004
Urine volume (ml)	1000(560,1400)	1000(500,1500)	0.217
Kt/V	2.23±0.58	2.20±0.56	0.647

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38 39 40	
41 42 43 44	
45 46 47	
48 49 50	
51 52 53 54	
55 56 57	
58 59 60	

Rs13306435	L/LA (n=123)	H/HA (n=97)	Р
AT	14	3	0.023
TT	109	94	
Rs1800796			
CC	77	61	0.986
CG	37	28	
GG	9	8	
Rs1800795			
GG	121	96	0.506
GC	2	1	

Table 4 IL-6 Gene Polymorphisms in Two Groups

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$\begin{array}{c} 2&3&4&5&6&7\\ 8&9&10&11&2&3&1\\ 1&1&1&1&1&1&2&2&2&2&2&2&2&2&2&2&2&2&2&2$	1 2 3 4
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51 52 53 54 55 56 57 58 59	46 47 48 49
56 57 58 59	51 52 53 54
	56 57 58 59

	L/LA (n=123)	H/HA (n=97)	Р
Rs10967717			
AA	15	16	0.652
AG	56	47	
GG	52	34	
Rs10967789			
CC	90	83	0.039
CG	33	14	
Rs2273718			
AG	14	4	0.081
GG	109	93	
Rs2273719			
AA	4	2	0.868
AG	36	27	
GG	83	68	
Rs3737188			
CC	6	1	0.373
СТ	44	31	
TT	73	65	
Rs542913			
CC	100	80	0.403
СТ	22	17	
TT	1	0	
Rs639225		6	
CC	40	18	0.047
СТ	53	46	
TT	30	33	
Rs657867			
AA	112	87	0.732
AG	11	10	
Rs9987817			
CC	97	74	0.730
СТ	22	22	
TT	4	1	

Table 5 TIE2 Gene Polymorphisms in Two Groups

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D/D0 glucose 0.41 ± 0.092 0.39 ± 0.085 0.424 TIE2 Rs639225CCCT/TTPD/P creatinine 0.61 ± 0.11 0.64 ± 0.11 0.056 D/D0 glucose 0.40 ± 0.09 0.39 ± 0.08 0.518 TIE2 Rs10967789CCCGPD/P creatinine 0.64 ± 0.12 0.61 ± 0.09 0.187	IL-6 Rs13306435	s13306435 AT TT		Р
TE2 Rs639225 CC CT/TT P D/P creatinine 0.61±0.11 0.64±0.11 0.056 D/D0 glucose 0.40±0.09 0.39±0.08 0.518 TE2 Rs10967789 CC CG P D/P creatinine 0.64±0.12 0.61±0.09 0.187 D/P creatinine 0.64±0.12 0.61±0.09 0.835 D/D0 glucose 0.39±0.09 0.40±0.06 0.835	D/P creatinine	0.61 ± 0.99	0.63 ± 0.12	0.357
D/P creatinine 0.61±0.11 0.64±0.11 0.056 D/D0 glucose 0.40±0.09 0.39±0.08 0.518 TE2 Rs10967789 CC CG P D/P creatinine 0.64±0.12 0.61±0.09 0.187 D/D0 glucose 0.39±0.09 0.40±0.06 0.839	D/D0 glucose	0.41 ± 0.092	0.39 ± 0.085	0.424
D/D0 glucose 0.40±0.09 0.39±0.08 0.518 TE2 Rs10967789 CC CG P D/P creatinine 0.64±0.12 0.61±0.09 0.187 D/D0 glucose 0.39±0.09 0.40±0.06 0.839	TIE2 Rs639225	CC	CT/TT	Р
TE2 Rs10967789 CC CG P D/P creatinine 0.64±0.12 0.61±0.09 0.187 D/D0 glucose 0.39±0.09 0.40±0.06 0.839	D/P creatinine	0.61 ± 0.11	0.64 ± 0.11	0.056
D/P creatinine 0.64±0.12 0.61±0.09 0.187 D/D0 glucose 0.39±0.09 0.40±0.06 0.839	D/D0 glucose	0.40 ± 0.09	0.39 ± 0.08	0.518
D/D0 glucose 0.39±0.09 0.40±0.06 0.839	TIE2 Rs10967789	CC	CG	Р
	D/P creatinine	$0.\overline{64 \pm 0.12}$	0.61 ± 0.09	0.187
	D/D0 glucose	0.39 ± 0.09	0.40 ± 0.06	0.839

Table 6 The parameters for peritoneal solute transport rate according to different genotypes

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Table 7 Multivariate Logistic Regression Model to Identify Factors Associated withHigh/High-Average Transport Status

	OR	95%CI	Р
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TIE2 Rs639225 (CC vs. CT/TT)	0.188	0.044~0.806	0.024
IL-6 Rs13306435 (AT vs. TT)	0.408	0.227~0.736	0.043
Age	0.966	0.930~1.004	0.081
Male	1.401	0.519~3.788	0.506
DM	3.28	0.952~11.360	0.060
Periods between operation and initial PET(d)	0.996	0.987~1.005	0.401
hs-CRP(mg/L)	1.081	0.964~1.212	0.182
Serum albumin (g/L)	0.898	0.796~1.014	0.083
TIE2 Rs10967789 (CC vs. CG)	1.061	0.371~1.632	0.197
Hemoglobin (g/L)	0.984	0.796~1.014	0.192