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BMJ Open

Assessing the Impact of Tobacco-Induced Volatile Organic Compounds on Cardiovascular Risk in a Cross-Sectional Cohort: Cardiovascular Injury Due to Tobacco Study

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| Keywords: | smoking, tobacco, electronic cigarette, cardiovascular risk, vascular injury, cigarettes |
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5 2 **Cardiovascular Risk in a Cross-Sectional Cohort: Cardiovascular Injury Due to**

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8 3 **Tobacco Study**

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10 4 Rachel J. Keith, Jessica L. Fetterman, Daniel W. Riggs, Tim O’Toole, Jessica Nystoriak,

11

12 5 Monica Holbrook, Pawel Lorkiewicz, Aruni Bhatnagar, Andrew DeFilippis*, Naomi M.

13

14

15 6 Hamburg*

16

17 7 Rachel J. Keith-Division of Cardiovascular Medicine, University of Louisville School of

18

19 8 Medicine 580 S. Preston St. Louisville KY, 40202 rachel.keith@louisville.edu 502-852-

20

21 9 4211

22

23

24 10 Jessica L. Fetterman- Vascular Biology Section, Whitaker Cardiovascular Institute,

25

26 11 Boston University School of Medicine Evans Building, Boston, MA USA

27

28 12 Jasmit Shah- University of Louisville School of Medicine Louisville, KY USA

29

30

31 13 Timothy O-Toole- Division of Cardiovascular Medicine, University of Louisville School of

32

33 14 Medicine Louisville, KY USA

34

35 15 Jessica L. Nystoriak- Division of Cardiovascular Medicine, University of Louisville

36

37 16 School of Medicine Louisville, KY USA

38

39

40 17 Monika Holbrook- Vascular Biology Section, Whitaker Cardiovascular Institute, Boston

41

42 18 University School of Medicine Boston, MA USA

43

44

45 19 Pawel Lorkiewicz- Division of Cardiovascular Medicine, University of Louisville School

46

47 20 of Medicine Louisville, KY USA

48

49 21 Aruni Bhatnagar- Division of Cardiovascular Medicine, University of Louisville School of

50

51 22 Medicine Louisville, KY USA

52

53

54

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56

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Andrew P. DeFilippis- Division of Cardiovascular Medicine, University of Louisville
School of Medicine Louisville, KY USA (co-senior author)

Naomi M. Hamburg- Vascular Biology Section, Whitaker Cardiovascular Institute,
Boston University School of Medicine Boston, MA USA (co-senior author)

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ABSTRACT

Introduction: Tobacco use leads to increased mortality, the majority of which is attributed to cardiovascular disease. Despite this knowledge, the early cardiovascular impact of tobacco product use is not well understood. Tobacco use increases exposure to harmful and potentially harmful constituents including volatile organic compounds (VOCs) such as acrolein and crotonaldehyde, which may contribute to cardiovascular risk. The link between exposure patterns, risk profiles and demographic distribution of tobacco product users, particularly users of new and emerging products, are not well known. Therefore, we designed the Cardiovascular Injury due to Tobacco Use (CITU) study to assess population characteristics, demographic features, exposure patterns and cardiovascular risk in relation to tobacco.

Methods and analysis: This is a cross-section observational study conducted in Boston MA and Louisville KY from 2014 through 2018. Healthy participants 21 to 45 years of age who use tobacco products, including ENDS, or who never used tobacco are being recruited. The study aims to recruit an evenly split cohort of African Americans and Caucasians that is sex balanced for evaluation of self-reported tobacco

exposure, VOC exposure and tobacco-induced injury profiling. Detailed information about participant’s demographics, health status and lifestyle is also collected.

Ethics and dissemination: The study protocol was approved institutional review boards at both participating universities. All study protocols will protect participant confidentiality. Results from the study will be disseminated via peer-reviewed journals and presented at scientific conferences.

Strengths and limitations

- Young age to allow for evaluation of early stage disease (e.g. inflammation, endothelial function) as opposed to end stage clinical consequence (e.g. myocardial infarction)
- Diverse tobacco product use allows for assessment of a wide range of tobacco-induced VOC exposure
- All study visits are in English introducing selection bias
- Data will inform regulatory agencies on the cardiovascular health effects of multiple tobacco products and the contribution of HPHCs

Keywords: Tobacco, smoking, electronic cigarette, vascular injury, cardiovascular risk, cigarettes.

INTRODUCTION

Tobacco product use and smoking are the leading causes of preventable deaths throughout the world. Of those deaths, one-third are attributed to cardiovascular disease

(CVD)¹. The cardiovascular (CV) effects of tobacco exposure can include atherogenesis, vascular injury, thrombosis, arrhythmias and inflammation² and may be attributable to the many different harmful and potentially harmful constituents (HPHCs) present in tobacco products.

The HPHCs found in tobacco products include volatile organic compounds (VOCs) of which reactive aldehydes, such as acrolein and crotonaldehyde, are likely the most significant contributors to CV toxicity³. High levels of aldehydes are present in cigarette smoke^{4 5} as well as smokeless tobacco (ST)⁶. Risk assessments, using the prevalence of each individual chemical weighed by its potency, suggest that the non-cancer risk of smoking is dominated by acrolein, which contributes 40-100 times more to risk than any other chemical present in cigarette smoke³.

Although HPHCs, including VOC reactive aldehydes, have been suspected to be major contributors to the toxicity of cigarette smoke for over 4 decades, their contribution to CV injury and early CVD risk has not been rigorously evaluated. Experimental studies in animal models suggest that because of low aldehyde-metabolizing capacity, CV tissues are highly sensitive to aldehydes and exposure to low levels of aldehydes can induce CV injury and accelerate CVD⁷⁻¹⁹. The WHO Study Group on Tobacco Product Regulation (TobReg) has marked acrolein, a VOC, along with 8 other cigarette constituents for monitoring and regulation²⁰ and the U.S. Environmental Protection Agency lists Acrolein as one of most hazardous air pollutants²¹. Nevertheless, the contribution of tobacco induced VOCs, including acrolein or other aldehydes, toward CV toxicity in humans has not been fully assessed. Greater understanding of how aldehydes affect cardiovascular health and disease will provide

new avenues for evaluating the toxicity of cigarette smoke and for assessing the injurious potential of new and emerging tobacco products, such as ENDS, which may also contain VOCs including acrolein²²⁻²⁴.

The latency period between tobacco exposure and the development of major clinical adverse health effects is long, therefore biomarkers that provide information over a shorter period allow for the identification of harm decades before clinical outcome data is available. Thus, the Cardiovascular Injury due To Tobacco Use (CITU) study evaluates the association of the urinary metabolites of 18 parent VOCs from tobacco exposure with a comprehensive set of CV biomarkers representative of early disease and predictive of future CV events.²⁵

METHODS AND DESIGN

Overall design

The CITU study is an investigator-initiated cross-sectional observational study of around 500 healthy participants 21 to 45 years of age who are never or current tobacco product users in two urban areas at Boston University (BU) and University of Louisville (UofL) (Boston, MA and Louisville, KY) designed to evaluate CV toxicity due to tobacco product use, with correlations to VOCs found in the tobacco products (Figure 1).

Figure 1. Cardiovascular Injury due to Tobacco Use

CITU is designed to assess how tobacco related VOC exposure contributes to cardiovascular risk factors. Our exposure measurements include a panel of 23 urinary metabolites of 18 parent VOCs and tobacco use patterns. Cardiovascular phenotyping

includes measures of injury, risk, vascular biomarkers and early vascular dysfunction. Tobacco use included use of traditional cigarettes, smokeless tobacco, waterpipe tobacco (hookah), electronic nicotine devices (ENDS), little cigars, cigarillos, pipes, cigars or any other form of tobacco that is available. Enrollment began in July 2014 and is ongoing.

Participant Eligibility Criteria

The goal of the study is to examine the impact of tobacco products on healthy young adults who could be classified as a current tobacco product users (Defined in table 1), or never-users (does not have lifetime use of any tobacco product); therefore we excluded participants if they had: 1) diagnosis of diabetes (HbA1c >7.0 or treatment for diabetes), hypertension (systolic blood pressure >140 mm Hg or diastolic blood pressure >90 mm Hg), hypothyroidism or hyperthyroidism, inflammatory conditions such as lupus or inflammatory bowel disease, HIV/AIDS, hepatitis, liver disease, anemia, cancer of any type or another medical condition that might compromise the successful completion of the study; 2) recipients of organ transplant or renal replacement therapy; 3) individuals that are taking the following medications: immunosuppressant agents estrogen, testosterone, anti TNF agents, certain biologics, Procrit, statins, beta-blockers or other cardiovascular medicine; 4) individuals using nutraceuticals or anabolic steroids beyond the recommended daily allowance; 5) body weight less than 100 pounds; 6) pregnant women; 7) prisoners and other vulnerable populations; and 8) active illness or infection. Participants are rescheduled or considered screen-failures and excluded from the study if symptomatic of an acute illness, i.e. viral upper respiratory infection, on study date.

Table 1. Tobacco product use classifications

| Classification | Qualification |
|------------------------|---|
| Never | Does not meet lifetime limits for any tobacco use (see below) |
| Smoker | >100 lifetime cigarettes and current use for the past year |
| Smokeless Tobacco User | >20 lifetime dips or chews and current use for the past year |
| Cigar/Cigarillo User | >20 lifetime cigars or cigarillos and current use for the past year |
| Pipe User | >20 lifetime pipefuls and current use for the past year |
| ENDS User | >20 lifetime vape sessions and current use for the past year |
| Hookah User | >20 lifetime hookah sessions and current use for the past year |

Study participants are screened prior to enrollment for current and past tobacco product use. Participants are characterized and assigned a use group based on self-reported patterns collected during the study visits.

Overall Study Procedure

Study participants fast for 8 h from food and 6 h from tobacco prior to the visit. All study visits occur before 11AM to limit effects due to circadian changes. All vascular function studies are completed after 10 min of supine positioning. All vascular studies are sent to the BU central lab for analysis. BU biologic samples have minimal processing and are shipped overnight to the UofL central laboratory at the completion of each study visit. Samples obtained at UofL are processed to a similar stage, then held overnight prior to analysis for standardization of time to measurement for all samples.

Study visits include a structured interview on demographics, socioeconomics, lifestyle, health, family history of heart disease, allergies, and tobacco use. All surveys are collected and kept in Research Electronic Data Capture (REDCap), a secure web application for building and managing online surveys and databases.

Exposure Variables

Tobacco Product Use & Particulate Matter Exposure

Comprehensive tobacco product exposure is assessed using a modified version of the National Health Interview survey on tobacco use²⁶. The survey is modified to include detailed information on electronic nicotine devices (ENDs) and other new or emerging tobacco products. Residential addresses are collected for assessment of ambient airborne particulate matter (PM_{2.5}) exposure and future correction of overall exposure. PM_{2.5} data from the day of the study visit, and 3 and 5 days prior to the study is collected from publicly available data associated with EPA monitoring stations. Other exposure variables, including occupation, are collected through interview.

VOC Measurements

Standard clean catch urine specimens are obtained from participants. We have developed a robust Core Lab that utilizes mass spectrometry procedures adopted from the Centers for Disease Control and Prevention (CDC) protocols, to quantify 23 urinary metabolites of tobacco smoking related toxins (aldehydes and other VOCs), including acrolein²⁷ (**Table 2**). The concentration values of analytes are then normalized to urinary creatinine levels measured using Infinity Creatinine Reagent (Thermo Fisher Scientific, MA) on a COBAS MIRA-plus analyzer (Roche, NJ).

Table 2 Exposure Variables (Please see end of article)

| <i>Parent compound</i> | <i>VOC metabolite</i> | <i>Common abbr.</i> |
|-------------------------------|---|----------------------------|
| Acetaldehyde | Acetic acid/Acetate | ACETATE |
| Acrolein | N-Acetyl-S-(2-carboxyethyl)-L-cysteine | CEMA |
| | N-Acetyl-S-(3-hydroxypropyl)-L-cysteine | 3HPMA |
| Acrylamide | N-Acetyl-S-(2-carbamoyl-ethyl)-L-cysteine | AAMA |

| | | |
|---|--|---------|
| | N-Acetyl-S-(2-carbamoyl-2-hydroxyethyl)-L-cysteine | GAMA |
| Acrylonitrile | N-Acetyl-S-(2-cyanoethyl)-L-cysteine | CYMA |
| Acrylonitrile, vinyl chloride, ethylene oxide | N-Acetyl-S-(2-hydroxyethyl)-L-cysteine | HEMA |
| Anabasine | Anabasine (free) | ANB |
| Anatabine | Anatabine (free) | ANTB |
| Benzene | N-Acetyl-S-(phenyl)-L-cysteine | PMA |
| | trans, trans-Muconic acid | MU |
| 1-Bromopropane | N-Acetyl-S-(n-propyl)-L-cysteine | BPMA |
| 1,3-Butadiene | N-Acetyl-S-(3,4-dihydroxybutyl)-L-cysteine | DHBMA |
| | N-Acetyl-S-(1-hydroxymethyl-2-propenyl)-L-cysteine | MHBMA1 |
| | N-Acetyl-S-(2-hydroxy-3-butenyl)-L-cysteine | MHBMA2 |
| | N-Acetyl-S-(4-hydroxy-2-buten-1-yl)-L-cysteine | MHBMA3 |
| Carbon-disulfide | 2-Thioxothiazolidine-4-carboxylic acid | TTCA |
| Crotonaldehyde | N-Acetyl-S-(3-hydroxypropyl-1-methyl)-L-cysteine | HPMMA |
| Cyanide | 2-Aminothiazoline-4-carboxylic acid | ATCA |
| N,N-Dimethylformamide | N-Acetyl-S-(N-methylcarbamoyl)-L-cysteine | AMCC |
| Ethylbenzene, styrene | Phenylglyoxylic acid | PGA |
| Formaldehyde | Formate | FORMATE |
| Nicotine | Nicotine | NIC |
| | Cotinine | COT |
| | 3-Hydroxycotinine | 3HC |

| | | |
|---------------------|---|------------|
| Propylene oxide | N-Acetyl-S-(2-hydroxypropyl)-L-cysteine | 2HPMA |
| Styrene | N-Acetyl-S-(1-phenyl-2-hydroxyethyl)-L-cysteine + | PHEMA |
| | N-Acetyl-S-(2-phenyl-2-hydroxyethyl)-L-cysteine | |
| | Mandelic acid | MA |
| Tetrachloroethylene | N-Acetyl-S-(trichlorovinyl)-L-cysteine | TCVMA |
| Toluene | N-Acetyl-S-(benzyl)-L-cysteine | BMA |
| Trichloroethylene | N-Acetyl-S-(1,2-dichlorovinyl)-L-cysteine | 1,2DCVMA |
| | N-Acetyl-S-(2,2-dichlorovinyl)-L-cysteine | 2,2DCVMA |
| Xylene | N-Acetyl-S-(2,4-dimethylphenyl)-L-cysteine + | DPMA |
| | N-Acetyl-S-(2,5-dimethylphenyl)-L-cysteine + | |
| | N-Acetyl-S-(3,4-dimethylphenyl)-L-cysteine | |
| | 2-Methylhippuric acid | 2MHA |
| | 3-Methylhippuric acid + 4-Methylhippuric acid | 3MHA+ 4MHA |

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174 *Urine is analyzed for 23 metabolites of 18 parent VOCs and tobacco alkaloids by UPLC-*
 175 *MS/MS. Analytes are listed as parent, metabolite and their common abbreviation.*

176

177 **Circulating Markers of Cardiovascular Injury**

178 To assess tobacco product-induced cardiovascular toxicity, we examine
 179 endothelial function, inflammatory mediators, biomarkers, and thrombosis. CV risk is
 180 defined through measurements of circulating angiogenic cells, lipid profile, and glucose
 181 metabolism^{25 28 29}. Plasma (BD367863 and BD366415) and serum (BD367814)
 182 samples are obtained from all participants for laboratory testing and long term

biobanking. Whole blood (BD366415) is obtained for flow cytometry on fresh samples at UofL pathology core. BU biologic samples have minimal processing and are shipped overnight to the UofL central laboratory at the completion of each study visit. Samples obtained at UofL are processed to a similar stage, then held overnight prior to analysis to standardize the time to measurement for all samples. The UofL central laboratory, as previously reported, will complete fasting and biomarker measurements (**Table 3**), with the exception of cytomics^{13 30}. For cytomic measurements, mononuclear cells are labeled with the peripheral blood phenotyping panel kit (Fluidigm). Samples are shipped at 4 degree C to Core Lab facilities at the University of Rochester for Mass cytometric analysis.

Table 3 Blood analysis

| |
|---|
| <u>Fasting Measurements</u> |
| LDL cholesterol, HDL cholesterol, total cholesterol, triglycerides, glucose, uric acid, SAA and fibrinogen |
| <u>Biomarkers</u> |
| CAC (1-15) ¹ , Platelet-monocyte aggregates, MP (1-5) ¹ , PF4, t-PA, TxA2, Factor VII, IL-6, CRP, D-dimer, PAI-1, s-ICAM-1, s-VCAM, s-thrombomodulin, s-TNFR1, MMP-2, MMP-3, MMP-9, cytomics, endothelin, E-selectin and P-selectin |
| 1: Fifteen different CAP subpopulations and 5 subtypes of microparticles were measured by flow cytometry. |

All participants who complete the study visit will have blood samples taken and processed. Flow cytometric analysis is completed on fresh samples. All other analysis will be completed on biobanked samples in batches LDL= low density lipoprotein. HDL= high density lipoprotein. SAA= serum amyloid A. CAC= circulating angiogenic cells. MP= microparticles. PF4= Platelet factor 4. t-PA= tissue plasminogen activator. TxA2=Thromboxane A. IL-6= Interleukin 6. CRP= C-reactive protein. PAI=- Plasminogen activator. s- ICAM- soluble intercellular adhesion protein inhibitor. s-

201 *VCAM= soluble vascular adhesion protein. TNFR1= Tumor necrosis factor receptor 1.*

202 *MMP- Matrix metalloproteinase.*

203 **Non-Invasive Vascular Function Testing**

204 Smoking, is associated with endothelial damage and vascular dysfunction^{31 32}.

205 Endothelial cells are exposed to circulating toxins and measures of endothelial function
206 are reflective of cardiovascular injury³³. Thus, we examine the non-invasive endothelial
207 vasodilator function using flow-mediated vasodilation^{34 35}, arterial stiffness with carotid-
208 femoral and carotid-radial pulse wave velocity³⁶, and peripheral vascular function with
209 ankle brachial index. All vascular imagers were trained at BU. Similar equipment and
210 software is used at both sites. All vascular studies are sent to the BU central lab for
211 analysis.

212 **Anthropometric measures**

213 Anthropometric measures included height, weight, waist and hip circumference
214 and body fat. All anthropometric measures are completed twice and the average
215 recorded. Standing height measurements are completed on a fixed stadiometer. Weight
216 measurements are completed on a digital scale to the nearest tenth of a pound. Waist
217 circumference is measured at the level of the umbilicus to the nearest tenth of a
218 centimeter. Hip circumference is measured at the maximal protrusion of the gluteal
219 muscle to the nearest tenth of a centimeter. Body fat percentage is calculated by the
220 bioelectrical impedance measured with the Omron fat loss monitor (HBF-306C).

221 **DATA ANALYSIS**

222 We expect that from this study we will be able to identify specific biomarkers of
223 cardiovascular injury due to tobacco use and the relationship of these biomarkers to

specific measures of tobacco exposure. For instance, we will identify which biomarkers are affected by tobacco use, and which ones are most sensitive; including their dose-dependence. Additionally we will examine the extent to which biomarkers are associated with exposure to nicotine versus exposure to HPHC of tobacco like aldehydes.

All statistical analysis will be performed using SAS version 9.4 software (SAS Institute, Inc., Cary, North Carolina), and a two-sided p-value of <0.05 will be considered significant for any statistical test. Demographics and other baseline characteristics will be summarized according to product group. The primary outcomes will be analyzed using multiple regression techniques. Appropriate Interaction variables will be tested for in the regression models and subgroup analyses will be conducted according to the following factors: significant interactions, sex, age, race, tobacco product group. Multiple imputation method will be used for missing data where appropriate. Sensitivity analysis using different analytic approaches, such as generalized linear models, as well as considering different covariate adjustments, will be used to build concordant results.

The dose-dependence of the changes in biomarkers will be determined by analyzing the data obtained from individuals that are exposed to different doses of a single product (e.g. smoking 0, <15, 15-20 and >20 cigarettes per day) and by comparing between tobacco products that have different doses of HPHC constituents. In the US the average cigarettes per day is between 15-20³⁷ and therefore this dose range distribution is reflective of general population exposure. Comparisons of the effects of novel tobacco products and smoking will be informative of the relative toxicity of the two products.

We believe that the methods employed in the current project are exquisitely sensitive and responsive to even low dose insults such as ambient air pollution¹³ allowing us to quantify tobacco product-induced changes with high precision. Moreover, levels of acrolein exposure vary between different individuals due to difference in puffing intensity and the time a cigarette is left smoldering. Thus, direct measurements of acrolein metabolites afford better estimates of acrolein exposure than machine yields. We expect to obtain wide variations in acrolein/crotonaldehyde exposure which will enable us to construct a dose-response relationship and identify which injury biomarkers are associated with aldehyde exposure and whether high levels of exposure are associated with high levels of injury, despite similar nicotine delivery.

We consider three major factors for balancing sample selection: age, gender, and race. Given that very few females use e-cigarette, only males will be enrolled in this group. With the balanced design to determine the main effects and interactions in selected scenarios, we justify the sample size. The analysis plan is primarily based on evaluating the effect of tobacco exposure on endothelial function (FMD), and the main biomarkers, EPCs, and platelet-monocyte aggregates (PMA). The sample size is justified in terms of the primary dependent measure, FMD, given the potential importance of this variable as a direct measure of the impact of tobacco exposure. The main comparisons are between non-tobacco users and tobacco users. Due to one control group, we will conservatively adjust our α (significance level) using a Bonferroni correction, and we will set $\alpha=0.01$. Based on preliminary data for FMD, we have observed mean \pm SD in smoker and nonsmoker groups to be 4.0 ± 1.6 and 6.8 ± 1.0 , respectively. We consider at least 25% (mean FMD=3.0 from 4.0) reduction from

smokers to non-smokers is meaningful. Using a two sample, one-sided t test with an α of 0.01 and 80% power (1- β), assuming a common SD of 1.3, we will need 34 evaluable subjects in each group. To examine dose response, smokers will be recruited in 3 groups (<15, 15-20 and >20 CPD). We will recruit 40 participants in each group; total group size = 120 participants. In **Table 4** we provide estimable effect size for different outcome measures.

Table 4 Minimal Detectable Differences in Endpoints at $\alpha=0.01$ and Power=80%

| Variable | Non-smokers | Smokers | n | p | Ref | MDD |
|-----------------------------|---------------------|----------------------|----|-------|-----|----------------------|
| Primary Functional Outcome | | | | | | 278 |
| FMD | 6.8 \pm 1% | 4.0 \pm 1.6% | 10 | <0.05 | 32 | 1.0 ²⁷⁹ |
| Primary Biomarkers | | | | | | 280 |
| EPC | 25 \pm 5 cell/ml | 10 \pm 3 cells/ml | 24 | 0.037 | 38 | 3.1 ²⁸¹ |
| PMA | 19.7 \pm 8.6% | 26.6 \pm 9% | 25 | 0.02 | 39 | 7.0 ²⁸² |
| EMP | 1.1 \pm 0.4 | 0.5 \pm 0.2 | 32 | <0.05 | 40 | 0.23 ²⁸³ |
| Other Biochemical Variables | | | | | | 284 |
| PF4 | 3.9 \pm 1.2 IU/ml | 5.0 \pm 2.6 IU/ml | 12 | <0.05 | 41 | 2.0 ²⁸⁵ |
| tPA | 3.0 \pm 0.6 ng/ml | 4.3 \pm 2.0 ng/ml | 20 | <0.05 | 42 | 1.6 ²⁸⁶ |
| TxA ₂ | 2.2 \pm 0.1 pg/ml | 3.3 \pm 0.02 pg/ml | 12 | <0.05 | 43 | 0.016 ²⁸⁷ |

PMA: Platelet
- monocyte

aggregates; EMP: Endothelial microparticles (CD62+/CD31+); MDD: minimal detectable difference. Values are mean \pm SD

ETHICS AND DISSEMINATION

The CITU study was approved at each institution by their institutional review board (BU #H-32613 and UofL #13.0590) and all participants provide written consent. No study related procedures will be completed until after participant consent.

Participants for the CITU study are being recruited in both Boston, MA and Louisville KY. The two populations show significant differences, therefore recruitment at two sites will ensure a range more reflective of the general population. Although overall racial and ethnic demographics for both cities show a clear majority of Caucasians (70%) and despite smokers typically male, we strive to, and currently are successful in, recruiting a population that was gender balanced and almost evenly split between Caucasian and African Americans. Despite this balanced recruitment, e-cigarette users have been reported as predominantly Caucasian and male⁴⁴, and thus far our recruitment mirrors these demographics. We expect very few Hispanic/Latino's to participate, due to data suggesting tobacco use, including ENDS, tends to be lower among Hispanic's/Latino's^{44 45}. Thus we have also opted to only recruit English speakers. We have carefully develop our recruitment strategy and exclusion criteria to protect vulnerable populations, which is important since many report a lower socioeconomic status and educational level in smokers in addition to higher rates of reported alcohol and drug use^{46 47}.

Our study is an observational study where participants have already assumed the risk of using tobacco. Study procedures pose minimal risk. Given the known harms associated with smoking, we will provide information on tobacco treatment when requested by the participant. Participant information is de-identified for analysis and reported in aggregate to protect privacy.

Completion of these studies will enable a greater understanding of the biological responses to use of a variety of tobacco products. Specifically, they will help to identify the constituents of these products; and how a panel of exposure and CV injury biomarkers are associated with these different constituents. This data will be available to the FDA and could help guide new policy measures to reduce or eliminate the harmful components of tobacco smoke and other nicotine products. The study is dedicated to the rapid dissemination of their rigorously characterized and well-controlled research findings to the public in the form of peer-reviewed publications. Subsequent to the initial full-length manuscript publications of the resources generated with funding from this program, the study will make them available to interested and qualified investigators upon written request. The study will provide relevant protocols of published data, upon request (presuming prior publication by the Center members). Participants will be provided a summary of the results as they become available. Finally press releases of relevant findings will inform the general population.

LIST OF ABBREVIATIONS

ABI- Ankle Brachial Index
CAC= circulating angiogenic cells
CRP= C-reactive protein
CVD- Cardiovascular disease
ENDS- Electronic nicotine Device (i.e. e-cigarette)
FACS- Fluorescence-activated cell sorting
FMD- Flow mediated dilation

338 HDL= high density lipoprotein

339 IL-6= Interleukin 6

340 MMP- Matrix metalloproteinase

341 MP= micoparticles

342 PAI=- Plasminogen activator

343 PF4= Platelet factor 4

344 PWV- Pulse wave velocity

345 SAA= serum amyloid A

346 s-ICAM- soluble intercellular adhesion protein inhibitor

347 s-VCAM= soluble vascular adhesion protein

348 TNFR1= Tumor necrosis factor receptor 1

349 t-PA= tissue plasminogen activator

350 TxA2=Thromboxane A

351 VOC- Volatile organic compound

352 W:H- ratio: Waist to hip ratio

353

354 **AUTHORS CONTRIBUTIONS**

355 Rachel Keith- Study design, study recruitment, study visits, statistical analysis and
356 manuscript preparation. Jessica Fetterman- study recruitment, study visits, manuscript
357 preparation and editing. Dan Riggs- statistical analysis, manuscript preparation and
358 editing. Tim O'Toole- Biomarker measurements, manuscript preparation and editing.
359 Jessica Nystoriak- study recruitment and study visits. Monica Holbrook- study
360 recruitment and study visits. Pawel Lorkiewicz- VOC measurements and manuscript

preparation. Aruni Bhatnagar- Study design, study funding and manuscript editing.

Andrew DeFilippis- Human subject assessment planning, manuscript preparation and editing. Naomi M. Hamburg- Study design, study funding, vascular core, manuscript preparation and editing.

COMPETING INTERESTS

None declared

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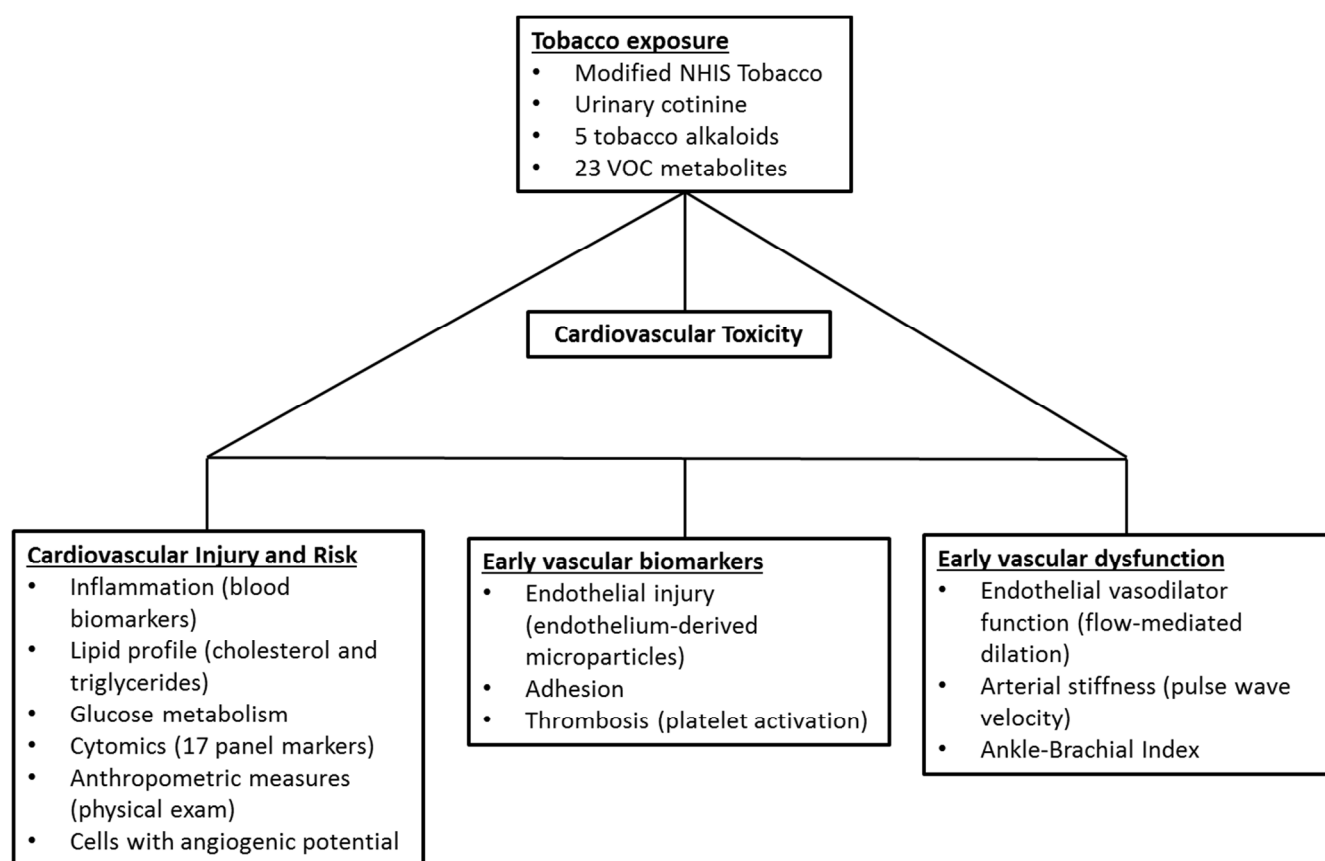
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Figure 1.



STROBE 2007 (v4) Statement—Checklist of items that should be included in reports of *cohort studies*

| Section/Topic | Item # | Recommendation | Reported on page # |
|---------------------------|--------|--|----------------------|
| Title and abstract | 1 | (a) Indicate the study’s design with a commonly used term in the title or the abstract | 1 |
| | | (b) Provide in the abstract an informative and balanced summary of what was done and what was found | 2-3 |
| Introduction | | | |
| Background/rationale | 2 | Explain the scientific background and rationale for the investigation being reported | 3-4 |
| Objectives | 3 | State specific objectives, including any prespecified hypotheses | 5 |
| Methods | | | |
| Study design | 4 | Present key elements of study design early in the paper | 5, 7 |
| Setting | 5 | Describe the setting, locations, and relevant dates, including periods of recruitment, exposure, follow-up, and data collection | 5, 7 |
| Participants | 6 | (a) Give the eligibility criteria, and the sources and methods of selection of participants. Describe methods of follow-up | 6-7 |
| | | (b) For matched studies, give matching criteria and number of exposed and unexposed | N/A |
| Variables | 7 | Clearly define all outcomes, exposures, predictors, potential confounders, and effect modifiers. Give diagnostic criteria, if applicable | 7-12 |
| Data sources/ measurement | 8* | For each variable of interest, give sources of data and details of methods of assessment (measurement). Describe comparability of assessment methods if there is more than one group | 7-12 |
| Bias | 9 | Describe any efforts to address potential sources of bias | 7 |
| Study size | 10 | Explain how the study size was arrived at | 14-16 |
| Quantitative variables | 11 | Explain how quantitative variables were handled in the analyses. If applicable, describe which groupings were chosen and why | 12-14 |
| Statistical methods | 12 | (a) Describe all statistical methods, including those used to control for confounding | 12-14 |
| | | (b) Describe any methods used to examine subgroups and interactions | 13 |
| | | (c) Explain how missing data were addressed | 13 |
| | | (d) If applicable, explain how loss to follow-up was addressed | N/A (study protocol) |
| | | (e) Describe any sensitivity analyses | 13 |
| Results | | | |

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|--------------------------|-----|--|----------------------|
| Participants | 13* | (a) Report numbers of individuals at each stage of study—eg numbers potentially eligible, examined for eligibility, confirmed eligible, included in the study, completing follow-up, and analysed | N/A (study protocol) |
| | | (b) Give reasons for non-participation at each stage | N/A (study protocol) |
| | | (c) Consider use of a flow diagram | N/A (study protocol) |
| Descriptive data | 14* | (a) Give characteristics of study participants (eg demographic, clinical, social) and information on exposures and potential confounders | N/A (study protocol) |
| | | (b) Indicate number of participants with missing data for each variable of interest | N/A (study protocol) |
| | | (c) Summarise follow-up time (eg, average and total amount) | N/A (study protocol) |
| Outcome data | 15* | Report numbers of outcome events or summary measures over time | N/A (study protocol) |
| Main results | 16 | (a) Give unadjusted estimates and, if applicable, confounder-adjusted estimates and their precision (eg, 95% confidence interval). Make clear which confounders were adjusted for and why they were included | N/A (study protocol) |
| | | (b) Report category boundaries when continuous variables were categorized | N/A (study protocol) |
| | | (c) If relevant, consider translating estimates of relative risk into absolute risk for a meaningful time period | N/A (study protocol) |
| Other analyses | 17 | Report other analyses done—eg analyses of subgroups and interactions, and sensitivity analyses | N/A (study protocol) |
| Discussion | | | |
| Key results | 18 | Summarise key results with reference to study objectives | N/A (study protocol) |
| Limitations | | | |
| Interpretation | 20 | Give a cautious overall interpretation of results considering objectives, limitations, multiplicity of analyses, results from similar studies, and other relevant evidence | 17 |
| Generalisability | 21 | Discuss the generalisability (external validity) of the study results | |
| Other information | | | |
| Funding | 22 | Give the source of funding and the role of the funders for the present study and, if applicable, for the original study on which the present article is based | 19 |

*Give information separately for cases and controls in case-control studies and, if applicable, for exposed and unexposed groups in cohort and cross-sectional studies.

Note: An Explanation and Elaboration article discusses each checklist item and gives methodological background and published examples of transparent reporting. The STROBE checklist is best used in conjunction with this article (freely available on the Web sites of PLoS Medicine at <http://www.plosmedicine.org/>, Annals of Internal Medicine at <http://www.annals.org/>, and Epidemiology at <http://www.epidem.com/>). Information on the STROBE Initiative is available at www.strobe-statement.org.

BMJ Open

Protocol to Assess the Impact of Tobacco-Induced Volatile Organic Compounds on Cardiovascular Risk in a Cross-Sectional Cohort: Cardiovascular Injury Due to Tobacco Study

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1 **Protocol to Assess the Impact of Tobacco-Induced Volatile Organic Compounds**
2 **on Cardiovascular Risk in a Cross-Sectional Cohort: Cardiovascular Injury Due to**
3 **Tobacco Study**

4 Rachel J. Keith, Jessica L. Fetterman, Daniel W. Riggs, Tim O’Toole, Jessica Nystoriak,
5 Monica Holbrook, Pawel Lorkiewicz, Aruni Bhatnagar, Andrew DeFilippis*, Naomi M.
6 Hamburg*

7 Rachel J. Keith-Division of Cardiovascular Medicine, University of Louisville School of
8 Medicine 580 S. Preston St. Louisville KY, 40202 rachel.keith@louisville.edu 502-852-
9 4211

10 Jessica L. Fetterman- Vascular Biology Section, Whitaker Cardiovascular Institute,
11 Boston University School of Medicine Evans Building, Boston, MA USA

12 Jasmit Shah- University of Louisville School of Medicine Louisville, KY USA

13 Timothy O-Toole- Division of Cardiovascular Medicine, University of Louisville School of
14 Medicine Louisville, KY USA

15 Jessica L. Nystoriak- Division of Cardiovascular Medicine, University of Louisville
16 School of Medicine Louisville, KY USA

17 Monika Holbrook- Vascular Biology Section, Whitaker Cardiovascular Institute, Boston
18 University School of Medicine Boston, MA USA

19 Pawel Lorkiewicz- Division of Cardiovascular Medicine, University of Louisville School
20 of Medicine Louisville, KY USA

21 Aruni Bhatnagar- Division of Cardiovascular Medicine, University of Louisville School of
22 Medicine Louisville, KY USA

Andrew P. DeFilippis- Division of Cardiovascular Medicine, University of Louisville
School of Medicine Louisville, KY USA (co-senior author)

Naomi M. Hamburg- Vascular Biology Section, Whitaker Cardiovascular Institute,
Boston University School of Medicine Boston, MA USA (co-senior author)

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ABSTRACT

Introduction: Tobacco use leads to increased mortality, the majority of which is attributed to cardiovascular disease. Despite this knowledge, the early cardiovascular impact of tobacco product use is not well understood. Tobacco use increases exposure to harmful and potentially harmful constituents including volatile organic compounds (VOCs) such as acrolein and crotonaldehyde, which may contribute to cardiovascular risk. The link between exposure patterns, risk profiles and demographic distribution of tobacco product users, particularly users of new and emerging products, are not well known. Therefore, we designed the Cardiovascular Injury due to Tobacco Use (CITU) study to assess population characteristics, demographic features, exposure patterns and cardiovascular risk in relation to tobacco.

Methods and analysis: We present the design and methodology of the CITU study a cross-section observational tobacco study conducted in Boston MA and Louisville KY starting in 2014. Healthy participants 21 to 45 years of age who use tobacco products, including ENDS, or who never used tobacco are being recruited. The study aims to recruit an evenly split cohort of African Americans and Caucasians that is sex balanced

for evaluation of self-reported tobacco exposure, VOC exposure and tobacco-induced injury profiling. Detailed information about participant’s demographics, health status and lifestyle is also collected.

Ethics and dissemination: The study protocol was approved institutional review boards at both participating universities. All study protocols will protect participant confidentiality. Results from the study will be disseminated via peer-reviewed journals and presented at scientific conferences.

Strengths and limitations

- Young age to allow for evaluation of early stage disease (e.g. inflammation, endothelial function) as opposed to end stage clinical consequence (e.g. myocardial infarction)
- Diverse tobacco product use allows for assessment of a wide range of tobacco-induced VOC exposure
- All study visits are in English introducing selection bias
- Data will inform regulatory agencies on the cardiovascular health effects of multiple tobacco products and the contribution of HPHCs

Keywords: Tobacco, smoking, electronic cigarette, vascular injury, cardiovascular risk, cigarettes.

INTRODUCTION

Tobacco product use and smoking are the leading causes of preventable deaths throughout the world. Of those deaths, one-third are attributed to cardiovascular disease (CVD)¹. The cardiovascular (CV) effects of tobacco exposure can include atherogenesis, vascular injury, thrombosis, arrhythmias and inflammation² and may be attributable to the many different harmful and potentially harmful constituents (HPHCs) present in tobacco products.

The HPHCs found in tobacco products include volatile organic compounds (VOCs) of which reactive aldehydes, such as acrolein and crotonaldehyde, are likely the most significant contributors to CV toxicity³. High levels of aldehydes are present in cigarette smoke^{4 5} as well as smokeless tobacco (ST)⁶. Risk assessments, using the prevalence of each individual chemical weighed by its potency, suggest that the non-cancer risk of smoking is dominated by acrolein, which contributes 40-100 times more to risk than any other chemical present in cigarette smoke³.

Although HPHCs, including VOC reactive aldehydes, have been suspected to be major contributors to the toxicity of cigarette smoke for over 4 decades, their contribution to CV injury and early CVD risk has not been rigorously evaluated. Experimental studies in animal models suggest that because of low aldehyde-metabolizing capacity, CV tissues are highly sensitive to aldehydes and exposure to low levels of aldehydes can induce CV injury and accelerate CVD⁷⁻¹⁸. The WHO Study Group on Tobacco Product Regulation (TobReg) has marked acrolein, a VOC, along with 8 other cigarette constituents for monitoring and regulation¹⁹ and the U.S. Environmental Protection Agency lists Acrolein as one of most hazardous air pollutants²⁰. Nevertheless, the contribution of tobacco induced VOCs, including acrolein

or other aldehydes, toward CV toxicity in humans has not been fully assessed. Greater understanding of how aldehydes affect cardiovascular health and disease will provide new avenues for evaluating the toxicity of cigarette smoke and for assessing the injurious potential of new and emerging tobacco products, such as ENDS, which may also contain VOCs including acrolein²¹⁻²³.

The latency period between tobacco exposure and the development of major clinical adverse health effects is long, therefore biomarkers that provide information over a shorter period allow for the identification of harm decades before clinical outcome data is available. Thus, in this paper we present the design and methodology of the Cardiovascular Injury due To Tobacco Use (CITU) study which will evaluate the association of the urinary metabolites of 18 parent VOCs from tobacco exposure with a comprehensive set of CV biomarkers representative of early disease and predictive of future CV events.²⁴

METHODS AND DESIGN

Overall design

The CITU study is an investigator-initiated cross-sectional observational study of around 500 healthy participants 21 to 45 years of age who are never or current tobacco product users in two urban areas at Boston University (BU) and University of Louisville (UofL) (Boston, MA and Louisville, KY) designed to evaluate CV toxicity due to tobacco product use, with correlations to VOCs found in the tobacco products (Figure 1).

Participant Eligibility Criteria

The goal of the study is to examine the impact of tobacco products on healthy young adults who could be classified as a current tobacco product users (Defined in table 1), or never-users (does not have lifetime use of any tobacco product). Participants were self-reported to be healthy therefore we excluded participants if they had: 1) diagnosis of clinical cardiovascular disease including but not limited to known heart attack, peripheral artery disease, heart failure or stroke; 2) diagnosis of diabetes (HbA1c >7.0 or treatment for diabetes), hypertension (systolic blood pressure >140 mm Hg or diastolic blood pressure >90 mm Hg), hypothyroidism or hyperthyroidism, inflammatory conditions such as lupus or inflammatory bowel disease, HIV/AIDS, hepatitis, liver disease, anemia, cancer of any type or another medical condition that might compromise the successful completion of the study; 2) recipients of organ transplant or renal replacement therapy; 3) individuals that are taking the following medications: immunosuppressant agents estrogen, testosterone, anti TNF agents, certain biologics, Procrit, statins, beta-blockers or other cardiovascular medicine; 4) individuals using nutraceuticals or anabolic steroids beyond the recommended daily allowance; 5) body weight less than 100 pounds; 6) pregnant women; 7) prisoners and other vulnerable populations; and 8) active illness or infection. Participants are rescheduled or considered screen-failures and excluded from the study if symptomatic of an acute illness, i.e. viral upper respiratory infection, on study date.

Table 1. Tobacco product use classifications

| Classification | Qualification |
|------------------------|---|
| Never | Does not meet lifetime limits for any tobacco use (see below) |
| Smoker | >100 lifetime cigarettes and current use for the past year |
| Smokeless Tobacco User | >20 lifetime dips or chews and current use for the past year |
| Cigar/Cigarillo User | >20 lifetime cigars or cigarillos and current use for the past year |
| Pipe User | >20 lifetime pipefuls and current use for the past year |

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| ENDS User | >20 lifetime vape sessions and current use for the past year |
| Hookah User | >20 lifetime hookah sessions and current use for the past year |

Study participants are screened prior to enrollment for current and past tobacco product use. Participants are characterized and assigned a use group based on self-reported patterns collected during the study visits.

Overall Study Procedure

Study participants fast for 8 h from food and 6 h from tobacco prior to the visit. All study visits occur before 11AM to limit effects due to circadian changes. All vascular function studies are completed after 10 min of supine positioning. All vascular studies are sent to the BU central lab for analysis. BU biologic samples have minimal processing and are shipped overnight to the UofL central laboratory at the completion of each study visit. Samples obtained at UofL are processed to a similar stage, then held overnight prior to analysis for standardization of time to measurement for all samples.

Study visits take approximately 90 minutes to complete and include a structured interview on demographics, socioeconomics, lifestyle, health, family history of heart disease, allergies, and tobacco use. (Figure 2) Participants were compensated appropriately for their time. All surveys are collected and kept in Research Electronic Data Capture (REDCap), a secure web application for building and managing online surveys and databases.

Exposure Variables

Tobacco Product Use & Particulate Matter Exposure

Comprehensive tobacco product exposure is assessed using a modified version of the National Health Interview survey on tobacco use²⁵. The survey is modified to include detailed information on electronic nicotine devices (ENDs) and other new or

emerging tobacco products. Residential addresses are collected for assessment of ambient airborne particulate matter (PM_{2.5}) exposure and future correction of overall exposure. PM_{2.5} data from the day of the study visit, and 3 and 5 days prior to the study is collected from publicly available data associated with EPA monitoring stations. Other exposure variables, including occupation, are collected through interview.

VOC Measurements

Standard clean catch urine specimens are obtained from participants. We have developed a robust Core Lab that utilizes mass spectrometry procedures adopted from the Centers for Disease Control and Prevention (CDC) protocols, to quantify 23 urinary metabolites of tobacco smoking related toxins (aldehydes and other VOCs), including acrolein²⁶ (**Table 2**). The concentration values of analytes are then normalized to urinary creatinine levels measured using Infinity Creatinine Reagent (Thermo Fisher Scientific, MA) on a COBAS MIRA-plus analyzer (Roche, NJ).

Table 2 Exposure Variables (Please see end of article)

| <i>Parent compound</i> | <i>VOC metabolite</i> | <i>Common abbr.</i> |
|------------------------|--|---------------------|
| Acetaldehyde | Acetic acid/Acetate | ACETATE |
| Acrolein | N-Acetyl-S-(2-carboxyethyl)-L-cysteine | CEMA |
| | N-Acetyl-S-(3-hydroxypropyl)-L-cysteine | 3HPMA |
| Acrylamide | N-Acetyl-S-(2-carbamoyl-ethyl)-L-cysteine | AAMA |
| | N-Acetyl-S-(2-carbamoyl-2-hydroxyethyl)-L-cysteine | GAMA |
| Acrylonitrile | N-Acetyl-S-(2-cyanoethyl)-L-cysteine | CYMA |

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|---|--|---------|
| Acrylonitrile, vinyl chloride, ethylene oxide | N-Acetyl-S-(2-hydroxyethyl)-L-cysteine | HEMA |
| Anabasine | Anabasine (free) | ANB |
| Anatabine | Anatabine (free) | ANTB |
| Benzene | N-Acetyl-S-(phenyl)-L-cysteine | PMA |
| | trans, trans-Muconic acid | MU |
| 1-Bromopropane | N-Acetyl-S-(n-propyl)-L-cysteine | BPMA |
| 1,3-Butadiene | N-Acetyl-S-(3,4-dihydroxybutyl)-L-cysteine | DHBMA |
| | N-Acetyl-S-(1-hydroxymethyl-2-propenyl)-L-cysteine | MHBMA1 |
| | N-Acetyl-S-(2-hydroxy-3-butenyl)-L-cysteine | MHBMA2 |
| | N-Acetyl-S-(4-hydroxy-2-buten-1-yl)-L-cysteine | MHBMA3 |
| Carbon-disulfide | 2-Thioxothiazolidine-4-carboxylic acid | TTCA |
| Crotonaldehyde | N-Acetyl-S-(3-hydroxypropyl-1-methyl)-L-cysteine | HPMMA |
| Cyanide | 2-Aminothiazoline-4-carboxylic acid | ATCA |
| N,N-Dimethylformamide | N-Acetyl-S-(N-methylcarbamoyl)-L-cysteine | AMCC |
| Ethylbenzene, styrene | Phenylglyoxylic acid | PGA |
| Formaldehyde | Formate | FORMATE |
| Nicotine | Nicotine | NIC |
| | Cotinine | COT |
| | 3-Hydroxycotinine | 3HC |
| Propylene oxide | N-Acetyl-S-(2-hydroxypropyl)-L-cysteine | 2HPMA |
| Styrene | N-Acetyl-S-(1-phenyl-2-hydroxyethyl)-L-cysteine + | PHEMA |

| | | |
|---------------------|--|------------|
| | N-Acetyl-S-(2-phenyl-2-hydroxyethyl)-L-cysteine | |
| | Mandelic acid | MA |
| Tetrachloroethylene | N-Acetyl-S-(trichlorovinyl)-L-cysteine | TCVMA |
| Toluene | N-Acetyl-S-(benzyl)-L-cysteine | BMA |
| Trichloroethylene | N-Acetyl-S-(1,2-dichlorovinyl)-L-cysteine | 1,2DCVMA |
| | N-Acetyl-S-(2,2-dichlorovinyl)-L-cysteine | 2,2DCVMA |
| Xylene | N-Acetyl-S-(2,4-dimethylphenyl)-L-cysteine + N-Acetyl-S-(2,5-dimethylphenyl)-L-cysteine + N-Acetyl-S-(3,4-dimethylphenyl)-L-cysteine | DPMA |
| | 2-Methylhippuric acid | 2MHA |
| | 3-Methylhippuric acid + 4-Methylhippuric acid | 3MHA+ 4MHA |
| | | |

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171 *Urine is analyzed for 23 metabolites of 18 parent VOCs and tobacco alkaloids by UPLC-*
 172 *MS/MS. Analytes are listed as parent, metabolite and their common abbreviation.*

173

174 **Circulating Markers of Cardiovascular Injury**

175 To assess tobacco product-induced cardiovascular toxicity, we examine
 176 endothelial function, inflammatory mediators, biomarkers, and thrombosis. CV risk is
 177 defined through measurements of circulating angiogenic cells, lipid profile, and glucose
 178 metabolism^{24 27 28}. Plasma (BD367863 and BD366415) and serum (BD367814)
 179 samples are obtained from all participants for laboratory testing and long term
 180 biobanking. Whole blood (BD366415) is obtained for flow cytometry on fresh samples at
 181 UofL pathology core. BU biologic samples have minimal processing and are shipped

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overnight to the UofL central laboratory at the completion of each study visit. Samples
obtained at UofL are processed to a similar stage, then held overnight prior to analysis
to standardize the time to measurement for all samples. The UofL central laboratory, as
previously reported, will complete fasting and biomarker measurements (**Table 3**), with
the exception of cytomics^{12 29}. For cytomic measurements, mononuclear cells are
labeled with the peripheral blood phenotyping panel kit (Fluidigm). Samples are shipped
at 4 degree C to Core Lab facilities at the University of Rochester for Mass cytometric
analysis.

Table 3 Blood analysis

| |
|---|
| <u>Fasting Measurements</u> |
| LDL cholesterol, HDL cholesterol, total cholesterol, triglycerides, glucose, uric acid, SAA and fibrinogen |
| <u>Biomarkers</u> |
| CAC (1-15) ¹ , Platelet-monocyte aggregates, MP (1-5) ¹ , PF4, t-PA, TxA2, Factor VII, IL-6, CRP, D-dimer, PAI-1, s-ICAM-1, s-VCAM, s-thrombomodulin, s-TNFR1, MMP-2, MMP-3, MMP-9, cytomics, endothelin, E-selectin and P-selectin |
| 1: Fifteen different CAP subpopulations and 5 subtypes of microparticles were measured by flow cytometry. |

All participants who complete the study visit will have blood samples taken and processed. Flow cytometric analysis is completed on fresh samples. All other analysis will be completed on biobanked samples in batches LDL= low density lipoprotein. HDL= high density lipoprotein. SAA= serum amyloid A. CAC= circulating angiogenic cells. MP= microparticles. PF4= Platelet factor 4. t-PA= tissue plasminogen activator. TxA2=Thromboxane A. IL-6= Interleukin 6. CRP= C-reactive protein. PAI=- Plasminogen activator. s- ICAM- soluble intercellular adhesion protein inhibitor. s- VCAM= soluble vascular adhesion protein. TNFR1= Tumor necrosis factor receptor 1. MMP- Matrix metalloproteinase.

Non-Invasive Vascular Function Testing

Smoking, is associated with endothelial damage and vascular dysfunction^{30 31}. Endothelial cells are exposed to circulating toxins and measures of endothelial function are reflective of cardiovascular injury³². Thus, we examine the non-invasive endothelial vasodilator function using flow-mediated vasodilation^{33 34}, arterial stiffness with carotid-femoral and carotid-radial pulse wave velocity³⁵, and peripheral vascular function with ankle brachial index. Flow mediated dilation was assessed with a 7.5MHZ ultrasound probe is used to image the brachial artery while a 10cm blood pressure cuff is attached

to the lower arm and a 3 lead ECG is attached to the patient. After baseline images and 10 cycles of Doppler images are captured using NIHEM R-wave triggered image capturing software, the blood pressure cuff is inflated to 200mmHg or 50mmHg higher than the systolic pressure. After the 5 minute occlusion, the cuff is released and the NIHEM software records two minutes of imaging. Images were analyzed by a single blinded analyzer using MIA vascular Research Tolls Brachial Analyzer for Research, version 6.8.5. All vascular imagers where trained at BU who have a previously reported reproducibility with intra- and inter-observer correlation coefficients of 0.98 and 0.99 for brachial diameter and 0.78 and 0.92 for FMD.³⁶ Similar equipment and software is used at both sites. All vascular studies are sent to the BU central lab for analysis.

Anthropometric measures

Anthropometric measures included height, weight, waist and hip circumference and body fat. All anthropometric measures are completed twice and the average recorded. Standing height measurements are completed on a fixed stadiometer. Weight measurements are completed on a digital scale to the nearest tenth of a pound. Waist circumference is measured at the level of the umbilicus to the nearest tenth of a centimeter. Hip circumference is measured at the maximal protrusion of the gluteal muscle to the nearest tenth of a centimeter. Body fat percentage is calculated by the bioelectrical impedance measured with the Omron fat loss monitor (HBF-306C).

DATA ANALYSIS

We expect that from this study we will be able to identify specific biomarkers of cardiovascular injury due to tobacco use and the relationship of these biomarkers to specific measures of tobacco exposure. For instance, we will identify which biomarkers

are affected by tobacco use, and which ones are most sensitive; including their dose-dependence. Additionally we will examine the extent to which biomarkers are associated with exposure to nicotine versus exposure to HPHC of tobacco like aldehydes.

All statistical analysis will be performed using SAS version 9.4 software (SAS Institute, Inc., Cary, North Carolina), and a two-sided p-value of <0.05 will be considered significant for any statistical test. Demographics and other baseline characteristics will be summarized according to product group. The primary outcomes will be analyzed using multiple regression techniques. Appropriate Interaction variables will be tested for in the regression models and subgroup analyses will be conducted according to the following factors: significant interactions, sex, age, race, tobacco product group. Multiple imputation method will be used for missing data where appropriate. Sensitivity analysis using different analytic approaches, such as generalized linear models, as well as considering different covariate adjustments, will be used to build concordant results.

The dose-dependence of the changes in biomarkers will be determined by analyzing the data obtained from individuals that are exposed to different doses of a single product (e.g. smoking 0, <15 , 15-20 and >20 cigarettes per day) and by comparing between tobacco products that have different doses of HPHC constituents. In the US the average cigarettes per day is between 15-20³⁷ and therefore this dose range distribution is reflective of general population exposure. Comparisons of the effects of novel tobacco products and smoking will be informative of the relative toxicity of the two products.

We believe that the methods employed in the current project are exquisitely sensitive and responsive to even low dose insults such as ambient air pollution¹² allowing us to quantify tobacco product-induced changes with high precision. Moreover, levels of acrolein exposure vary between different individuals due to difference in puffing intensity and the time a cigarette is left smoldering. Thus, direct measurements of acrolein metabolites afford better estimates of acrolein exposure than machine yields. We expect to obtain wide variations in acrolein/crotonaldehyde exposure which will enable us to construct a dose-response relationship and identify which injury biomarkers are associated with aldehyde exposure and whether high levels of exposure are associated with high levels of injury, despite similar nicotine delivery.

Sample size

The sample size is justified in terms of the primary dependent measure, FMD, given the potential importance of this variable as a direct measure of the impact of tobacco exposure. The main comparisons are between non-tobacco users and tobacco users. Due to one control group, we will conservatively adjust our α (significance level) using a Bonferroni correction, and we will set $\alpha=0.01$. Based on preliminary data for FMD, we have observed mean \pm SD in smoker and nonsmoker groups to be 4.0 ± 1.6 and 6.8 ± 1.0 , respectively. We consider at least 25% (mean FMD=3.0 from 4.0) reduction from smokers to non-smokers is meaningful. Using a two sample, one-sided t test with an α of 0.01 and 80% power ($1-\beta$), assuming a common SD of 1.3, we will need 34 evaluable subjects in each group. We will recruit a total of 120 tobacco using participants per site. This over sampling will allow us to look at multiple endpoints and for associations with VOCs.

ETHICS AND DISSEMINATION

The CITU study was approved at each institution by their institutional review board (BU #H-32613 and UofL #13.0590) and all participants provide written consent. No study related procedures will be completed until after participant consent.

Participants for the CITU study are being recruited in both Boston, MA and Louisville KY. The two populations show significant differences, therefore recruitment at two sites will ensure a range more reflective of the general population. Although overall racial and ethnic demographics for both cities show a clear majority of Caucasians (70%) and despite smokers typically male, we strive to, and currently are successful in, recruiting a population that was gender balanced and almost evenly split between Caucasian and African Americans. Despite this balanced recruitment, e-cigarette users have been reported as predominantly Caucasian and male³⁸, and thus far our recruitment mirrors these demographics. We expect very few Hispanic/Latino's to participate, due to data suggesting tobacco use, including ENDS, tends to be lower among Hispanic's/Latino's^{38 39}. Thus we have also opted to only recruit English speakers. We have carefully develop our recruitment strategy and exclusion criteria to protect vulnerable populations, which is important since many report a lower socioeconomic status and educational level in smokers in addition to higher rates of reported alcohol and drug use^{40 41}.

Our study is an observational study where participants have already assumed the risk of using tobacco. Study procedures pose minimal risk. Given the known harms associated with smoking, we will provide information on tobacco treatment when

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3 298 requested by the participant. Participant information is de-identified for analysis and
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5 299 reported in aggregate to protect privacy.
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8 300 Completion of these studies will enable a greater understanding of the biological
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10 301 responses to use of a variety of tobacco products. Specifically, they will help to identify
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12 302 the constituents of these products; and how a panel of exposure and CV injury
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14 303 biomarkers are associated with these different constituents. This data will be available
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16 304 to the FDA and could help guide new policy measures to reduce or eliminate the
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18 305 harmful components of tobacco smoke and other nicotine products. The study is
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20 306 dedicated to the rapid dissemination of their rigorously characterized and well-controlled
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22 307 research findings to the public in the form of peer-reviewed publications. Subsequent to
23
24 308 the initial full-length manuscript publications of the resources generated with funding
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26 309 from this program, the study will make them available to interested and qualified
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28 310 investigators upon written request. The study will provide relevant protocols of published
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30 311 data, upon request (presuming prior publication by the Center members). Participants
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32 312 will be provided a summary of the results as they become available. Finally press
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34 313 releases of relevant findings will inform the general population.
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42 315 **LIST OF ABBREVIATIONS**

- 43
44 316 ABI- Ankle Brachial Index
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46 317 CAC= circulating angiogenic cells
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48 318 CRP= C-reactive protein
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50 319 CVD- Cardiovascular disease
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53 320 ENDS- Electronic nicotine Device (i.e. e-cigarette)
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- 321 FACS- Fluorescence-activated cell sorting
- 322 FMD- Flow mediated dilation
- 323 HDL= high density lipoprotein
- 324 IL-6= Interleukin 6
- 325 MMP- Matrix metalloproteinase
- 326 MP= micoparticles
- 327 PAI-- Plasminogen activator
- 328 PF4= Platelet factor 4
- 329 PWV- Pulse wave velocity
- 330 SAA= serum amyloid A
- 331 s-ICAM- soluble intercellular adhesion protein inhibitor
- 332 s-VCAM= soluble vascular adhesion protein
- 333 TNFR1= Tumor necrosis factor receptor 1
- 334 t-PA= tissue plasminogen activator
- 335 TxA2=Thromboxane A
- 336 VOC- Volatile organic compound
- 337 W:H- ratio: Waist to hip ratio
- 338

339 AUTHORS CONTRIBUTIONS

340 Rachel Keith- Study design, study recruitment, study visits, statistical analysis and
 341 manuscript preparation. Jessica Fetterman- study recruitment, study visits, manuscript
 342 preparation and editing. Dan Riggs- statistical analysis, manuscript preparation and
 343 editing. Tim O'Toole- Biomarker measurements, manuscript preparation and editing.

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3 344 Jessica Nystoriak- study recruitment and study visits. Monica Holbrook- study
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5 345 recruitment and study visits. Pawel Lorkiewicz- VOC measurements and manuscript
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7 346 preparation. Aruni Bhatnagar- Study design, study funding and manuscript editing.
8
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10 347 Andrew DeFilippis- Human subject assessment planning, manuscript preparation and
11
12 348 editing. Naomi M. Hamburg- Study design, study funding, vascular core, manuscript
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14 349 preparation and editing.

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17 350 **COMPETING INTERESTS**

18
19 351 None declared

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25
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27
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34
35 358 (CTP). The content is solely the responsibility of the authors and does not necessarily
36
37 359 represent the official views of the NIH or the Food and Drug Administration.

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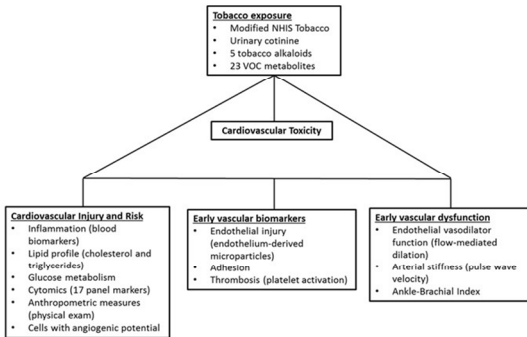
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Figure 1. Cardiovascular Injury due to Tobacco Use

CITU is designed to assess how tobacco related VOC exposure contributes to cardiovascular risk factors. Our exposure measurements include a panel of 23 urinary metabolites of 18 parent VOCs and tobacco use patterns. Cardiovascular phenotyping includes measures of injury, risk, vascular biomarkers and early vascular dysfunction. Tobacco use included use of traditional cigarettes, smokeless tobacco, waterpipe tobacco (hookah), electronic nicotine devices (ENDS), little cigars, cigarillos, pipes, cigars or any other form of tobacco that is available. Enrollment began in July 2014 and is ongoing.

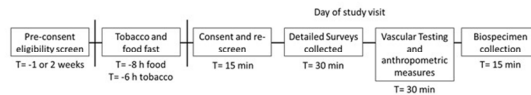
Figure 2. Study Visit Design

Study flow chart for interested participants from screening through study completion. Potential participants are pre-screened for eligibility prior to enrollment. Potential participants are asked to fast from tobacco for a minimum of 6 hours prior to the study visit. On the day of the visit the study lasts approximately 90 minute.



CITU is designed to assess how tobacco related VOC exposure contributes to cardiovascular risk factors. Our exposure measurements include a panel of 23 urinary metabolites of 18 parent VOCs and tobacco use patterns. Cardiovascular phenotyping includes measures of injury, risk, vascular biomarkers and early vascular dysfunction. Tobacco use included use of traditional cigarettes, smokeless tobacco, waterpipe tobacco (hookah), electronic nicotine devices (ENDS), little cigars, cigarillos, pipes, cigars or any other form of tobacco that is available. Enrollment began in July 2014 and is ongoing.

338x190mm (96 x 96 DPI)



Study flow chart for interested participants from screening through study completion. Potential participants are pre-screened for eligibility prior to enrollment. Potential participants are asked to fast from tobacco for a minimum of 6 hours prior to the study visit. On the day of the visit the study lasts approximately 90 minute.

338x190mm (96 x 96 DPI)

STROBE 2007 (v4) Statement—Checklist of items that should be included in reports of *cohort studies*

| Section/Topic | Item # | Recommendation | Reported on page # |
|---------------------------|--------|--|----------------------|
| Title and abstract | 1 | (a) Indicate the study’s design with a commonly used term in the title or the abstract | 1 |
| | | (b) Provide in the abstract an informative and balanced summary of what was done and what was found | 2-3 |
| Introduction | | | |
| Background/rationale | 2 | Explain the scientific background and rationale for the investigation being reported | 3-4 |
| Objectives | 3 | State specific objectives, including any prespecified hypotheses | 5 |
| Methods | | | |
| Study design | 4 | Present key elements of study design early in the paper | 5, 7 |
| Setting | 5 | Describe the setting, locations, and relevant dates, including periods of recruitment, exposure, follow-up, and data collection | 5, 7 |
| Participants | 6 | (a) Give the eligibility criteria, and the sources and methods of selection of participants. Describe methods of follow-up | 6-7 |
| | | (b) For matched studies, give matching criteria and number of exposed and unexposed | N/A |
| Variables | 7 | Clearly define all outcomes, exposures, predictors, potential confounders, and effect modifiers. Give diagnostic criteria, if applicable | 7-12 |
| Data sources/ measurement | 8* | For each variable of interest, give sources of data and details of methods of assessment (measurement). Describe comparability of assessment methods if there is more than one group | 7-12 |
| Bias | 9 | Describe any efforts to address potential sources of bias | 7 |
| Study size | 10 | Explain how the study size was arrived at | 14-16 |
| Quantitative variables | 11 | Explain how quantitative variables were handled in the analyses. If applicable, describe which groupings were chosen and why | 12-14 |
| Statistical methods | 12 | (a) Describe all statistical methods, including those used to control for confounding | 12-14 |
| | | (b) Describe any methods used to examine subgroups and interactions | 13 |
| | | (c) Explain how missing data were addressed | 13 |
| | | (d) If applicable, explain how loss to follow-up was addressed | N/A (study protocol) |
| | | (e) Describe any sensitivity analyses | 13 |
| Results | | | |

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|--------------------------|-----|--|----------------------|
| Participants | 13* | (a) Report numbers of individuals at each stage of study—eg numbers potentially eligible, examined for eligibility, confirmed eligible, included in the study, completing follow-up, and analysed | N/A (study protocol) |
| | | (b) Give reasons for non-participation at each stage | N/A (study protocol) |
| | | (c) Consider use of a flow diagram | N/A (study protocol) |
| Descriptive data | 14* | (a) Give characteristics of study participants (eg demographic, clinical, social) and information on exposures and potential confounders | N/A (study protocol) |
| | | (b) Indicate number of participants with missing data for each variable of interest | N/A (study protocol) |
| | | (c) Summarise follow-up time (eg, average and total amount) | N/A (study protocol) |
| Outcome data | 15* | Report numbers of outcome events or summary measures over time | N/A (study protocol) |
| Main results | 16 | (a) Give unadjusted estimates and, if applicable, confounder-adjusted estimates and their precision (eg, 95% confidence interval). Make clear which confounders were adjusted for and why they were included | N/A (study protocol) |
| | | (b) Report category boundaries when continuous variables were categorized | N/A (study protocol) |
| | | (c) If relevant, consider translating estimates of relative risk into absolute risk for a meaningful time period | N/A (study protocol) |
| Other analyses | 17 | Report other analyses done—eg analyses of subgroups and interactions, and sensitivity analyses | N/A (study protocol) |
| Discussion | | | |
| Key results | 18 | Summarise key results with reference to study objectives | N/A (study protocol) |
| Limitations | | | |
| Interpretation | 20 | Give a cautious overall interpretation of results considering objectives, limitations, multiplicity of analyses, results from similar studies, and other relevant evidence | 17 |
| Generalisability | 21 | Discuss the generalisability (external validity) of the study results | |
| Other information | | | |
| Funding | 22 | Give the source of funding and the role of the funders for the present study and, if applicable, for the original study on which the present article is based | 19 |

*Give information separately for cases and controls in case-control studies and, if applicable, for exposed and unexposed groups in cohort and cross-sectional studies.

Note: An Explanation and Elaboration article discusses each checklist item and gives methodological background and published examples of transparent reporting. The STROBE checklist is best used in conjunction with this article (freely available on the Web sites of PLoS Medicine at <http://www.plosmedicine.org/>, Annals of Internal Medicine at <http://www.annals.org/>, and Epidemiology at <http://www.epidem.com/>). Information on the STROBE Initiative is available at www.strobe-statement.org.

BMJ Open

Protocol to Assess the Impact of Tobacco-Induced Volatile Organic Compounds on Cardiovascular Risk in a Cross-Sectional Cohort: Cardiovascular Injury Due to Tobacco Study

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| Primary Subject Heading: | Cardiovascular medicine |
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| Keywords: | smoking, tobacco, electronic cigarette, cardiovascular risk, vascular injury, cigarettes |
| | |

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1 **Protocol to Assess the Impact of Tobacco-Induced Volatile Organic Compounds**
2 **on Cardiovascular Risk in a Cross-Sectional Cohort: Cardiovascular Injury Due to**
3 **Tobacco Study**

4 Rachel J. Keith, Jessica L. Fetterman, Daniel W. Riggs, Tim O’Toole, Jessica Nystoriak,
5 Monica Holbrook, Pawel Lorkiewicz, Aruni Bhatnagar, Andrew DeFilippis*, Naomi M.
6 Hamburg*

7 Rachel J. Keith-Division of Cardiovascular Medicine, University of Louisville School of
8 Medicine 580 S. Preston St. Louisville KY, 40202 rachel.keith@louisville.edu 502-852-
9 4211

10 Jessica L. Fetterman- Vascular Biology Section, Whitaker Cardiovascular Institute,
11 Boston University School of Medicine Evans Building, Boston, MA USA

12 Daniel W. Riggs- University of Louisville School of Medicine Louisville, KY USA

13 Timothy O-Toole- Division of Cardiovascular Medicine, University of Louisville School of
14 Medicine Louisville, KY USA

15 Jessica L. Nystoriak- Division of Cardiovascular Medicine, University of Louisville
16 School of Medicine Louisville, KY USA

17 Monika Holbrook- Vascular Biology Section, Whitaker Cardiovascular Institute, Boston
18 University School of Medicine Boston, MA USA

19 Pawel Lorkiewicz- Division of Cardiovascular Medicine, University of Louisville School
20 of Medicine Louisville, KY USA

21 Aruni Bhatnagar- Division of Cardiovascular Medicine, University of Louisville School of
22 Medicine Louisville, KY USA

Andrew P. DeFilippis- Division of Cardiovascular Medicine, University of Louisville
School of Medicine Louisville, KY USA (co-senior author)
Naomi M. Hamburg- Vascular Biology Section, Whitaker Cardiovascular Institute,
Boston University School of Medicine Boston, MA USA (co-senior author)

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ABSTRACT

Introduction: Tobacco use leads to increased mortality, the majority of which is attributed to cardiovascular disease. Despite this knowledge, the early cardiovascular impact of tobacco product use is not well understood. Tobacco use increases exposure to harmful and potentially harmful constituents including volatile organic compounds (VOCs) such as acrolein and crotonaldehyde, which may contribute to cardiovascular risk. The link between exposure patterns, risk profiles and demographic distribution of tobacco product users, particularly users of new and emerging products, are not well known. Therefore, we designed the Cardiovascular Injury due to Tobacco Use (CITU) study to assess population characteristics, demographic features, exposure patterns and cardiovascular risk in relation to tobacco.

Methods and analysis: We present the design and methodology of the CITU study a cross-section observational tobacco study conducted in Boston MA and Louisville KY starting in 2014. Healthy participants 21 to 45 years of age who use tobacco products, including ENDS, or who never used tobacco are being recruited. The study aims to recruit an evenly split cohort of African Americans and Caucasians that is sex balanced

for evaluation of self-reported tobacco exposure, VOC exposure and tobacco-induced injury profiling. Detailed information about participant’s demographics, health status and lifestyle is also collected.

Ethics and dissemination: The study protocol was approved institutional review boards at both participating universities. All study protocols will protect participant confidentiality. Results from the study will be disseminated via peer-reviewed journals and presented at scientific conferences.

Strengths and limitations

- Young age to allow for evaluation of early stage disease (e.g. inflammation, endothelial function) as opposed to end stage clinical consequence (e.g. myocardial infarction)
- Diverse tobacco product use allows for assessment of a wide range of tobacco-induced VOC exposure
- All study visits are in English introducing selection bias
- Data will inform regulatory agencies on the cardiovascular health effects of multiple tobacco products and the contribution of HPHCs

Keywords: Tobacco, smoking, electronic cigarette, vascular injury, cardiovascular risk, cigarettes.

INTRODUCTION

Tobacco product use and smoking are the leading causes of preventable deaths throughout the world. Of those deaths, one-third are attributed to cardiovascular disease (CVD)¹. The cardiovascular (CV) effects of tobacco exposure can include atherogenesis, vascular injury, thrombosis, arrhythmias and inflammation² and may be attributable to the many different harmful and potentially harmful constituents (HPHCs) present in tobacco products.

The HPHCs found in tobacco products include volatile organic compounds (VOCs) of which reactive aldehydes, such as acrolein and crotonaldehyde, are likely the most significant contributors to CV toxicity³. High levels of aldehydes are present in cigarette smoke^{4 5} as well as smokeless tobacco (ST)⁶. Risk assessments, using the prevalence of each individual chemical weighed by its potency, suggest that the non-cancer risk of smoking is dominated by acrolein, which contributes 40-100 times more to risk than any other chemical present in cigarette smoke³.

Although HPHCs, including VOC reactive aldehydes, have been suspected to be major contributors to the toxicity of cigarette smoke for over 4 decades, their contribution to CV injury and early CVD risk has not been rigorously evaluated. Experimental studies in animal models suggest that because of low aldehyde-metabolizing capacity, CV tissues are highly sensitive to aldehydes and exposure to low levels of aldehydes can induce CV injury and accelerate CVD⁷⁻¹⁸. The WHO Study Group on Tobacco Product Regulation (TobReg) has marked acrolein, a VOC, along with 8 other cigarette constituents for monitoring and regulation¹⁹ and the U.S. Environmental Protection Agency lists Acrolein as one of most hazardous air pollutants²⁰. Nevertheless, the contribution of tobacco induced VOCs, including acrolein

or other aldehydes, toward CV toxicity in humans has not been fully assessed. Greater understanding of how aldehydes affect cardiovascular health and disease will provide new avenues for evaluating the toxicity of cigarette smoke and for assessing the injurious potential of new and emerging tobacco products, such as ENDS, which may also contain VOCs including acrolein²¹⁻²³.

The latency period between tobacco exposure and the development of major clinical adverse health effects is long, therefore biomarkers that provide information over a shorter period allow for the identification of harm decades before clinical outcome data is available. Thus, in this paper we present the design and methodology of the Cardiovascular Injury due To Tobacco Use (CITU) study which will evaluate the association of the urinary metabolites of 18 parent VOCs from tobacco exposure with a comprehensive set of CV biomarkers representative of early disease and predictive of future CV events.²⁴

METHODS AND DESIGN

Overall design

The CITU study is an investigator-initiated cross-sectional observational study of around 500 healthy participants 21 to 45 years of age who are never or current tobacco product users in two urban areas at Boston University (BU) and University of Louisville (UofL) (Boston, MA and Louisville, KY) designed to evaluate CV toxicity due to tobacco product use, with correlations to VOCs found in the tobacco products (**Figure 1**).

Participant Eligibility Criteria

The goal of the study is to examine the impact of tobacco products on healthy young adults who could be classified as a current tobacco product users (Defined in table 1), or never-users (does not have lifetime use of any tobacco product). Participants were self-reported to be healthy therefore we excluded participants if they had: 1) diagnosis of clinical cardiovascular disease including but not limited to known heart attack, peripheral artery disease, heart failure or stroke; 2) diagnosis of diabetes (HbA1c >7.0 or treatment for diabetes), hypertension (systolic blood pressure >140 mm Hg or diastolic blood pressure >90 mm Hg), hypothyroidism or hyperthyroidism, inflammatory conditions such as lupus or inflammatory bowel disease, HIV/AIDS, hepatitis, liver disease, anemia, cancer of any type or another medical condition that might compromise the successful completion of the study; 2) recipients of organ transplant or renal replacement therapy; 3) individuals that are taking the following medications: immunosuppressant agents estrogen, testosterone, anti TNF agents, certain biologics, Procrit, statins, beta-blockers or other cardiovascular medicine; 4) individuals using nutraceuticals or anabolic steroids beyond the recommended daily allowance; 5) body weight less than 100 pounds; 6) pregnant women; 7) prisoners and other vulnerable populations; and 8) active illness or infection. Participants are rescheduled or considered screen-failures and excluded from the study if symptomatic of an acute illness, i.e. viral upper respiratory infection, on study date.

Table 1. Tobacco product use classifications

| Classification | Qualification |
|------------------------|---|
| Never | Does not meet lifetime limits for any tobacco use (see below) |
| Smoker | >100 lifetime cigarettes and current use for the past year |
| Smokeless Tobacco User | >20 lifetime dips or chews and current use for the past year |
| Cigar/Cigarillo User | >20 lifetime cigars or cigarillos and current use for the past year |
| Pipe User | >20 lifetime pipefuls and current use for the past year |

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| ENDS User | >20 lifetime vape sessions and current use for the past year |
| Hookah User | >20 lifetime hookah sessions and current use for the past year |

Study participants are screened prior to enrollment for current and past tobacco product use. Participants are characterized and assigned a use group based on self-reported patterns collected during the study visits.

Overall Study Procedure

Study participants fast for 8 h from food and 6 h from tobacco prior to the visit. All study visits occur before 11AM to limit effects due to circadian changes. All vascular function studies are completed after 10 min of supine positioning. All vascular studies are sent to the BU central lab for analysis. BU biologic samples have minimal processing and are shipped overnight to the UofL central laboratory at the completion of each study visit. Samples obtained at UofL are processed to a similar stage, then held overnight prior to analysis for standardization of time to measurement for all samples.

Study visits take approximately 90 minutes to complete and include a structured interview on demographics, socioeconomics, lifestyle, health, family history of heart disease, allergies, and tobacco use. (Figure 2) Participants were compensated appropriately for their time. All surveys are collected and kept in Research Electronic Data Capture (REDCap), a secure web application for building and managing online surveys and databases.

Exposure Variables

Tobacco Product Use & Particulate Matter Exposure

Comprehensive tobacco product exposure is assessed using a modified version of the National Health Interview survey on tobacco use²⁵. The survey is modified to include detailed information on electronic nicotine devices (ENDs) and other new or

156 emerging tobacco products. Residential addresses are collected for assessment of
157 ambient airborne particulate matter (PM_{2.5}) exposure and future correction of overall
158 exposure. PM_{2.5} data from the day of the study visit, and 3 and 5 days prior to the study
159 is collected from publicly available data associated with EPA monitoring stations. Other
160 exposure variables, including occupation, are collected through interview.

161 VOC Measurements

162 Humans are exposed to VOCs from a variety of sources including indoor and
163 outdoor environments as well as diet. The most significant sources of ambient exposure
164 ambient are air pollution, car exhaust, household products, personal hygiene products,
165 and solvents^{26 27}. Although concurrent exposures from multiple sources could confound
166 attribution to smoking, the levels of urinary metabolites of these VOCs in smokers far
167 exceeds those measured in non-smokers exposed to typical sources of VOCs²⁸.

168 Standard clean catch urine specimens are obtained from participants. Though
169 only a single urine time point is collected, previous studies show spot urine
170 measurements correlate well with 24-hour urine collections²⁹. Many VOC metabolites
171 have relatively short half-lives that range from 2 - 25.2h,^{30 31} but given the constant
172 pattern of tobacco product use by most users, spot collection reflects recurrent use.
173 Moreover, even though some VOC metabolites, such as HPMA, are known vary with
174 time of day,²⁹ synchronizing the study visits and requiring a tobacco fast is likely to
175 minimize diurnal variations in metabolism. Our past work has shown that spot-urine
176 collected at the same time of day reliably reflects daily VOC exposure and is correlated
177 to CVD risk³².

We have developed a robust Core Lab that utilizes mass spectrometry procedures adopted from the Centers for Disease Control and Prevention (CDC) protocols, to quantify 23 urinary metabolites of tobacco smoking related toxins (aldehydes and other VOCs), including acrolein³³ (**Table 2**). The concentration values of analytes are then normalized to urinary creatinine levels measured using Infinity Creatinine Reagent (Thermo Fisher Scientific, MA) on a COBAS MIRA-plus analyzer (Roche, NJ).

Table 2 Exposure Variables (Please see end of article)

| <i>Parent compound</i> | <i>VOC metabolite</i> | <i>Common abbr.</i> |
|---|--|---------------------|
| Acetaldehyde | Acetic acid/Acetate | ACETATE |
| Acrolein | N-Acetyl-S-(2-carboxyethyl)-L-cysteine | CEMA |
| | N-Acetyl-S-(3-hydroxypropyl)-L-cysteine | 3HPMA |
| Acrylamide | N-Acetyl-S-(2-carbamoyl)-L-cysteine | AAMA |
| | N-Acetyl-S-(2-carbamoyl-2-hydroxyethyl)-L-cysteine | GAMA |
| Acrylonitrile | N-Acetyl-S-(2-cyanoethyl)-L-cysteine | CYMA |
| Acrylonitrile, vinyl chloride, ethylene oxide | N-Acetyl-S-(2-hydroxyethyl)-L-cysteine | HEMA |
| Anabasine | Anabasine (free) | ANB |
| Anatabine | Anatabine (free) | ANTB |
| Benzene | N-Acetyl-S-(phenyl)-L-cysteine | PMA |
| | trans, trans-Muconic acid | MU |

| | | |
|-----------------------|--|----------|
| 1-Bromopropane | N-Acetyl-S-(n-propyl)-L-cysteine | BPMA |
| 1,3-Butadiene | N-Acetyl-S-(3,4-dihydroxybutyl)-L-cysteine | DHBMA |
| | N-Acetyl-S-(1-hydroxymethyl-2-propenyl)-L-cysteine | MHBMA1 |
| | N-Acetyl-S-(2-hydroxy-3-butenyl)-L-cysteine | MHBMA2 |
| | N-Acetyl-S-(4-hydroxy-2-buten-1-yl)-L-cysteine | MHBMA3 |
| Carbon-disulfide | 2-Thioxothiazolidine-4-carboxylic acid | TTCA |
| Crotonaldehyde | N-Acetyl-S-(3-hydroxypropyl-1-methyl)-L-cysteine | HPMMA |
| Cyanide | 2-Aminothiazoline-4-carboxylic acid | ATCA |
| N,N-Dimethylformamide | N-Acetyl-S-(N-methylcarbamoyl)-L-cysteine | AMCC |
| Ethylbenzene, styrene | Phenylglyoxylic acid | PGA |
| Formaldehyde | Formate | FORMATE |
| Nicotine | Nicotine | NIC |
| | Cotinine | COT |
| | 3-Hydroxycotinine | 3HC |
| Propylene oxide | N-Acetyl-S-(2-hydroxypropyl)-L-cysteine | 2HPMA |
| Styrene | N-Acetyl-S-(1-phenyl-2-hydroxyethyl)-L-cysteine + | PHEMA |
| | N-Acetyl-S-(2-phenyl-2-hydroxyethyl)-L-cysteine | |
| | Mandelic acid | MA |
| Tetrachloroethylene | N-Acetyl-S-(trichlorovinyl)-L-cysteine | TCVMA |
| Toluene | N-Acetyl-S-(benzyl)-L-cysteine | BMA |
| Trichloroethylene | N-Acetyl-S-(1,2-dichlorovinyl)-L-cysteine | 1,2DCVMA |
| | N-Acetyl-S-(2,2-dichlorovinyl)-L-cysteine | 2,2DCVMA |

| | | |
|--------|--|------------|
| Xylene | N-Acetyl-S-(2,4-dimethylphenyl)-L-cysteine + N-Acetyl-S-(2,5-dimethylphenyl)-L-cysteine + N-Acetyl-S-(3,4-dimethylphenyl)-L-cysteine | DPMA |
| | 2-Methylhippuric acid | 2MHA |
| | 3-Methylhippuric acid + 4-Methylhippuric acid | 3MHA+ 4MHA |

Urine is analyzed for 23 metabolites of 18 parent VOCs and tobacco alkaloids by UPLC-MS/MS. Analytes are listed as parent, metabolite and their common abbreviation.

Circulating Markers of Cardiovascular Injury

To assess tobacco product-induced cardiovascular toxicity, we examine endothelial function, inflammatory mediators, biomarkers, and thrombosis. CV risk is defined through measurements of circulating angiogenic cells, lipid profile, and glucose metabolism^{24 34 35}. Plasma (BD367863 and BD366415) and serum (BD367814) samples are obtained from all participants for laboratory testing and long term biobanking. Whole blood (BD366415) is obtained for flow cytometry on fresh samples at UofL pathology core. BU biologic samples have minimal processing and are shipped overnight to the UofL central laboratory at the completion of each study visit. Samples obtained at UofL are processed to a similar stage, then held overnight prior to analysis to standardize the time to measurement for all samples. The UofL central laboratory, as previously reported, will complete fasting and biomarker measurements (Table 3), with the exception of cytomics^{12 36}. For cytomic measurements, mononuclear cells are labeled with the peripheral blood phenotyping panel kit (Fluidigm).Samples are shipped

at 4 degree C to Core Lab facilities at the University of Rochester for Mass cytometric analysis.

Table 3 Blood analysis

Fasting Measurements

LDL cholesterol, HDL cholesterol, total cholesterol, triglycerides, glucose, uric acid, SAA and fibrinogen

Biomarkers

CAC (1-15)¹, Platelet-monocyte aggregates, MP (1-5)¹, PF4, t-PA, TxA2, Factor VII, IL-6, CRP, D-dimer, PAI-1, s-ICAM-1, s-VCAM, s-thrombomodulin, s-TNFR1, MMP-2, MMP-3, MMP-9, cytomix, endothelin, E-selectin and P-selectin

1: Fifteen different CAP subpopulations and 5 subtypes of microparticles were measured by flow cytometry.

All participants who complete the study visit will have blood samples taken and processed. Flow cytometric analysis is completed on fresh samples. All other analysis will be completed on biobanked samples in batches LDL= low density lipoprotein. HDL= high density lipoprotein. SAA= serum amyloid A. CAC= circulating angiogenic cells. MP= microparticles. PF4= Platelet factor 4. t-PA= tissue plasminogen activator. TxA2=Thromboxane A. IL-6= Interleukin 6. CRP= C-reactive protein. PAI= Plasminogen activator. s- ICAM- soluble intercellular adhesion protein inhibitor. s-VCAM= soluble vascular adhesion protein. TNFR1= Tumor necrosis factor receptor 1. MMP- Matrix metalloproteinase.

Non-Invasive Vascular Function Testing

Smoking, is associated with endothelial damage and vascular dysfunction^{37 38}. Endothelial cells are exposed to circulating toxins and measures of endothelial function are reflective of cardiovascular injury³⁹. Thus, we examine the non-invasive endothelial vasodilator function using flow-mediated vasodilation^{40 41}, arterial stiffness with carotid-femoral and carotid-radial pulse wave velocity⁴², and peripheral vascular function with

ankle brachial index. Flow mediated dilation was assessed with a 7.5MHZ ultrasound probe is used to image the brachial artery while a 10cm blood pressure cuff is attached to the lower arm and a 3 lead ECG is attached to the patient. After baseline images and 10 cycles of Doppler images are captured using NIHEM R-wave triggered image capturing software, the blood pressure cuff is inflated to 200mmHg or 50mmHg higher than the systolic pressure. After the 5 minute occlusion, the cuff is released and the NIHEM software records two minutes of imaging. Images were analyzed by a single blinded analyzer using MIA vascular Research Tolls Brachial Analyzer for Research, version 6.8.5. All vascular imagers where trained at BU who have a previously reported reproducibility with intra- and inter-observer correlation coefficients of 0.98 and 0.99 for brachial diameter and 0.78 and 0.92 for FMD.⁴³ Similar equipment and software is used at both sites. All vascular studies are sent to the BU central lab for analysis.

Anthropometric measures

Anthropometric measures included height, weight, waist and hip circumference and body fat. All anthropometric measures are completed twice and the average recorded. Standing height measurements are completed on a fixed stadiometer. Weight measurements are completed on a digital scale to the nearest tenth of a pound. Waist circumference is measured at the level of the umbilicus to the nearest tenth of a centimeter. Hip circumference is measured at the maximal protrusion of the gluteal muscle to the nearest tenth of a centimeter. Body fat percentage is calculated by the bioelectrical impedance measured with the Omron fat loss monitor (HBF-306C).

DATA ANALYSIS

We expect that from this study we will be able to identify specific biomarkers of cardiovascular injury due to tobacco use and the relationship of these biomarkers to specific measures of tobacco exposure. For instance, we will identify which biomarkers are affected by tobacco use, and which ones are most sensitive; including their dose-dependence. Additionally we will examine the extent to which biomarkers are associated with exposure to nicotine versus exposure to HPHC of tobacco like aldehydes.

Sample size

The sample size is justified in terms of the primary dependent measure, FMD, given the potential importance of this variable as a direct measure of the impact of tobacco exposure. The main comparisons are between non-tobacco users and tobacco users. Due to one control group, we will conservatively adjust our α (significance level) using a Bonferroni correction, and we will set $\alpha=0.01$. Based on preliminary data for FMD, we have observed mean \pm SD in smoker and nonsmoker groups to be 4.0 ± 1.6 and 6.8 ± 1.0 , respectively. We consider at least 25% (mean FMD=3.0 from 4.0) reduction from smokers to non-smokers is meaningful. Using a two sample, one-sided t test with an α of 0.01 and 80% power ($1-\beta$), assuming a common SD of 1.3, we will need 34 evaluable subjects in each group. We will recruit a total of 120 tobacco using participants per site. This over sampling will allow us to look at multiple endpoints and for associations with VOCs.

Analysis Plan

All statistical analysis will be performed using SAS version 9.4 software (SAS Institute, Inc., Cary, North Carolina), and a two-sided p-value of <0.05 will be considered

significant for any statistical test. Demographics and other baseline characteristics will be summarized according to product group. Differences in VOC's between product groups will be tested using ANOVA for normally distributed data or Kruskal-Wallis test for non-normal data. The association between primary outcomes of vascular function as well as circulating markers of cardiovascular injury with individual VOC levels will be analyzed using multiple regression models, adjusting for appropriate confounders. Additionally, because we have multiple VOC's, which are highly correlated, we will use methods such as LASSO to identify the VOC's that are most associated with the outcomes of interest. Multipollutant approaches, such as principal component analysis (PCA), will be used to test whether overall VOC exposure is associated with the health outcomes. Interaction variables will be tested for in the regression models and subgroup analyses will be conducted according to the following factors: significant interactions, sex, age, race, tobacco product group. Multiple imputation method will be used for missing data where appropriate. Sensitivity analysis using different analytic approaches, such as generalized linear models, as well as considering different covariate adjustments, will be used to build concordant results.

The dose-dependence of the changes in biomarkers will be determined by analyzing the data obtained from individuals that are exposed to different doses of a single product (e.g. smoking 0, <10, 10-20 and >20 cigarettes per day) and by comparing between tobacco products that have different doses of HPHC constituents. In the US the average cigarettes per day is between 10-20⁴⁴ and therefore this dose range distribution is reflective of general population exposure. Comparisons of the

effects of novel tobacco products and smoking will be informative of the relative toxicity of the two products.

We believe that the methods employed in the current project are exquisitely sensitive and responsive to even low dose insults such as ambient air pollution¹² allowing us to quantify tobacco product-induced changes with high precision. Moreover, levels of acrolein exposure vary between different individuals due to difference in puffing intensity and the time a cigarette is left smoldering. Thus, direct measurements of acrolein metabolites afford better estimates of acrolein exposure than machine yields. We expect to obtain wide variations in acrolein/crotonaldehyde exposure which will enable us to construct a dose-response relationship and identify which injury biomarkers are associated with aldehyde exposure and whether high levels of exposure are associated with high levels of injury, despite similar nicotine delivery.

ETHICS AND DISSEMINATION

The CITU study was approved at each institution by their institutional review board (BU #H-32613 and UofL #13.0590) and all participants provide written consent. No study related procedures will be completed until after participant consent.

Participants for the CITU study are being recruited in both Boston, MA and Louisville KY. The two populations show significant differences, therefore recruitment at two sites will ensure a range more reflective of the general population. Although overall racial and ethnic demographics for both cities show a clear majority of Caucasians (70%) and despite smokers typically male, we strive to, and currently are successful in, recruiting a population that was gender balanced and almost evenly split between Caucasian and African Americans. Despite this balanced recruitment, e-cigarette users

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312 have been reported as predominantly Caucasian and male⁴⁵, and thus far our
313 recruitment mirrors these demographics. We expect very few Hispanic/Latino's to
314 participate, due to data suggesting tobacco use, including ENDS, tends to be lower
315 among Hispanic's/Latino's ^{45 46}. Thus we have also opted to only recruit English
316 speakers. We have carefully develop our recruitment strategy and exclusion criteria to
317 protect vulnerable populations, which is important since many report a lower
318 socioeconomic status and educational level in smokers in addition to higher rates of
319 reported alcohol and drug use ^{47 48}.

320 Our study is an observational study where participants have already assumed
321 the risk of using tobacco. Study procedures pose minimal risk. Given the known harms
322 associated with smoking, we will provide information on tobacco treatment when
323 requested by the participant. Participant information is de-identified for analysis and
324 reported in aggregate to protect privacy.

325 Completion of these studies will enable a greater understanding of the biological
326 responses to use of a variety of tobacco products. Specifically, they will help to identify
327 the constituents of these products; and how a panel of exposure and CV injury
328 biomarkers are associated with these different constituents. This data will be available
329 to the FDA and could help guide new policy measures to reduce or eliminate the
330 harmful components of tobacco smoke and other nicotine products. The study is
331 dedicated to the rapid dissemination of their rigorously characterized and well-controlled
332 research findings to the public in the form of peer-reviewed publications. Subsequent to
333 the initial full-length manuscript publications of the resources generated with funding
334 from this program, the study will make them available to interested and qualified

investigators upon written request. The study will provide relevant protocols of published data, upon request (presuming prior publication by the Center members). Participants will be provided a summary of the results as they become available. Finally press releases of relevant findings will inform the general population.

LIST OF ABBREVIATIONS

ABI- Ankle Brachial Index
CAC= circulating angiogenic cells
CRP= C-reactive protein
CVD- Cardiovascular disease
ENDS- Electronic nicotine Device (i.e. e-cigarette)
FACS- Fluorescence-activated cell sorting
FMD- Flow mediated dilation
HDL= high density lipoprotein
IL-6= Interleukin 6
MMP- Matrix metalloproteinase
MP= micoparticles
PAI=- Plasminogen activator
PF4= Platelet factor 4
PWV- Pulse wave velocity
SAA= serum amyloid A
s-ICAM- soluble intercellular adhesion protein inhibitor
s-VCAM= soluble vascular adhesion protein

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3 358 TNFR1= Tumor necrosis factor receptor 1
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5 359 t-PA= tissue plasminogen activator
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8 360 TxA2=Thromboxane A
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10 361 VOC- Volatile organic compound
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12 362 W:H- ratio: Waist to hip ratio
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17 364 **AUTHORS CONTRIBUTIONS**

18
19 365 Rachel Keith- Study design, study recruitment, study visits, statistical analysis and
20
21 366 manuscript preparation. Jessica Fetterman- study recruitment, study visits, manuscript
22
23 367 preparation and editing. Dan Riggs- statistical analysis, manuscript preparation and
24
25 368 editing. Tim O'Toole- Biomarker measurements, manuscript preparation and editing.
26
27 369 Jessica Nystoriak- study recruitment and study visits. Monica Holbrook- study
28
29 370 recruitment and study visits. Pawel Lorkiewicz- VOC measurements and manuscript
30
31 371 preparation. Aruni Bhatnagar- Study design, study funding and manuscript editing.
32
33 372 Andrew DeFilippis- Human subject assessment planning, manuscript preparation and
34
35 373 editing. Naomi M. Hamburg- Study design, study funding, vascular core, manuscript
36
37 374 preparation and editing.

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39 375 **COMPETING INTERESTS**

40
41 376 None declared

42
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44
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46
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48
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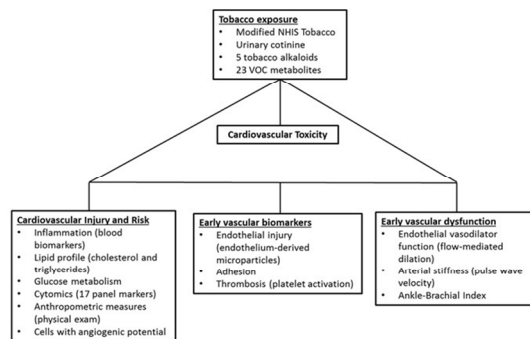
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Figure 1. Cardiovascular Injury due to Tobacco Use

CITU is designed to assess how tobacco related VOC exposure contributes to cardiovascular risk factors. Our exposure measurements include a panel of 23 urinary metabolites of 18 parent VOCs and tobacco use patterns. Cardiovascular phenotyping includes measures of injury, risk, vascular biomarkers and early vascular dysfunction. Tobacco use included use of traditional cigarettes, smokeless tobacco, waterpipe tobacco (hookah), electronic nicotine devices (ENDS), little cigars, cigarillos, pipes, cigars or any other form of tobacco that is available. Enrollment began in July 2014 and is ongoing.

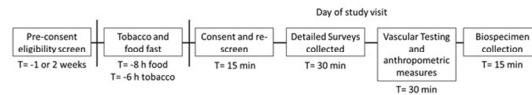
Figure 2. Study Visit Design

Study flow chart for interested participants from screening through study completion. Potential participants are pre-screened for eligibility prior to enrollment. Potential participants are asked to fast from tobacco for a minimum of 6 hours prior to the study visit. On the day of the visit the study lasts approximately 90 minute.



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108x60mm (300 x 300 DPI)



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108x60mm (300 x 300 DPI)

STROBE 2007 (v4) Statement—Checklist of items that should be included in reports of *cohort studies*

| Section/Topic | Item # | Recommendation | Reported on page # |
|---------------------------|--------|--|----------------------|
| Title and abstract | 1 | (a) Indicate the study’s design with a commonly used term in the title or the abstract | 1 |
| | | (b) Provide in the abstract an informative and balanced summary of what was done and what was found | 2-3 |
| Introduction | | | |
| Background/rationale | 2 | Explain the scientific background and rationale for the investigation being reported | 3-4 |
| Objectives | 3 | State specific objectives, including any prespecified hypotheses | 5 |
| Methods | | | |
| Study design | 4 | Present key elements of study design early in the paper | 5, 7 |
| Setting | 5 | Describe the setting, locations, and relevant dates, including periods of recruitment, exposure, follow-up, and data collection | 5, 7 |
| Participants | 6 | (a) Give the eligibility criteria, and the sources and methods of selection of participants. Describe methods of follow-up | 6-7 |
| | | (b) For matched studies, give matching criteria and number of exposed and unexposed | N/A |
| Variables | 7 | Clearly define all outcomes, exposures, predictors, potential confounders, and effect modifiers. Give diagnostic criteria, if applicable | 7-12 |
| Data sources/ measurement | 8* | For each variable of interest, give sources of data and details of methods of assessment (measurement). Describe comparability of assessment methods if there is more than one group | 7-12 |
| Bias | 9 | Describe any efforts to address potential sources of bias | 7 |
| Study size | 10 | Explain how the study size was arrived at | 14-16 |
| Quantitative variables | 11 | Explain how quantitative variables were handled in the analyses. If applicable, describe which groupings were chosen and why | 12-14 |
| Statistical methods | 12 | (a) Describe all statistical methods, including those used to control for confounding | 12-14 |
| | | (b) Describe any methods used to examine subgroups and interactions | 13 |
| | | (c) Explain how missing data were addressed | 13 |
| | | (d) If applicable, explain how loss to follow-up was addressed | N/A (study protocol) |
| | | (e) Describe any sensitivity analyses | 13 |
| Results | | | |

| | | | |
|-------------------|-----|--|----------------------|
| Participants | 13* | (a) Report numbers of individuals at each stage of study—eg numbers potentially eligible, examined for eligibility, confirmed eligible, included in the study, completing follow-up, and analysed | N/A (study protocol) |
| | | (b) Give reasons for non-participation at each stage | N/A (study protocol) |
| | | (c) Consider use of a flow diagram | N/A (study protocol) |
| Descriptive data | 14* | (a) Give characteristics of study participants (eg demographic, clinical, social) and information on exposures and potential confounders | N/A (study protocol) |
| | | (b) Indicate number of participants with missing data for each variable of interest | N/A (study protocol) |
| | | (c) Summarise follow-up time (eg, average and total amount) | N/A (study protocol) |
| Outcome data | 15* | Report numbers of outcome events or summary measures over time | N/A (study protocol) |
| Main results | 16 | (a) Give unadjusted estimates and, if applicable, confounder-adjusted estimates and their precision (eg, 95% confidence interval). Make clear which confounders were adjusted for and why they were included | N/A (study protocol) |
| | | (b) Report category boundaries when continuous variables were categorized | N/A (study protocol) |
| | | (c) If relevant, consider translating estimates of relative risk into absolute risk for a meaningful time period | N/A (study protocol) |
| Other analyses | 17 | Report other analyses done—eg analyses of subgroups and interactions, and sensitivity analyses | N/A (study protocol) |
| Discussion | | | |
| Key results | 18 | Summarise key results with reference to study objectives | N/A (study protocol) |
| Limitations | | | |
| Interpretation | 20 | Give a cautious overall interpretation of results considering objectives, limitations, multiplicity of analyses, results from similar studies, and other relevant evidence | 17 |
| Generalisability | 21 | Discuss the generalisability (external validity) of the study results | |
| Other information | | | |
| Funding | 22 | Give the source of funding and the role of the funders for the present study and, if applicable, for the original study on which the present article is based | 19 |

*Give information separately for cases and controls in case-control studies and, if applicable, for exposed and unexposed groups in cohort and cross-sectional studies.

Note: An Explanation and Elaboration article discusses each checklist item and gives methodological background and published examples of transparent reporting. The STROBE checklist is best used in conjunction with this article (freely available on the Web sites of PLoS Medicine at <http://www.plosmedicine.org/>, Annals of Internal Medicine at <http://www.annals.org/>, and Epidemiology at <http://www.epidem.com/>). Information on the STROBE Initiative is available at www.strobe-statement.org.