To cite: Huang N. Zhu X. Shu Z.

et al. Association between

elevated serum REG I $\alpha$  levels

and eGFR decline in patients

with chronic kidney disease:

a cross-sectional study in

eastern China. BMJ Open

bmjopen-2024-086874

Prepublication history

and additional supplemental

material for this paper are available online. To view these

files, please visit the journal

bmjopen-2024-086874).

NH, XZ and ZS contributed

Received 27 March 2024

Accepted 08 January 2025

Check for updates

C Author(s) (or their

BMJ Group.

end of article.

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online (https://doi.org/10.1136/

2025;15:e086874. doi:10.1136/

# **BMJ Open** Association between elevated serum **REG I***α* levels and eGFR decline in patients with chronic kidney disease: a cross-sectional study in eastern China

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#### ABSTRACT

Objectives This study aimed to investigate the relationship between serum regenerating protein I $\alpha$  (REG  $I\alpha$ ) levels and estimated glomerular filtration rate (eGFR) and to evaluate the diagnostic efficiency of REG I $\alpha$  in chronic kidnev disease (CKD).

Design This is a cross-sectional study. Setting The study was conducted in eastern China between August 2022 and August 2023.

Participants A total of 880 participants aged over 18 years were enrolled, with 220 non-CKD participants (111 males, 50.45%) and 660 patients with CKD (366 males, 55.45%). CKD was diagnosed based on the Kidney Disease: Improving Global Outcomes (KDIGO) 2012 guidelines. Exclusion criteria included participation in other trials, acute kidney injury, end-stage kidney disease undergoing renal replacement therapy, pregnancy, active infections, gastrointestinal or pancreatic inflammation, history of gastrointestinal or pancreatic resections, cancer and mental disorders.

**Results** Serum REG I $\alpha$  was significantly higher in the CKD group (125.54 (60.28-303.39) ng/mL) compared with those in the non-CKD group (24.62 (14.09-37.32) ng/mL, p<0.001). Positive correlations were observed between serum REG I $\alpha$  and serum creatinine. cvstatin C (Cvs-C). and kidney injury molecule 1 (KIM-1), while a negative correlation was identified with eGFR. After adjusting for sex, diabetes, hypertension and fasting blood glucose, the multivariate regression analysis demonstrated a significant association between serum REG I $\alpha$  and eGFR (OR=1.737 (1.263-2.388), p=0.001). Furthermore, serum REG I $\alpha$  levels increased progressively with declining kidney function categorised by eGFR (p<0.001). In CKD screening, serum REG I $\alpha$  demonstrated strong diagnostic performance, with an area under the receiver operating characteristic curves (AUC) of 0.860 (0.813-0.899), providing a sensitivity of 71.63%, a specificity of 86.89%, a positive predictive value of 94.30% and a negative predictive value of 46.85%. Additionally, serum REG I $\alpha$ exhibited an AUC of 0.769 (0.712-0.819) for identifying high- and very-high-risk CKD based on KDIGO risk stratification. Its sensitivity significantly outperformed serum Cys-C and KIM-1 (82.80% vs 75.16% and 36.94%, respectively).

 The DeLong test was applied to statistically compare areas under the receiver operating characteristic curve values among the biomarkers of CKD, enhancing the reliability of diagnostic performance assessments.
Conclusions This study provided compelling evidence that serum REG lα levels were notably elevated in patients with CKD and closely associated with kidney function. REG lα levels were notably elevated in patients with CKD and closely associated with kidney function.
Conclusions This study provided compelling evidence that serum REG lα levels were notably elevated in patients with CKD and closely associated with kidney function. REG lα and the risk of chronic kidney function.
This is the first study provided compelling evidence that serum REG lα levels were notably elevated in patients with CKD and closely associated with kidney function. REG lα may serve as a promising biomarker for CKD detection and risk stratification.
Conclusions This study provided compelling evidence that serum REG la levels were notably elevated in patients with CKD and closely associated with kidney function. REG Iα may serve as a promising biomarker for CKD detection and risk stratification.
Chronic kidney disease (CKD) encompasses a wide range of underlying etiologies and exhibits variable progression rates, <sup>1–2</sup> and may become the fifth leading cause of death worldwide by 2040.<sup>3</sup> The endpoint of CKD, may become the fifth leading cause of death **3** worldwide by 2040.<sup>3</sup> The endpoint of CKD, known as end-stage kidney disease (ESKD), is characterised by a loss of approximately 90% of kidney function, rendering long-term survival without renal replacement therapy impossible.<sup>2</sup> The high prevalence, low detection rate, severe clinical outcomes and substantial economic burden of CKD underscore its importance as a critical global health

issue.<sup>4</sup> Early prevention, detection and treatment are key to improving patient outcomes and slowing the progression to ESKD.

Current biomarkers such as serum creatinine (Scr), blood urea nitrogen (BUN), estimated glomerular filtration rate (eGFR) and urine albuminuria creatine ratio (UACR) are routinely used to evaluate CKD severity.<sup>5-8</sup> In 2012, the Kidney Disease: Improving Global Outcomes (KDIGO) has taken the eGFR and UACR to create risk stratification strategies, assisting in the tailored management of CKD and guiding treatment for those at a higher risk of progression.<sup>9</sup> Emerging biomarkers, including cystatin C (Cys-C), kidney injury molecule 1 (KIM-1) and \beta2-microglobulin, have demonstrated the potential ability to enhance the precision of CKD screening, either independently or in conjunction with traditional markers.<sup>231011</sup> However, most biomarkers have not yet met clinical expectations in terms of sensitivity, specificity and practicality.<sup>12–15</sup> Few biomarkers are capable of effectively detecting CKD while simultaneously assessing progression risk. Therefore, identifying a novel biomarker that can both monitor kidney function decline and stratify CKD progression risk remains of paramount importance.

Regenerating protein I $\alpha$  (REG I $\alpha$ ), a 16 kDa protein primarily secreted by the pancreas and intestine,<sup>16</sup> is also referred to as pancreatic stone protein (PSP).<sup>17</sup> It plays a vital role in cellular proliferation and regeneration processes.<sup>18</sup><sup>19</sup> Recent studies have reported the presence of REG Ia in patients with various kidney diseases, suggesting its involvement in renal pathology.<sup>20 21</sup> Our previous studies also have further demonstrated that serum REG Ia levels are elevated in patients with diabetic kidney disease (DKD), consistent with the findings of Sobajima and others.<sup>22–24</sup> These observations highlight the potential role of REG Ia as a biomarker for kidney insufficiency.

In this study, we aimed to investigate the relationship between serum REG Ia levels and kidney function, assess its potential as a screening tool for CKD and evaluate its role as a biomarker for kidney function and disease progression.

#### **METHODS**

#### **Study subjects**

Participants were recruited from Zhongda Hospital between August 2022 and August 2023. The study was approved by the ethics committee (approval number: 2022ZDSYLL204-P01), with a clinical study registration of ChiCTR2300072247. Informed consent was acquired from all participants.

The inclusion criteria were as follows: (1) non-CKD participant: age >18 years; (2) patients with CKD: age >18 years and diagnosed with CKD in accordance with the KIDGO 2012 guidelines.<sup>1</sup> The exclusion criteria were as follows: (1) enrolled in another trial; (2) acute kidney injury; (3) patients with ESKD who are undergoing renal replacement therapy; (4) pregnancy; (5) active infection; | from (ABES http://bmjopen.bmj.com/ on June 7, 2025 at Agence Bibliographique de l

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6) acute or chronic inflammation of the gastrointestinal or pancreatic resections; (8) cancer and (9) mental disorders. EGFR was calculated using the Chronic Kidney Disease Fidemiology Collaboration equation.<sup>25</sup> CKD stages were classified based on the U.S. National Kidney Foundation, while risk stratification of CKD progression was performed according to the KDIGO guidelines.<sup>26, 27</sup> Subgroup analyses were conducted among patients with and different degrees of kidney function impairment. A sample size calculation was conducted to ensure sufficient statistical power to detect associations between serum REG Iα and eGFR. The parameters included an expected elevated REG Iα proportion (P1=0.3), a two-sufficient statistical power to detect associations between serum and finite population correction. A total of 880 participants were included to enhance the robustness and entities and finite population correction. A total of 880 participants were included to enhance the robustness and finite population correction. A total of the peripheral blood samples. Blood samples were conflucted at 3500 pm for 15 min, and the upper serum was collected to the study. Baseline demographic data were collected using stam for fisting and took about 3mL of the peripheral blood samples. Blood samples were immediately stored at -80°C for subsequent analyses. The following clinical blood samples. Blood samples were extracted from the clinical laboratory of the Hospital: Scr, BUN, uric acid (UA), Cys-C, myoglobin, UACR and fast blood glucose (FBG). The laboratory implements internal and external quality control procedures directed by a Chinese Quality Control Laboratory. The serum REG Ia levels were determined using a double antibody sandwich ELISA, as previously described.<sup>28</sup> Serum KIM-1 was measured using an ELISA kit (KE00136) from Proteintech.

#### **Statistical analysis**

, and Statistical analyses were conducted using SPSS 20.0, Med-Calc and GraphPad Prism 8.0. Continuous variables were summarised as mean±SD for normally distributed data or as median with IQR for non-normally distributed data. For categorical variables, the frequency with a percentage of each category was calculated. Normality was assessed using graphical methods (Q-Q plots) and the Shapiro-Wilk test. For variables that did not meet normality, **2** we used nonparametric descriptors and methods. We used Student's t-test for normally distributed continuous variables, Mann-Whitney U test for nonnormally distributed continuous variables and  $\chi^2$ or Fisher's exact test for categorical variables for two group comparisons. Tukey's multiple comparison test was employed to examine the differences in biomarker values across three or more groups, thereby avoiding the issue of multiple comparisons.

Spearman's rank correlation analyses and ordinal logistic regression were used to measure the associations between serum REG Ia and other biomarkers of kidney function. Multivariate logistic regression analysis was used to identify the independent factors of kidney dysfunction. The study included age, BUN, UA, serum myoglobin, serum Cys-C, serum KIM-1/100 and serum REG I $\alpha$ /100 into an ordinal multiple logistic regression model, while adjusting for sex, diabetes, hypertension and FBG. The multivariate logistic regression model also incorporates the above covariates. The area under the receiver operating characteristic curve (AUC) was plotted to analyse the ability of serum Cys-C, KIM-1 and REG Iα to screen the patients with CKD and detect the high- and very-high-risk patients. The study assessed to evaluate accuracy using sensitivity, specificity, negative predictive value (NPV), positive predictive value (PPV) and accuracy, and applied the DeLong test to statistically analyse AUC differences between receiver operating characteristic (ROC) curves. P value of <0.050 using two-tailed tests was considered statistically significant.

### RESULTS

#### Baseline characteristics of the study population

Overall, a total of 880 participants were enrolled, comprising 220 non-CKD participants and 660 patients with CKD (online supplemental data figure 1). Significant differences were observed between patients with CKD and non-CKD participants in terms of age, complication diseases (diabetes and hypertension), kidney function biomarkers (Scr, BUN, UA, eGFR, Cys-C and KIM-1) and serum myoglobin (all p<0.001, table 1). Serum REG Iα levels were significantly elevated in patients with CKD (125.54 (60.28, 303.39) ng/mL) compared with non-CKD participants (24.62 (14.09, 37.32) ng/mL, p<0.001, table 1). Biomarker trends revealed that serum Cys-C, REG Iα and myoglobin levels progressively increased as eGFR declined (figure 1: A1, C1 and D1). Serum KIM-1 levels also exhibited significant differences between the non-CKD and CKD groups (G2 to G5, p<0.010, figure 1: B1). Further analyses confirmed that cardiovascular disease (online supplemental data figure 2) did not influence the distribution of serum REG Ia and a myoglobin levels. Diabetes had no influence on serum ō REG Ia in patients with CKD (online supplemental data figure 3).

Table 1     Clinical characteristics of the study population at baseline examination					
	Non-CKD participants	Patients with CKD	P value		
Number	220	660	-		
Demographics					
Age (years)	53 (40–62)	62 (50–72)	< 0.001		
Sex (male, %)	111 (50.45)	366 (55.45)	0.212		
Smoking (yes, %)	41 (18.64)	142 (21.52)	0.141		
Drinking (yes, %)	30 (13.64)	84 (12.73)	0.823		
Complication diseases					
Diabetes (yes, %)	75 (34.09)	350 (53.03)	< 0.001		
Hypertension (yes, %)	65 (29.55)	501 (75.91)	< 0.001		
Cardiovascular disease (yes, %)	44 (20.00)	149 (22.58)	0.453		
Laboratory measurements					
Fast blood glucose (mmol/L)	5.45 (4.99–6.67)	5.34 (4.57–7.11)	0.008		
Serum creatinine (µmol/L)	64.00 (55.25–76.00)	126.00 (83.00–418.50)	< 0.001		
Blood urea nitrogen (mmol/L)	5.20 (4.30–6.28)	10.30 (7.00–18.60)	< 0.001		
Serum uric acid (µmol/L)	301.00 (256.00–365.25)	354.00 (290.00–443.00)	< 0.001		
Estimated glomerular filtration rate (mL/ min/1.73 m <sup>2</sup> )	102.37 (95.73–112.36)	44.03 (10.96–77.09)	<0.001		
Serum cystatin C (mg/L)	1.10 (0.99–1.24)	1.79 (1.23–3.50)	< 0.001		
Serum kidney injury molecule 1 (pg/mL)	186.28 (57.22–266.88)	247.72 (175.10–334.13)	< 0.001		
Serum myoglobin (ng/mL)	46.81 (33.49–58.00)	64.04 (35.96–112.51)	< 0.001		
Serum regenerating protein Ia (ng/mL)	24.62 (14.09–37.32)	125.54 (60.28–303.39)	< 0.001		

Data are presented in quartiles. Student's t-tests were used for normally distributed continuous variables. Mann-Whitney U tests were used for non-normally distributed continuous variables.  $\chi^2$  or Fisher's exact tests were used for categorical variables for two group comparisons. CKD, chronic kidney disease.



**Figure 1** Distribution of serum Cys-C, serum KIM-1, serum REG Ia and serum myoglobin in different groups. A1, B1, C1 and D1 show distributions in all participants: the CKD groups were classified in accordance with eGFR levels as described in the Methods section. A1: Distribution of serum Cys-C. B1: Distribution of serum KIM-1. C1: Distribution of serum REG Ia. D1: Distribution of serum myoglobin. A2, B2, C2 and D2 show distributions in patients with CKD: the patients were classified in accordance with KDIGO 2012 risk stratification as described in the Methods section. A2: Distribution of serum Cys-C. B2: Distribution of serum KIM-1. C2: Distribution of serum REG Ia. D2: Distribution of serum Myoglobin. \*p<0.050, \*\*p<0.010 and \*\*\*p<0.001. Tukey's multiple comparison tests were conducted to examine the differences in different groups. CKD, chronic kidney disease; Cys-C, cystatin C; KDIGO, Kidney Disease Improving Global Outcomes; KIM-1, kidney injury molecule 1; REG Ia, regenerating protein Ia.

#### Relationship between serum REG I $\alpha$ and kidney function

This study explored the relationship between serum REG I $\alpha$  levels and kidney function biomarkers. In figure 2, Spearman correlation analyses demonstrated a strong positive correlation between serum REG I $\alpha$  and Scr (r=0.753, p<0.001), BUN (r=0.733, p<0.001), serum Cys-C (r=0.678, p<0.001) and serum KIM-1 (r=0.217, p<0.001). A significant negative correlation was observed between serum REG I $\alpha$  and eGFR (r=-0.789, p<0.001). A comprehensive summary of these correlations was provided in online supplemental data table 1. Ordinal logistic regression analysis carried out in all participants, with eGFR as a grade-dependent variable, revealed that serum REG I $\alpha$ /100 levels were significantly associated with eGFR (OR=1.737, 95% CI: 1.263 to 2.388, p=0.001, table 2).

#### Subgroup analysis in patients with CKD

To explore the relationship between serum REG I $\alpha$  levels and CKD progression risk, 256 patients with CKD were included in the sub-research and classified by eGFR and UACR levels in accordance with KDIGO risk stratification guidelines.<sup>1</sup> The patients were stratified into four KDIGO risk groups: low risk (18.00%), moderate risk (20.70%), high risk (17.20%) and very high risk (44.10%, online supplemental data table 2). Serum REG I $\alpha$  levels significantly increased with higher CKD risk categories, reaching 184.38 (108.81, 314.71) ng/mL in the very-high-risk group (p<0.001, figure 1: C2). Similar trends were observed for serum Cys-C (2.57 (1.94, 3.53) mg/L) and myoglobin (67.00 (51.70, 113.68) ng/mL, p<0.001, figure 1: A2, D2). However, the serum KIM-1 did not exhibit an increasing trend (p>0.050, figure 1: B2). Multiple logistic regression analysis demonstrated that serum REG I $\alpha$ /100 was an independent influencing factor for high- and very-highrisk CKD (OR=1.799, 95% CI: 1.088 to 2.975, p=0.022, table 2).

## Ability of serum REG I $\alpha$ in screening patients with kidney dysfunction

ROC analysis evaluated the utility of serum REG I $\alpha$  as a screening tool for CKD and its ability to stratify CKD risk (figure 3 and online supplemental data table 3). Serum REG I $\alpha$  demonstrated an AUC of 0.860 (95% CI: 0.813 to 0.899) for detecting CKD compared with Scr (0.850, 95% CI: 0.801 to 0.890) and serum Cys-C (0.842, 95% CI: 0.793 to 0.883). At a cut-off value of 70.82 ng/mL, serum REG I $\alpha$  had a sensitivity of 71.63%, specificity of 86.89%, PPV of 94.30% and NPV of 46.85%. Serum KIM-1 showed a lower AUC than serum REG I $\alpha$ , measuring 0.714 (95% CI: 0.656 to 0.767, p<0.001). Serum myoglobin had the lowest AUC among the five biomarkers, measuring 0.642 (95% CI: 0.581 to 0.699, p<0.001).

For distinguishing high- and very-high-risk CKD, serum REG Iα demonstrated superior performance compared with serum KIM-1 (AUC=0.769 (0.712–0.819) vs 0.528 (0.465–0.590), p<0.010, online supplemental data table 3). Serum Cys-C had the highest AUC (0.865 (0.817–0.904), p<0.010) among the three biomarkers. Serum REG Iα,

with a cut-off value of 76.05 ng/mL, exhibited sensitivity of 82.80%, specificity of 62.63%, PPV of 77.38% and NPV of 69.32%. Notably, serum REG Iα showed significantly higher sensitivity than serum Cys-C and KIM-1.

#### DISCUSSION

This is the first study to systematically evaluate serum REG I $\alpha$  levels in a broad CKD population and confirm its upregulation in patients with CKD. First, serum REG I $\alpha$  levels increased progressively with declining eGFR and correlated strongly with conventional kidney function biomarkers (Scr, BUN, Cys-C and KIM-1). Second, serum REG I $\alpha$  emerged as an independent risk factor for patients classified as high and very high risk according to the KDIGO risk stratification. Third, serum REG I $\alpha$  emerged robust diagnostic performance, showing higher sensitivity than serum KIM-1 for identifying CKD and distinguishing risk progression.

REG Ia, a low-molecular-weight protein (16 kDa), was initially discovered in the pancreas and was identified as PSP due to its role in inhibiting the formation of calcium carbonate stones in pancreatic ducts.<sup>29 30</sup> Immunohistochemical studies have shown overexpression of REG Ia in impaired kidneys, particularly in proximal tubules and thick ascending limbs of Henle's loops.<sup>20</sup> Previous studies also reported elevated REG Ia levels in DKD, suggesting its involvement in tubular dysfunction and kidney injury.<sup>22 31 32</sup> Our previous studies provided evidence that serum REG Ia levels were notably elevated in patients with type 2 diabetes mellitus who had kidney dysfunction. We also observed that pregnant women with early renal dysfunction exhibited similar elevations in serum REG Ia levels.<sup>24 33</sup> Notably, the serum REG Ia levels remained  $\exists$ significantly elevated when the scope of this study was expanded to the entire CKD population.

We incorporated myoglobin (17.6 kDa)<sup>34</sup> to investigate ≥ whether the elevated level of serum REG Ia represents a universal phenomenon of accumulation in CKD. ğ The myoglobin was identified as a biomarker for acute myocardial ischemia and rhabdomyolysis.<sup>35</sup> Some studies indicated that serum myoglobin levels increase in kidney impairment. This accumulation was primarily attributed to slower clearance in CKD.<sup>36</sup> We showed that serum levels of myoglobin have no differences between non-CKD participants and patients with early-stage CKD (group G1 and group G2). There was also a gradual increase in **O** patients with medium- to end-stage CKD (group G3 to g group G5). The possible reason was as follows: in the early **3** stages of kidney injury, compensatory glomerular hypertrophy with a hyperfiltration state was known to exist. This mechanism allowed some low-molecular-weight protein to be filtered through the glomerular basement membrane, resulting in no discernible difference during the early stage of CKD (eGFR  $\geq 60 \text{ mL/min}/1.73 \text{ m}^2$ ) and even appearing as a transient decrease. With progression to medium and end stages of CKD (eGFR <60 mL/ min/1.73 m<sup>2</sup>), the glomerular basement membrane



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**Figure 2** Correlations between serum REG Ia and different markers in all participants. A: correlation between serum REG Ia and Scr (r=0.753, p<0.001), B: correlation between serum REG Ia and BUN (r=0.733, p<0.001), C: correlation between serum REG Ia and serum Cys-C (r=0.678, p<0.001), D: correlation between serum REG Ia and serum KIM-1 (r=0.217, p<0.001) and E: correlation between serum REG Ia and eGFR (r=-0.789, p<0.001). BUN, blood urea nitrogen; Cys-C, cystatin C; eGFR, estimated glomerular filtration rate; KIM-1, kidney injury molecule 1; REG Ia, regenerating protein Ia; Scr, serum creatinine.

Table 2     Logistic regression analyses showing the relationship between variables and kidney function					
Ordinal logistic regression*		Multivariate logistic regression†			
P value	OR (95% CI)	P value	OR (95% CI)		
0.050	1.020 (1.001 to 1.041)	0.009	0.966 (0.942 to 0.992)		
<0.001	1.266 (1.165 to 1.376)	<0.001	1.440 (1.216 to 1.706)		
0.337	1.001 (0.999 to 1.004)	0.085	1.003 (1.000 to 1.007)		
0.165	1.005 (0.998 to 1.013)	0.148	1.136 (0.897 to 1.559)		
<0.001	6.784 (4.016 to 11.460)	0.071	1.853 (0.949 to 3.620)		
0.133	1.069 (0.980 to 1.167)	0.122	1.243 (0.943 to 1.639)		
0.001	1.737 (1.263 to 2.388)	0.022	1.799 (1.088 to 2.975)		
	n analyses showing th Ordinal logistic reg P value 0.050 <0.001 0.337 0.165 <0.001 0.133 0.001	P value     OR (95% Cl)       0.050     1.020 (1.001 to 1.041)       <0.001	P value     OR (95% Cl)     P value       0.050     1.020 (1.001 to 1.041)     0.009       <0.001		

\* The ordinal multiple logistic regression shows variables independently associated with eGFR levels in all participants.

† The multivariate logistic regression analysis identified the independent influencing factors for high- and very-high-risk patients with CKD in accordance with KDIGO risk stratification. The analyses included age, BUN, UA, serum myoglobin, serum Cys-C, serum KIM-1/100 and serum REG Ia/100 into ordinal multiple logistic regression model, while adjusting for sex, diabetes, hypertension and FBG. The multivariate logistic regression model also incorporates the above covariates.

‡ years.

§ mmol/L.

¶µmol/L.

\* ng/mL.

†† mg/L.

‡‡ pg/mL.

BUN, blood urea nitrogen; CKD, chronic kidney disease; Cys-C, cystatin C; eGFR, estimated glomerular filtration rate; FBG, fast blood glucose; KDIGO, Kidney Disease Improving Global Outcomes; KIM-1, kidney injury molecule 1; REG Ια, regenerating protein Ια; UA, uric acid.



 Serum REG I $\alpha$ , AUC = 0.860 (0.813-0.899);
 Serum Creatinine, AUC = 0.850 (0.801-0.890);
 Serum Cys-C, AUC = 0.842 (0.793-0.883);
 Serum KIM-1, AUC = 0.714 (0.656-0.767);
 Serum Myoglobin, AUC = 0.642 (0.581-0.699);

Figure 3 Ability of screening patients with CKD. The AUC of serum REG Ia was 0.860 (95% CI: 0.813 to 0.899) and that of Scr was 0.850 (95% CI: 0.801 to 0.890), while that of serum Cys-C was 0.842 (95% CI: 0.793 to 0.883). Serum KIM-1 had an AUC of 0.714 (95% CI: 0.656 to 0.767), and serum myoglobin had an AUC of 0.642 (95% CI: 0.581 to 0.699). The AUC of serum REG Ia was similar to Scr and serum Cys-C (p>0.050) and was significantly higher than serum KIM-1 and serum myoglobin (p<0.001). The DeLong tests were applied to analyse AUC differences between receiver operating characteristic curves. AUC, area under the receiver operating characteristic curve; CKD, chronic kidney disease; Cys-C, cystatin C; KIM-1, kidney injury molecule 1; REG Ia, regenerating protein Ia.

thickened and led to a significant decline in glomerular filtration function, ultimately causing protein accumulation in serum.<sup>37</sup> The serum levels of REG Ia showed a gradual increase in medium to end stages of CKD, which a was similar to the rise observed in serum myoglobin levels. However, it exhibited a significant increase in the early stage of CKD, which was different with serum myoglobin. This suggested that, in addition to the sensitive accumulation of serum REG I $\alpha$  as a low-molecular-weight protein, there might be a specific production when it comes to kidney impairment. The unique behaviour of serum REG Iα, compared with low-molecular-weight protein, highlights its dual role: accumulation due to reduced glomerular clearance and potentially increased production in response to kidney injury. Unlike myoglobin, which remains stable in early-stage CKD, serum REG Ia levels were significantly elevated even at the ultra-early stage of kidney dysfunction, suggesting a sensitive and specific response to renal impairment.

The aetiology underlying the upregulation of REG I $\alpha$ production in patients with CKD remains elusive. Two potential mechanisms are considered as follows. First, REG Ia resists apoptosis and promotes cell proliferation in different inflammation situations.<sup>38–42</sup> Studies have shown that cytokines such as IL-6 can increase the proliferation of REG Ia, which is involved in cell regeneration and repair.<sup>42 43</sup> In CKD, the occurrence and progression of the disease are closely related to the expression of multiple inflammatory cytokines, including IL-1, IL-6 and TNF- $\alpha$ .<sup>37 44 45</sup> Under the stimulation of chronic inflammation, different types of renal cells secrete REG Ia locally to participate in kidney antiapoptosis and proliferation and

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against kidney fibrosis in the development of CKD. The secreted REG Ia enters the circulation through reabsorption by kidney tubules, resulting in a significant increase in serum REG Ia levels. Thus, REG Ia might serve as an inflammatory factor involved in kidney diseases. Second, REG Ia is primarily synthesised in the pancreas and released into circulation.<sup>18 46</sup> A hypothesis suggests that a cross-talk mechanism exists between the pancreas and kidney when renal damage occurs. This interaction may prompt the pancreas to produce more REG I $\alpha$  in response to kidney injury, resulting in the elevation in serum. Although the exact cause of REG Ia upregulation remains unclear, these two proposed mechanisms provided potential insights into the relationship between REG Ia and CKD.

At present, the assessment of CKD generally focuses on glomerular filtration capacity, which is characterised by Scr, UACR and eGFR.<sup>2</sup> The stabilities of these factors are compromised by age, dietary intake and physical activity. In general, the performance of eGFR is poor for populations outside of North America, Australia and Europe, and accuracy decreases at the extremes in the distribution of age and body composition.<sup>7</sup> Therefore, new biomarkers of kidney impairment have been discovered. Cys-C is produced by all nucleated cells at a steady rate and freely filtered by the kidney.<sup>47</sup> The accumulation of its levels is observed in the case of glomerular filtration dysfunction, with limited impact and strong stability.<sup>48</sup> Eventually, serum Cys-C is incorporated into the eGFR equation as a reliable and sensitive indicator of kidney function. The sensitivity of serum Cys-C is heightened in the early stages of CKD compared with creatinine.<sup>4950</sup> Another biomarker is KIM-1,<sup>51 52</sup> which is secreted following kidney proximal tubular injury and has emerged as a sensitive and specific biomarker of acute kidney injury and CKD progression.<sup>53–56</sup> Compared with serum KIM-1, the REG Ia had several advantages as follows. This study demonstrated that levels of serum REG I $\alpha$  increase significantly earlier than KIM-1, making it a better marker for the early detection of renal injury. In addition, serum REG Ia was sensitive in distinguishing between different stages of CKD. Its ability to discriminate early from advanced stages of CKD provides valuable diagnostic and prognostic information. Serum REG Ia also exhibited better AUC, sensitivity and specificity, enhancing its diagnostic performance in identifying patients with CKD. In summary, the advantages of serum REG Ia over serum KIM-1 primarily lie in its ability to detect renal injury earlier, its sensitivity in differentiating various stages of CKD and its better diagnostic performance in identifying patients with CKD. These features collectively underscore its potential as a potential biomarker for CKD.

In this study, serum REG Ia was strongly correlated with Scr and Cys-C and had a similar performance to Scr and Cys-C in detecting patients with CKD. The serum levels of REG I $\alpha$  were more likely related to glomerular filtration capacity according to the correlation analyses, although the mechanism of its production and degradation or

CKD. These results highlight the potential application of patients with CKD and detecting high- and very-high-risk of patients with CKD and the assessment of CKD risk. The sensitivity in identifying high- and very-high-risk of serum REG Iα as a valuable biomarker in the screening of patients with CKD and the assessment of CKD risk. These results highlight the potential application of CKD participants, and they had higher rates of CKD.<sup>2</sup> Diabetes accounts for 30%–50% of all CKD causes of the increased risk of developing is associated with blood pressure control. These findings were consistent with the typical aetiology of CKD aetiology in our study.

team has endeavoured to conduct a pre-subgroup analā ysis within the available constraints. We found that CKD etiologies (eg, IgA nephropathy, membranous nephropathy and DKD) have no effect on serum REG levels. To enhance causality and generalisability, longitudinal ≥ designs with larger sample sizes and diverse populations should be considered in forthcoming research. Second, ß although robust biomarkers such as creatinine and Cys-C are already available for the diagnosis of CKD, the potential efficacy of REG Ia as a combined biomarker is antic-S ipated to be revealed in subsequent large-scale analyses. Third, the precise source of elevated REG Ia in CKD remains unclear, warranting further mechanistic investigations. Fourth, to address residual confounding, future studies should incorporate additional covariates such as og drug use and different causes of CKD and conduct sensitivity analyses to assess potential biases.

Moreover, we acknowledged the potential impact of self-reporting and recall bias in our questionnaire-based demographic data, especially among patients with CKD prone to cognitive and emotional challenges. Factors, such as cognitive impairment, symptom complexity, emotional stress, health literacy variations and medication effects, can skew reporting accuracy. To mitigate risks, we used standardised questionnaires with clear instructions. However, given CKD's clinical complexity,

This study provided compelling evidence that serum REG I $\alpha$  is significantly upregulated in patients with CKD and strongly associated with kidney function. Serum REG I $\alpha$  demonstrated notable diagnostic sensitivity and utility in CKD risk stratification, underscoring its potential as a valuable biomarker for detecting kidney function decline and identifying patients with high-risk CKD.

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Acknowledgements The biological samples were provided by the Biobank of Zhongda Hospital, Southeast University.

**Contributors** LL acted as guarantor. LL conceptualised the study. BW reviewed and edited the manuscript. NH and XZ were responsible for data analysis and wrote the original draft. NH, SC, XW, HW, ZS and XH were responsible for the inclusion of the population, collection of samples and data, and the experiments. JB and JS were responsible for guidance on data analysis. PC and X-x H were responsible for the analysis of kidney biopsy tissues from patients with CKD. RG provided excellent technical support for this study.

**Funding** This work was supported by the National Natural Science Foundation of China (No. 81970717, 82000740, 82170845 and 82200929), the Key Research & Development Program (No. BE2022853), the Medical Key Discipline of Jiangsu Province (ZDXK202203) and the Open Project of Key Laboratory of Environmental Medicine Engineering of Ministry of Education.

Competing interests None declared.

**Patient and public involvement** Patients and/or the public were not involved in the design, or conduct, or reporting, or dissemination plans of this research.

#### Patient consent for publication Not applicable.

Ethics approval This study involves human participants. The study was approved by the ethics committee of Zhongda Hospital (2022ZDSYLL204-P01) and was in compliance with the Helsinki Declaration. The study had a clinical study registration number of ChiCTR2300072247. Participants gave informed consent to participate in the study before taking part.

Provenance and peer review Not commissioned; externally peer reviewed.

Data availability statement Data are available upon reasonable request. The data underlying this article is available from the corresponding author under reasonable request.

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### REFERENCES

- 1 Stevens PE, Levin A, Kidney Disease: Improving Global Outcomes Chronic Kidney Disease Guideline Development Work Group Members. Evaluation and management of chronic kidney disease: synopsis of the kidney disease: improving global outcomes 2012 clinical practice guideline. *Ann Intern Med* 2013;158:825–30.
- 2 Webster AC, Nagler EV, Morton RL, *et al.* Chronic Kidney Disease. *Lancet* 2017;389:1238–52.
- 3 Kalantar-Zadeh K, Jafar TH, Nitsch D, et al. Chronic kidney disease Lancet 2021;398:786–802.
- 4 Kovesdy CP. Epidemiology of chronic kidney disease: an update 2022. *Kidney Int Suppl (2011)* 2022;12:7–11.
- 5 Hemmelgarn BR, Manns BJ, Lloyd A, et al. Relation between kidney function, proteinuria, and adverse outcomes. JAMA 2010;303:423–9.
- 6 Tonelli M, Muntner P, Lloyd A, et al. Using proteinuria and estimated glomerular filtration rate to classify risk in patients with chronic kidney disease: a cohort study. Ann Intern Med 2011;154:12–21.
- 7 Earley A, Miskulin D, Lamb EJ, *et al.* Estimating equations for glomerular filtration rate in the era of creatinine standardization: a systematic review. *Ann Intern Med* 2012;156:785–95.
- 8 Inker LA, Levey AS, Pandya K, et al. Early change in proteinuria as a surrogate end point for kidney disease progression: an individual patient meta-analysis. Am J Kidney Dis 2014;64:74–85.
- 9 Andrassy KM. Comments on 'KDIGO 2012 Clinical Practice Guideline for the Evaluation and Management of Chronic Kidney Disease'. *Kidney Int* 2013;84:622–3.
- 10 Colhoun HM, Marcovecchio ML. Biomarkers of diabetic kidney disease. *Diabetologia* 2018;61:996–1011.
- 11 Singh D, Whooley MA, Ix JH, *et al.* Association of cystatin C and estimated GFR with inflammatory biomarkers: the Heart and Soul Study. *Nephrol Dial Transplant* 2007;22:1087–92.
- 12 Kerschbaum J, Rudnicki M, Dzien A, et al. Intra-individual variability of eGFR trajectories in early diabetic kidney disease and lack of performance of prognostic biomarkers. Sci Rep 2020;10:19743.
- 13 Colombo M, Looker HC, Farran B, *et al.* Serum kidney injury molecule 1 and  $\beta_2$ -microglobulin perform as well as larger biomarker panels for prediction of rapid decline in renal function in type 2 diabetes. *Diabetologia* 2019;62:156–68.
- 14 Heinzel A, Kammer M, Mayer G, et al. Validation of Plasma Biomarker Candidates for the Prediction of eGFR Decline in Patients With Type 2 Diabetes. *Diabetes Care* 2018;41:1947–54.
- 15 Woo KS, Choi JL, Kim BR, et al. Clinical usefulness of serum cystatin C as a marker of renal function. *Diabetes Metab J* 2014;38:278–84.
- 16 Terazono K, Yamamoto H, Takasawa S, *et al*. A novel gene activated in regenerating islets. *J Biol Chem* 1988;263:2111–4.
- 17 Lopes D, Chumbinho B, Bandovas JP, *et al.* Pancreatic stone protein as a biomarker of sepsis. *Crit Care* 2022;26:100.
- 18 Graf R, Schiesser M, Reding T, *et al.* Exocrine meets endocrine: pancreatic stone protein and regenerating protein--two sides of the same coin. *J Surg Res* 2006;133:113–20.
- 19 Watanabe T, Yonekura H, Terazono K, et al. Complete nucleotide sequence of human reg gene and its expression in normal and tumoral tissues. The reg protein, pancreatic stone protein, and pancreatic thread protein are one and the same product of the gene. J Biol Chem 1990;265:7432–9.
- 20 Verdier JM, Dussol B, Casanova P, et al. Evidence that human kidney produces a protein similar to lithostathine, the pancreatic inhibitor of CaCO3 crystal growth. *Eur J Clin Invest* 1992;22:469–74.
- 21 Tatemichi N, Takahashi C, Hayakawa S, et al. Enzyme immunoassay and characterization of pancreatic stone proteins in human urine. J Clin Lab Anal 1993;7:365–70.
- 22 Sobajima H, Niwa T, Shikano M, *et al.* Urinary excretion of pancreatic stone protein in diabetic nephropathy. *Intern Med* 1998;37:500–3.
- 23 Yang J, Li L, Raptis D, *et al*. Pancreatic stone protein/regenerating protein (PSP/reg): a novel secreted protein up-regulated in type 2 diabetes mellitus. *Endocrine* 2015;48:856–62.

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- 24 Zhu H, Zhu X, Lin H, *et al*. Association of Serum PSP/REG Iα with Renal Function in Type 2 Diabetes Mellitus. *J Diabetes Res* 2020;2020:9787839.
- 25 Levey AS, Stevens LA, Schmid CH, et al. A new equation to estimate glomerular filtration rate. Ann Intern Med 2009;150:604–12.
- 26 National Kidney F. K/DOQI clinical practice guidelines for chronic kidney disease: evaluation, classification, and stratification. *Am J Kidney Dis* 2002;39:S1–266.
- 27 Levey AS, Eckardt K-U, Dorman NM, *et al.* Nomenclature for kidney function and disease: report of a Kidney Disease: Improving Global Outcomes (KDIGO) Consensus Conference. *Kidney Int* 2020;97:1117–29.
- 28 Keel M, Härter L, Reding T, et al. Pancreatic stone protein is highly increased during posttraumatic sepsis and activates neutrophil granulocytes. Crit Care Med 2009;37:1642–8.
- 29 Bimmler D, Graf R, Scheele GA, et al. Pancreatic stone protein (lithostathine), a physiologically relevant pancreatic calcium carbonate crystal inhibitor? J Biol Chem 1997;272:3073–82.
- 30 Okamoto H. The Reg gene family and Reg proteins: with special attention to the regeneration of pancreatic beta-cells. *J Hepatobiliary Pancreat Surg* 1999;6:254–62.
- 31 Koziolek M, Mueller GA, Dihazi GH, et al. Urine E-cadherin: A Marker for Early Detection of Kidney Injury in Diabetic Patients. J Clin Med 2020;9:639.
- 32 Wang X, Wu H, Yang G, *et al.* REG1A and RUNX3 Are Potential Biomarkers for Predicting the Risk of Diabetic Kidney Disease. *Front Endocrinol (Lausanne)* 2022;13:935796.
- 33 Zhu X, Dong B, Reding T, et al. Association of Serum PSP/ REG Iα with Renal Function in Pregnant Women. *Biomed Res Int* 2019;2019:6970890.
- 34 Jiang J, Xia M, Gong H, et al. Effect of magnetic field modification on oxidative stability of myoglobin in sarcoplasm systems. Food Chem 2024;436:137691.
- 35 Lenglet A, Liabeuf S, Desjardins L, et al. Prognostic implications of plasma myoglobin levels in patients with chronic kidney disease. Int J Artif Organs 2012;35:959–68.
- 36 Hart PM, Feinfeld DA, Briscoe AM, et al. The effect of renal failure and hemodialysis on serum and urine myoglobin. *Clin Nephrol* 1982;18:141–3.
- 37 Piwowar A, Knapik-Kordecka M, Fus I, et al. Urinary activities of cathepsin B, N-acetyl-beta-D-glucosaminidase, and albuminuria in patients with type 2 diabetes mellitus. *Med Sci Monit* 2006;12:CR210–4.
- 38 Mueller CM, Zhang H, Zenilman ME. Pancreatic reg I binds MKP-1 and regulates cyclin D in pancreatic-derived cells. *J Surg Res* 2008;150:137–43.
- 39 Unno M, Nata K, Noguchi N, et al. Production and characterization of Reg knockout mice: reduced proliferation of pancreatic beta-cells in Reg knockout mice. *Diabetes* 2002;51 Suppl 3:S478–83.
- 40 Wang J, Koyota S, Zhou X, et al. Expression and localization of regenerating gene I in a rat liver regeneration model. *Biochem Biophys Res Commun* 2009;380:472–7.
- 41 Nakagawa K, Takasawa S, Nata K, et al. Prevention of Reg I-induced β-cell apoptosis by IL-6/dexamethasone through activation of HGF gene regulation. *Biochim Biophys Acta* 2013;1833:2988–95.
- 42 Sekikawa A, Fukui H, Fujii S, et al. REG lalpha protein mediates an anti-apoptotic effect of STAT3 signaling in gastric cancer cells. *Carcinogenesis* 2008;29:76–83.

- 43 Tohma Y, Dohi Y, Shobatake R, et al. Reg Gene Expression in Periosteum after Fracture and Its In Vitro Induction Triggered by IL-6. Int J Mol Sci 2017;18:2257.
- 44 Shlipak MG, Fried LF, Crump C, *et al.* Elevations of inflammatory and procoagulant biomarkers in elderly persons with renal insufficiency. *Circulation* 2003;107:87–92.
- 45 Galkina E, Ley K. Leukocyte recruitment and vascular injury in diabetic nephropathy. *J Am Soc Nephrol* 2006;17:368–77.
- 46 Jin CX, Hayakawa T, Ko SBH, et al. Pancreatic stone protein/ regenerating protein family in pancreatic and gastrointestinal diseases. Intern Med 2011;50:1507–16.
- 47 Abrahamson M, Olafsson I, Palsdottir A, et al. Structure and expression of the human cystatin C gene. *Biochem J* 1990;268:287–94.
- 48 Chew JSC, Saleem M, Florkowski CM, et al. Cystatin C--a paradigm of evidence based laboratory medicine. *Clin Biochem Rev* 2008;29:47–62.
- 49 Levey AS, Fan L, Eckfeldt JH, et al. Cystatin C for glomerular filtration rate estimation: coming of age. Clin Chem 2014;60:916–9.
- 50 Benoit SW, Ciccia EA, Devarajan P. Cystatin C as a biomarker of chronic kidney disease: latest developments. *Expert Rev Mol Diagn* 2020;20:1019–26.
- 51 Ichimura T, Bonventre JV, Bailly V, et al. Kidney injury molecule-1 (KIM-1), a putative epithelial cell adhesion molecule containing a novel immunoglobulin domain, is up-regulated in renal cells after injury. J Biol Chem 1998;273:4135–42.
- 52 Han WK, Bailly V, Abichandani R, *et al.* Kidney Injury Molecule-1 (KIM-1): a novel biomarker for human renal proximal tubule injury. *Kidney Int* 2002;62:237–44.
- 53 Ichimura T, Hung CC, Yang SA, et al. Kidney injury molecule-1: a tissue and urinary biomarker for nephrotoxicant-induced renal injury. Am J Physiol Renal Physiol 2004;286:F552–63.
- 54 Sabbisetti VS, Waikar SS, Antoine DJ, et al. Blood kidney injury molecule-1 is a biomarker of acute and chronic kidney injury and predicts progression to ESRD in type I diabetes. J Am Soc Nephrol 2014;25:2177–86.
- 55 Vaidya VS, Niewczas MA, Ficociello LH, *et al.* Regression of microalbuminuria in type 1 diabetes is associated with lower levels of urinary tubular injury biomarkers, kidney injury molecule-1, and N-acetyl-β-D-glucosaminidase. *Kidney Int* 2011;79:464–70.
- 56 Panduru NM, Sandholm N, Forsblom C, et al. Kidney injury molecule-1 and the loss of kidney function in diabetic nephropathy: a likely causal link in patients with type 1 diabetes. *Diabetes Care* 2015;38:1130–7.
- 57 Pugliese G, Solini A, Bonora E, et al. Distribution of cardiovascular disease and retinopathy in patients with type 2 diabetes according to different classification systems for chronic kidney disease: a crosssectional analysis of the renal insufficiency and cardiovascular events (RIACE) Italian multicenter study. *Cardiovasc Diabetol* 2014;13:59.
- 58 Murton M, Goff-Leggett D, Bobrowska A, et al. Burden of Chronic Kidney Disease by KDIGO Categories of Glomerular Filtration Rate and Albuminuria: A Systematic Review. Adv Ther 2021;38:180–200.
- 59 Nichols GA, Déruaz-Luyet A, Brodovicz KG, *et al.* Kidney disease progression and all-cause mortality across estimated glomerular filtration rate and albuminuria categories among patients with vs. without type 2 diabetes. *BMC Nephrol* 2020;21:167.
- 60 Gimeno-Orna JA, Rodríguez-Padial L, Anguita-Sánchez M, et al. Association of the KDIGO Risk Classification with the Prevalence of Heart Failure in Patients with Type 2 Diabetes. J Clin Med 2021;10:4634.