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Long-Term Human Papillomavirus Vaccination Effectiveness and Immunity in Rwandan Women Living with and without Human Immunodeficiency Virus

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Long-Term Human Papillomavirus Vaccination Effectiveness and Immunity in Rwandan Women Living with and without Human Immunodeficiency Virus

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Abstract

Introduction. Prophylactic human papillomavirus (HPV) vaccines have been shown to be highly effective in protecting women against cervical infections, high-grade abnormalities, and cancer caused by the targeted HPV types. However, the evidence for their effectiveness in women living with human immunodeficiency virus (HIV) (WLWH) is less clear.

Methods. WLWH and HIV-negative women who likely did (birth cohorts 1996 and later) and WLWH who likely did not (birth cohorts before 1996) receive HPV vaccination (n=757 for each of the three groups) through the Rwanda national vaccination program will be recruited into the study. Between groups, we will compare cervicovaginal, anal and oral prevalent and 6-month persistent HPV6/11/16/18 infections as measured using a modified AmpFire HPV genotyping assay that tests for 15 high-risk or intermediate-risk HPV genotypes, HPV6, and HPV11. We will also compare the HPV immune response in 548 HPV-vaccinated WLWH to 548 HPV-vaccinated HIV-negative women using an anti-HPV16 and -HPV18 ELISA. Vaccination status will be confirmed through national vaccination records.

Analysis. We will calculate point prevalence and prevalence of 6-month persisting infections by individual HPV-type specific infections and groups of infections for each anatomic site and for each group of women. Results will be stratified by age at vaccination, age at enrollment, and number of doses (3 vs. 2) as well as other factors possibly associated with HPV prevalence. Differences in endpoints between groups, overall and between sub-groups, will be tested for statistical significance ($p < 0.05$) using Fisher's exact or Pearson chi-square test. Differences in geometric mean titers (GMT) and seropositivity will be tested for statistical significance using the Mann-Whitney and Fisher's exact tests, respectively.

Ethics and Dissemination. The study was approved by the Albert Einstein College of Medicine Institutional Review Board and the Rwanda National Ethics Committee. Results will be disseminated through publication in peer-reviewed journals.

Key Words: Human papillomavirus (HPV), vaccination, cervical, anal, oral, cancer, Rwanda

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STRENGTHS AND LIMITATIONS OF THIS STUDY

Please note that ‘Strengths and limitations of this study’ should consist of 3-5 bullet points.

- The study is being conducted in Rwanda, a high cervical-cancer burden country
- National introduction of HPV vaccine in Rwanda in 2011 and relatively high prevalence of HIV makes a study of long-term HPV vaccination effective in women living with HIV (WLWH) possible.
- The study is not a randomized control trial, but an observational study of populations selected from different groups of women.
- A convenience sample of Rwandan WLWH living in and around Kigali will be enrolled and therefore the study population is not representative of all Rwanda or WLWH living elsewhere.

INTRODUCTION

Current prophylactic vaccines against human papillomavirus (HPV), the necessary cause of virtually all cervical cancer^{1 2}, are based on the self-assembly of recombinantly expressed L1 protein in cell lines into virus-like particles (VLPs) that resemble native viral capsids but without the viral genome necessary for viral replication. The first generation of prophylactic HPV vaccines, Gardasil® (Merck & Co, Kenilworth, NJ, USA)³ and Cervarix™ (GlaxoSmithKline, Wavre, Belgium)³, targeted HPV16 and HPV18 (HPV16/18), which cause approximately 70% of cervical cancers.⁴ Gardasil® also targets HPV6 and HPV11 (HPV6/11), non-high-risk HPV types responsible for approximately 90% of anogenital warts (*Condylomata acuminata*).⁵

Several countries, including Australia⁶⁻¹², Scotland¹³, Denmark¹⁴, and the U.S.¹⁵⁻¹⁷, were early adopters of HPV vaccination and have documented reductions in infections, diseases, and cervical abnormalities related to the HPV vaccine-targeted types. A meta-analysis on the impact of HPV vaccination found reductions in anogenital warts, HPV infections, cervical intraepithelial neoplasia (CIN) grade 2 (CIN2) or more severe diagnoses (CIN2+) among girls and women, and on anogenital warts diagnoses among girls, women, boys, and men.¹⁸ Recent reports from Finland¹⁹, Sweden²⁰, and England²¹ now provided real-world evidence that HPV vaccination prevents cervical cancer.

Cervical cancer was included as an acquired immunodeficiency syndrome (AIDS)-defining disease in adolescents and adults in 1993.^{22 23} Women living with human immunodeficiency (WLWH) virus, the cause of AIDS, have a significantly elevated risk of cervical cancer^{24 25}, due to an impaired immune response to HPV, compared to HIV-uninfected (HIV[-]) women. Meta-analyses of HIV/AIDS cohorts reported a 6-fold increased incidence of cervical cancer compared

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88 to the general female population/HIV[-] women.^{24 26} People living with HIV (PLWH) are also at
89 an increased risk of other HPV-related malignancies, notably anal and oropharyngeal cancers.²⁴
90 ²⁷ HIV co-infection has a profound impact on the natural history of HPV, thereby increasing the
91 risk of ICC. HIV co-infection increases the 1) likelihood of cervical and anal HPV persistence
92 and 2) likelihood of cervical and anal HPV infections progressing to precancer.^{28 29}

93 Data on HPV-vaccine efficacy and effectiveness are lacking for WLWH.^{24 25} To date, most
94 studies of HPV vaccine in WLWH focused on immunogenicity and safety. Generally, HPV
95 vaccination in PLWH has been well tolerated, safe, and resulted in good immune responses³⁰⁻³⁴;
96 one study found lower seroconversion and anti-HPV antibody titers among PLWH compared to
97 perinatally HIV-exposed, uninfected youth.³⁴ Studies have noted an impact of HIV disease status
98 (CD4 counts and viral suppression) on the immune responses to HPV vaccination.³⁵⁻³⁷ Studies
99 have reported lower seroconversion and antibody titers in WLWH not taking (vs. taking)
100 antiretroviral therapy (ART) and with lower (vs. higher) CD4 counts.^{38 39} Similarly, another
101 study found peak antibody titers to be 2- to 3-fold higher in mid-adult WLWH with full HIV
102 viral suppression compared to those not suppressed.⁴⁰ Interestingly, higher anti-HPV18 titers but
103 similar anti-HPV16 titers were reported in response to HPV vaccination by Cervarix® compared
104 to Gardasil® in PLWH but the differences in the former was primarily due to differences in the
105 immune responses in the WLWH.⁴¹

106 The few studies of HPV vaccine effectiveness in WLWH provided promising but inconclusive
107 evidence. Immunogenicity studies in WLWH have been of insufficient sample size to address
108 efficacy/effectiveness. Studies of HPV vaccination in select PLWH at high risk of anal cancer
109 have been limited in sample size because of the high HPV anal exposure (and likely
110 misclassification of exposure) to targeted HPV types prior to enrollment⁴² so that few truly

incident events could be observed. Of those studies that did have endpoints, one recent trial of Gardasil® in PLWH aged 27 and older concluded that “This double-blind, randomized trial did not find a benefit from HPV vaccination to prevent anal HPV infection or anal high-grade squamous intraepithelial lesions” and favorable but inconclusive benefit for protection against oral HPV.⁴³

The paucity of HPV-vaccine effectiveness data in WLWH is especially a concern for the prevention of cervical cancer in sub-Saharan Africa (SSA), where cervical cancer is the most common cause of cancer death in women.⁴⁴ Almost one quarter of the global burden of cervical cancer occurs in SSA and an estimated 20% of cervical cancer in SSA is attributable to HIV coinfection.²⁶ Evidence of the protective effects of HPV vaccination in SSA WLWH is needed, especially since most long-term public health planning to address cervical cancer in SSA depends on the unproven effectiveness of these vaccines.

Rwanda, a central/east African country that experiences a high burden of cervical cancer⁴⁵, is the ideal locale to answer questions about the long-term effectiveness and immunogenicity of Gardasil® in WLWH for the following reasons. First, in 2011-13, Rwanda, through a donation from Merck, launched a national HPV vaccination program.⁴⁶ In 2011, over 92,000 girls in primary school grade six (~12 years old) were vaccinated with three doses of Gardasil®. During 2012 and 2013, a catch-up vaccination program targeted girls in secondary school grade three (~15 years old).⁴⁶ In 2014, HPV vaccination was supported by GAVI⁴⁷ and reverted to vaccinating 12-year-old girls.⁴⁸ In 2015, Rwanda switched from 3 doses to 2 doses, 6 months apart, for vaccinating 12-year-old girls. In all years, Rwanda achieved $\geq 90\%$ annual coverage with the recommended number of HPV vaccine doses in the target population.^{46 47 49 50} Thus, Rwanda is one of the earliest and most successful adopters of HPV vaccination globally.

134 Second, the prevalence of HIV in Rwanda is 2.7% among 15-30 year olds.⁵¹ The excellent HIV
135 care program in Rwanda allows for easy and efficient recruitment and following of WLWH in an
136 observational study of HPV-vaccination impact. Third, given Rwanda's nearly complete national
137 coverage with Gardasil®, HPV-vaccinated and unvaccinated WLWH are easily recruited based
138 on birth year. Rwanda has excellent national vaccination records so that retrospectively the small
139 number of participants misclassified by HPV vaccination status based on birth year subsequently
140 can be correctly categorized.

141 The Einstein/Rwanda/DRC Consortium for Research in HIV/HPV/Malignancies therefore
142 launched an observational cohort of HPV-vaccinated and -unvaccinated WLWH and vaccinated
143 HIV[-] women to study the long-term effectiveness of HPV vaccination on cervicovaginal, anal,
144 and oral HPV carriage in Rwandan WLWH in late 2021. We will compare HPV-vaccinated
145 WLWH to HPV-vaccinated HIV[-] women to measure the relative long-term effectiveness and
146 immunogenicity of HPV vaccination in WLWH. We will compare HPV-vaccinated WLWH to
147 HPV-unvaccinated WLWH to measure the reduction of HPV burden in WLWH attributable to
148 HPV vaccination. As a secondary goal, we will conduct a natural history study to investigate
149 determinants, including cervicovaginal microbiome, of short-term HPV persistence in young
150 WLWH and HIV[-] women living in a SSA setting.

METHODS

Study Population and Setting

At community health centers in Kigali, 2,271 women aged 18-28 years will be enrolled: 757 WLWH (Group 1) and 757 HIV[-] women (Group 2) who did receive HPV vaccination (birth cohorts 1996 and later) and 757 WLWH (Group 3) who did not receive HPV vaccination (birth cohorts before 1996). Most Rwandan women born in 2003 or later will have been vaccinated with two doses of Gardasil®, women born between 1996 and 2002 will have been vaccinated with three doses of Gardasil®, and women born in 1995 or earlier will be unvaccinated. Within the HPV-vaccinated Groups 1 and 2, the study will enroll at least 274 women who received three doses and 274 women who received two doses of the HPV vaccine, with the remaining 209 not selected for the number of doses and may have received three or two doses.

We will recruit by HIV status and age, which will serve as a useful and very good proxy for HPV-vaccination status, which otherwise would be difficult to determine in real time at enrollment. HPV-vaccination status then will be confirmed by retrospective review of the HPV vaccination records kept by the Rwanda Ministry of Health (MoH), who are collaborating on this project. The number of women recruited will therefore be stratified by age (as proxy for vaccine status) and HIV status. Because this study has age-specific enrollment goals for WLWH and HIV[-] women, once those enrollment goals are met for each study group, the respective groups will be closed and other eligible women for that study group will be excluded from participation.

In Kigali, the study staff will work with five public health facilities that participate in the Central Africa International Epidemiology Databases to Evaluate AIDS (IeDEA) program (ca-iedea.org) and the WE-ACTx private health facility (**Table 1**). If needed, the staff will work with up to four

Table 1. Study enrollment sites

IeDEA Sites	Non-IeDEA sites
1. Busanza health center (HC)	1. Cor-unum HC
2. Gikondo HC	2. Kacyiru HC
3. Kicukiro HC	3. Remera HC
4. Nyarugunga HC	4. Rwampara HC
5. Rwanda Military Hospital	
6. WE-ACTx private clinic	

other public health clinics to recruit additional participants. We will recruit women attending collaborating health facilities and from the surrounding communities.

Study Design

Eligibility Criteria. Women are eligible to participate if they: live in Rwanda; are ages 18-28 years who are known to be living with HIV or ages 18-25 years whose HIV status is unknown and consent to HIV testing to confirm HIV status; have had sex; are physically and mentally able and willing to participate in the study; and are willing to provide written and signed or thumb printed, informed consent. A summary of the target population by HIV and HPV vaccination statuses are shown in **Table 2**.

Women are ineligible to participate if they: have positive pregnancy test or report to be pregnant at the time of visit or less than 6 weeks post-partum (will be asked to make an appointment 6 or more weeks post-partum); have a history of hysterectomy and no longer have a cervix; have a history of treatment for cervical abnormalities after cervical cancer screening, have a history of cervical cancer; report no previous sexual activity; and/or HIV status is unknown and date of birth is 12/31/1995 or earlier. Women who report menstruating at the time of visit will be asked to make a new appointment 2 weeks later.

Table 2. Target population by HIV status and age

HIV Status	Approximate number of HPV vaccine doses	Birth year	Age at year of study enrollment (years)		N
			in 2021	in 2023	
WLWH	3 doses	1996-2002	19-25	21-26	274
	2 doses	2003-2005	16-18*	18-20	274
	2 or 3 doses	1996-2005	19-25	18-26	209
	0 doses - unvaccinated	1993-1995	26-30*	28-31*	757
HIV[-]	3 doses	1996-2002	19-25	21-26	274
	2 doses	2003-2005	16-18*	18-20	274
	2 or 3 doses	1996-2005	19-25	18-26	209
TOTAL					2,271

*Only women aged 18-28 are eligible for enrollment.

Enrollment Visit. The enrollment visit is summarized in **Figure 1**. After eligibility is confirmed and consent given, eligible, consenting women (participants) will provide a urine sample for pregnancy testing to confirm they are not pregnant. The consent form obtains the participants' permission to extract from medical records information related to their HIV care including CD4 count, viral load, ART use, and age of initiation of HIV care as well as HPV vaccination status.

Participants recruited into Group 2, whose current HIV status is unknown, will have a rapid HIV test followed by a confirmatory test if the initial test result is invalid (inconclusive) or reactive (positive) according to the Rwanda National HIV Diagnostic Testing Guidelines.^{52 53} If the newly HIV-diagnosed participant chooses not to exit the study, she will be reclassified into either Group 1 or 3, depending on HPV vaccination status.

Participants will complete a baseline questionnaire on socio-demographics and risk factors (e.g., sexual behavior) for HPV, undergo blood collection for HPV serology, and finally cervicovaginal, anal, and oral specimen collection for HPV testing.

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Specimen Collection. The study nurse first will collect one 5 mL tube of blood for EDTA plasma from all participants. For the oral specimen collection, participants will alternate between a 5-second squish and a 5-second gargle of 10 mL of saline for 30 seconds and then spit out the specimen into sterile specimen container. The study nurse will then collect a cervicovaginal specimen using the AmpFire specimen collection brush (Atila BioSystems, Mountain View, CA, USA), placing it into the posterior vagina without a speculum, turning the brush 3 turns left and right. Then the study nurse will withdraw the brush, place it into collection tube, snap off the handle to break it, and seal the collection tube. Finally, the study nurse will collect an anal specimen inserting a water-moistened Dacron swab into the anal canal, turning the swab 2-3 turns left and right, before removing the swab. The study nurse will insert the swab into the collection tube, snap off the handle, and seal the collection tube.

Six-Month Follow-Up Visit of HPV-positive Women. The follow-up visit is summarized in **Figure 2**. Participants positive for HPV on any sample will have a 6-month follow-up visit to measure 6-month HPV type-specific persistence - a surrogate endpoint recommended by the World Health Organization (WHO).⁵⁴ At young ages, the risk of precancer is very low, but for safety purposes, participants will be offered colposcopy if they have a 6-month HPV type-specific persistent high-risk HPV cervical infection and anoscopy if they have a 6-month HPV type-specific persistent HPV16/18 anal infection. At this follow-up visit, participants will since the last visit if HIV negative at baseline, a rapid HIV test. They then will have cervicovaginal, anal, and/or oral specimens collected only from those tissue sites that were HPV positive at baseline for HPV testing as described.

Clinical Management. A summary of the management of HPV-positive results is shown in **Figure 3**. Women with a 6-month, HPV type-specific persistent cervicovaginal HPV infection

for one or more of the 13 high risk HPV (hrHPV) types (HPV16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, and 68), regardless of HIV status, will be referred for colposcopy and, as needed, treatment.

Following application of dilute acetic acid to the cervix, digital images of the cervix will be taken using a study-provided contemporary digital camera (e.g., Samsung A21S or similar cellular phone camera). Then colposcopy-guided biopsies will be taken of all acetowhite, visible cervical abnormalities.⁵⁵ Colposcopic impression and biopsy locations will be recorded.

Biopsies will be read by a local pathologist (and a second one if second opinion is required) and a US pathologist. All participants with a biopsy diagnosis of CIN grade 3 (CIN3) or more severe diagnoses (CIN3+) as rendered by any pathologist and WLWH with persistent HPV16- or HPV18 infection will undergo treatment by ablation or, if ineligible for ablative treatments, excision according to WHO guidelines.⁵⁶⁻⁵⁸ In accordance with US management guidelines⁵⁹, those diagnosed with CIN2 will not undergo immediate treatment due the likelihood of its regression and low risk of ICC following a CIN2 diagnosis especially in young women⁶⁰, and the possible increased risk in negative reproductive outcomes such as pre-term delivery following treatment.⁶¹ They will be advised to seek clinical follow-up in a year.

WLWH with 6-month persistent anal HPV16 or HPV18 infection will undergo anoscopy; HIV[-] participants with 6-month persistent HPV16- or HPV18 infection will not undergo anoscopy because only ~10% of anal cancer is due to HPV16 or HPV18 among HIV⁶², the low absolute risk of anal cancer²⁷, and the possibility of morbidity from treating anal abnormalities. A biopsy will be taken from acetowhite lesions. The anoscopy impression and location of biopsies will be recorded for clinical management files and saved in the participant record for study purposes.

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3 253 Those with a biopsy diagnosis of anal intraepithelial neoplasia (AIN) grade 3 (AIN3) or anal
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5 254 cancer will be treated or referred for anal cancer management, respectively. Those diagnosed
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7 255 with AIN grade 2 (AIN2) will not undergo immediate treatment due to the low risk of anal
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9 256 cancer following a AIN2 diagnosis and the morbidity associated with treatment. Those with
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11 257 untreated AIN2 or 6-month persistent anal hrHPV by non-HPV16/18 types will be advised to
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13 258 seek clinical follow-up in a year.
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18 259 There are no management guidelines or evidence-based intervention for 6-month HPV type-
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20 260 specific persistent oral HPV infection. Therefore, participants will not receive any clinical
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22 261 intervention for HPV type-specific persistent oral infection.
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26 262 Other participants, including hrHPV-negative WLWH and/or women with 6-month persistent
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28 263 *low-risk* HPV infection, will exit the study without further follow-up visits. Those diagnosed
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30 264 with cervical or anal cancer will be referred for cancer care.
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34 265 *Study Outcomes.* Main outcomes will be prevalence and 6-month type-specific persistence of
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36 266 cervicovaginal (as an excellent proxy for cervical sampling⁶³), anal, and/or oral infections by
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38 267 HPV6/11/16/18 as well as anti-HPV16 and -HPV18 geometric mean titers. Cervical and anal
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40 268 biopsy specimens diagnosed as CIN2 or more severe diagnoses (CIN2+) or AIN2 or more severe
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42 269 diagnoses AIN2+, respectively, will also be tested for HPV for a secondary analysis to measure
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44 270 the effects of HPV vaccination on the prevalence of HPV type-specific precursors to anogenital
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46 271 cancer. In an exploratory aim, we will examine risk factors, including the cervicovaginal
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48 272 microbiome⁶⁴ and current and past HIV status [positive vs. negative and current antiretroviral
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50 273 therapy, CD4 counts, and HIV viral load], for HPV type-specific persistence in WLWH and
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52 274 HIV[-] women living in Rwanda.
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275 *HPV Testing.* We will use a modified version of the AmpFire HPV genotyping assay
276 (Genotyping High Risk HPV Real Time Fluorescent Detection; Atila Biosystems, Mountain
277 View, CA, USA) for HPV genotyping of cervicovaginal, anal, and oral specimens. AmpFire uses
278 real-time PCR to detect 15 individual HPV genotypes including 13 hrHPV types and 2
279 intermediate-risk HPV types (HPV53 and HPV66). The modified AmpFire HPV genotyping
280 assay will detect the 17 individual HPV genotypes, including vaccine targeted HPV6 and
281 HPV11, in 5 assay reactions. The AmpFire testing platform was previously established at the
282 study lab and validated against other commercially available assays at Rwanda Military Hospital
283 (RMH).⁶⁵⁻⁶⁸ The assay will be run per the manufacturer's instructions.

284 Anogenital samples, collected into a dry tube, will be processed directly according to the
285 manufacturer's instructions. Residual lysed anogenital specimens will be neutralized according
286 to the manufacturer's protocol and stored at -20°C for future use.

287 Upon arrival to the lab, the oral rinse specimens will be stored at 4°C for processing the next
288 working lab day. Oral rinse specimens in saline will first be concentrated by centrifugation to
289 enrich the sample before processing and then stored frozen at -20°C until tested for HPV. After
290 testing, residual specimens will be discarded.

291 Formalin-fixed paraffin-embedded tissues diagnosed as CIN2+ or AIN2+ will similarly have
292 HPV genotyping using the AmpFire system according to the manufacturer's protocol.

293 *HPV Serology.* Anti-HPV16 and HPV18 IgG antibody geometric mean titers (GMTs) will be
294 measured from plasma by an VLP-based ELISA using a previously described method.⁶⁹⁻⁷¹ A
295 total of randomly selected 1,096 sera (548 from Groups 1 and 2) from HPV-vaccinated
296 participants (stratified by HIV status and 3 vs. 2 HPV vaccine doses) and 108 sera from

297 unvaccinated participants (Group 3) will be run in total. First, 20% of each group will be run at
298 the HPV Serology Laboratory at the National Cancer Institute (NCI) Frederick National
299 Laboratories, then the results will be replicated ($\geq 90\%$ correlation) at the study lab in Rwanda
300 (masked to the original results), and then the remaining 80% of the testing will be finished at the
301 study lab. As an additional quality control measure, a 10% random sample of specimens will be
302 re-run at the HPV Serology Laboratory (masked to the original results) (inter-laboratory
303 reliability) and another 10% random sample of specimens at the study lab (intra-laboratory
304 reliability). Positive (e.g., IS standards^{72 73}) and negative controls (e.g., negative plasma) will be
305 included in some testing plates to monitor assay performance per WHO recommendations.⁷⁴

306 *Pathology.* All histopathology slides will be scanned at the study lab and reviewed by a local
307 pathologist, with a second review if second opinion is required, and then the Einstein study
308 pathologist.⁷⁵ If the diagnoses are concordant, no further review of the case will be performed. If
309 the diagnosis is discordant and at least one pathologist diagnoses CIN2+, the biopsy slide will be
310 subjected to a joint review and consensus diagnosis. For negative/cervical intraepithelial
311 neoplasia grade 1 pairs of diagnoses, which does not influence our analyses or the care of the
312 participants, there will be no joint review.

313 **Analysis and Statistical Power**

314 *Analyses.* We will calculate point prevalence and the prevalence of 6-month persisting infection
315 by individual HPV-type specific infections, with binomial 95% confidence intervals (95%CI),
316 for each anatomic site and for the woman (all anatomic sites) for Gardasil®-vaccinated WLWH,
317 Gardasil®-vaccinated HIV[-] women and unvaccinated WLWH. We likewise will calculate
318 point prevalence and the prevalence of 6-month persisting infection with 95%CI for all HPV

types in aggregate and in sub-groups of HPV types according to the protection afforded by Gardasil®: Gardasil®-targeted types (HPV6, 11, 16, and 18), Gardasil®-untargeted types for which there might be cross-protection (HPV31, 33, and 45)^{18 76 77}, and Gardasil®-untargeted HPV types for which there is little or no evidence of cross-protection (HPV35, 39, 51, 52, 53, 56, 58, 59, 66, and 68). Results will be stratified by age and number (3 vs. 2) of doses as well as other factors possibly related to HPV prevalence. Differences in prevalence of these HPV type sub-groups (targeted, possible cross-protection, and untargeted) between the three study groups of women will be tested using Fisher's exact or Pearson chi-square test. Differences in prevalence of these HPV type sub-groups by age, number of doses (3 vs. 2), and other factors within the group, will be tested for statistical significance ($p < 0.05$) using a Fisher's exact or Pearson chi-square test.

Notably, study groups of participants are fundamentally different populations (vs. a randomized control trial that would recruit from the same population and, as result of randomization, enroll similar, representative populations in each arm). Specifically, there are known differences in age and therefore possible differences in sexual activity between Gardasil®-vaccinated (Group 1) vs. -unvaccinated WLWH (Group 2), and possible differences in sexual behaviors between Gardasil®-vaccinated WLWH (Group 1) vs. HIV[-] women (Group 3) (since HIV infection predominately is sexually transmitted in this population). Therefore, we will use a relative measure of effectiveness to account for the differences in age and possible differences in exposure to HPV. We will use logistic regression to calculate the odds ratio (OR) of the point prevalence and the prevalence of 6-month persisting HPV infections of Gardasil®-targeted HPV types, individually (HPV6, 11, 16, *or* 18) and in aggregate (HPV6, 11, 16, *and* 18), vs. untargeted HPV genotypes for which there is no evidence of cross-protection, for each anatomic

site individually (cervix, anus, OR oral cavity) and combined (cervix, anus, AND oral cavity).
That is, we also will compare the ratio between study groups of participants as follows:

$$\frac{N_{\text{HPV6/11/16/18}}}{N_{\text{HPV35/39/51/52/53/56/58/59/66/68}}} [\text{vaccinated WLWH}] \text{ (Group 1)}$$
$$N_{\text{HPV6/11/16/18}}/N_{\text{HPV35/39/51/52/53/56/58/59/66/68}} [\text{unvaccinated WLWH or HIV[-] women}] \text{ (Group 2 or Group 3)}$$

This will help us to account/adjust for differences, above what can be achieved statistically, in HPV exposure, prevalence, and persistence due to differences in age (n.b., prevalence equals incidence times duration; prevalent infections tend to be more persistent with increasing age⁷⁸) and sexual behaviors between groups. Additional logistic regression models may be used to adjust specifically on other factors, including age, number of doses, and sexual behaviors, to account for population differences between study groups.

Differences in GMT and seropositivity will be tested for statistical significance using the Mann-Whitney and Fisher’s exact tests, respectively. ANOVA and logistic regression models will be used to adjust for/assess the association of other factors (e.g., age at vaccination, age at enrollment into the study, number of doses, current and past HIV status, and detection of HPV genotypes, etc.) with GMT and seropositivity, respectively.

Sample Size/Power. We made the following assumptions: 1) at least 30% prevalence of HPV infection (of any anatomic site) at baseline, 25% of which will by Gardasil®-target HPV types and 55% will be untargeted HPV types, 2) a 10% loss to follow-up (LTFU) in 6 months, and 3) at least 70% of HPV infections persist for 6 months. We justify an HPV prevalence of at least 30% among WLWH less than 30 years of age in this study, based on 30% prevalence of high-risk HPV infection of the cervix alone in 30-34 year-olds WLWH from our previous cervical cancer screening study in Rwanda.⁷⁹ We expect that the HPV prevalence may be higher since the

prevalence of HPV, like that of other sexually transmitted infections, tends to peak about 5-10 years after the median age of sexual initiation in a population^{80 81}, which in Rwanda is around 17 years of age.^{82 83} However, we conservatively used 30% prevalence to ensure adequate statistical power. We justify our assumption of a maximum of 10% loss to follow-up over 6 months, given our experience working in Rwanda over the last ~20 years. We justify 70% of prevalent HPV infections persisting for 6 months, given that 36-50% of prevalent HPV infections persist 6-months in HIV[-] women^{84 85} and WLWH have an impaired immunity to HPV compared to HIV[-] women.^{24 25}

Sample size calculation relates to the relative effectiveness measure as discussed. Under above assumptions and accounting for 10% misclassification of HPV vaccine status, a sample size of 757 vaccinated and 757 unvaccinated WLWH will provide $\geq 80\%$ power ($p=0.05$) to detect an OR of ≤ 0.5 for 6-month persistent HPV6/11/16/18 infections relative to HPV35/39/51/52/53/56/58/59/66/68 (but not HPV31/33/45 because of possible cross protection), in Gardasil®-vaccinated (Group 1) vs. -unvaccinated WLWH (Group 3). Consequently, because there will be more participants with a prevalent infection than 6-month persistent HPV infection (because there are no losses to follow-up or HPV viral clearance for which to account), there will be $\geq 80\%$ power ($p=0.05$) to detect a OR of ≤ 0.75 in point prevalence of HPV6/11/16/18 infection relative to HPV35/39/51/52/53/56/58/59/66/68, in Gardasil®-vaccinated (Group 1) vs. -unvaccinated WLWH (Group 3).

Using the variance of GMT for HPV antibodies from Einstein *et al.*^{86 87}, a sample size of 274 will provide 80% power ($p=0.05$) to detect a 30% difference in GMT between HPV-vaccinated WLWH (Group 1) and HIV[-] (Group 2) women for the same number of doses. If there is no appreciable difference in the GMT between those who got three or two doses, we can combine

those groups with different doses. In that case, with a sample size of 548, there will be 80% power ($p=0.05$) to detect a 22% difference between HPV-vaccinated WLWH (Group 1) and HIV[-] women (Group 2).

Patient and Public Involvement

Patients or the public WERE NOT involved in the design, or conduct, or reporting, or dissemination plans of our research

DISCUSSION

Limitations

Several limitations in the proposed study are worth noting. First, our study is not a randomized control trial, but an observational study of populations selected from different groups of women. We have proposed several approaches to account/adjust for those age differences but as with any observational study, these techniques may be unable to completely control for biases.

Populations recruited into this study are primarily from Kigali and therefore are not representative of all Rwanda, nor are they representative of other populations in SSA or elsewhere in the world. In 2018-9, almost 80% of Rwandan PLWH had a suppressed HIV viral load⁸⁸, a higher percentage than for PLWH populations living in many SSA and other countries.⁸⁹ Therefore, these results may not be generalizable to all WLWH populations, especially populations of WLWH who are severely immunocompromised.

We are using 6-month HPV type-specific persistence as a proxy for cancer risk but ideally, we want to measure the impact of Gardasil® on high-grade cervical and anal abnormalities as a

more proximal surrogate for cancer risk. The population will be still too young to have large numbers of these endpoints although we will evaluate these endpoints in secondary analyses.

Strengths

Despite the noted limitations, there are some important strengths of the study that warrant highlighting. First, the early introduction of HPV vaccination that is documented through a national registry, excellent HIV care including a national database and patient tracking, and relatively high prevalence of HIV provides a unique opportunity to study the impact of HPV vaccination in WLWH, the population at the highest risk of cervical^{29 30} that therefore would gain the greatest benefit from it, living in Rwanda. Although a comparative effectiveness randomized control trial of different HPV vaccines is probably still warranted to determine how best to protect this most vulnerable population⁹⁰, these data may provide some of the first evidence of long-term effectiveness of HPV vaccination in WLWH and inform such a trial. A second strength is that the study is being conducted in Rwanda, a high cervical-cancer burden country.⁴⁵ The findings from this study, if positive, may encourage other high-burden countries to accelerate the introduction of HPV vaccination or at least target WLWH and PLWH populations. Finally, HPV DNA and serology testing are being done locally for this study could potentially be transferred and replicated in other LMICs that want to monitor and evaluate HPV-vaccine impact in their populations.

Importantly, as a result of implementing this research protocol, we will continue to expand the local capacity to conduct state-of-the-art HPV and molecular epidemiologic research by 1) establishing next-generation sequencing (NGS) technology to perform cervicovaginal, anal, and oral microbiota characterization for this study that will enable locally conducted studies of the

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human genome and genomic testing for personalized medicine in Rwanda; 2) building upon current ELISA capabilities at the study lab to perform titration of plasma antibodies, a skill that can be applied to other studies of vaccine response; 3) enhancing data capture and management through increased capacity to use REDCap⁹¹⁻⁹³; and 4) migrating current HPV vaccine records for participants living in Kigali into a common electronic database, which will allow us to conduct studies more easily on the impact of HPV vaccination on outcomes. Importantly, this will allow linkage of HPV-vaccination status to both the Rwanda National HIV Registry and the Rwanda Cancer Registry, the latter of which we, in collaboration with the Rwanda MoH, helped re-establish.⁹⁴ This will allow investigations of the long-term impact of HPV vaccination on cancer incidence in Rwandan WLWH and HIV[-] women and, as a result of herd protection, Rwandan men.

As building research capacity in Rwanda is a major goal of this project, all members of the research team will be asked and supported to lead at least one analysis and one manuscript preparation, based on interests and expertise. Analytic and publication responsibilities will be divided equally and collaboratively among both Rwandan and U.S. investigators.

ETHICS AND DISSEMINATION

The study was approved by the Albert Einstein College of Medicine Institutional Review Board (IRB#: 2021-13087) and the Rwanda National Ethics Committee.

All participant data collected will be entered or transferred to a secure REDCap database, with access to personal information restricted to the local staff. Study data will be stored in REDCap and maintained on password protected study computers behind the institution firewall.

Copies of signed consents will be stored in locked file cabinets in a locked room, with access restricted to study personnel only, at the study lab. Data will be entered or transferred to a secure REDCap database, with access to personal information restricted to the local staff.

Study data will be stored in REDCap and maintained on password protected study computers behind the institution firewall. REDCap is a secure, web-based application designed to support data capture for research studies providing: 1), an intuitive interface for validated data entry; 2) audit trails for tracking data manipulation and export procedures; 3) automated export procedures for seamless data downloads to common statistical packages; and 4) procedures for importing data from external sources. REDCap is hosted on a secure server and has undergone a Governance Risk & Compliance Assessment by All REDCap electronic data files shared with Albert Einstein College of Medicine will be maintained by the HIPAA-compliant Epidemiology Study Management and Informatics Core Facility (ESMI) at Einstein. The Albert Einstein College of Medicine policy on use of REDCap can be found at: http://ric.einstein.yu.edu/ric_files/REDCap%20Appropriate%20Use%20Policy.pdf

Medical, screening, and preventive services, all of which are minimally invasive, safe, and outpatient, and have been done in millions of people, provided by the study are on par with or better than international standards-of-care, most of which have very low risks of even minor adverse events or harms. Positive test results and diagnoses may result in psychologic distress and anxiety. Of note, pregnancy and HIV testing may cause pre- and post-test anxiety. Therefore, pre- and post-testing counseling will be provided as needed. Any participant who tests HIV positive will be referred to the health facility's HIV clinic for HIV management following the Rwanda MoH's HIV management guidelines.

We will publish a series of reports in peer-review scientific journals to disseminate these results. We will share the results with the Rwanda MoH. Data from this study will be made available by request in accordance with NIH policies and Rwandan laws.

ETHICS STATEMENT

Patient consent for publication not required.

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- **Contributorship statement:** GM, PEC, and KA designed the study initially. GM, AP, PEC, and KA refined and finalized the overall study design and wrote the grant proposal that is the basis for this protocol. GM, FS, NH, AM, JCG, AP, KA, and PEC drafted and finalized the written protocol. GM, FS, NH, AM, JCG, BM, FK, AP, PT, KA, and PEC implemented it. PEC drafted this manuscript. All authors have read, revised, and approved the final manuscript. GM and FS are co-first authors; KA and PEC are co-senior authors.
- **Competing interests statement:** Dr. Castle is the Director of the Division of Cancer Prevention at the NCI, the NIH institute that funds this research. However, Dr. Castle has recused himself from any decisional or financial authority over this grant or any extramural HPV grants funded by the NCI. Other authors claim no competing interests. The study is receiving HPV genotyping tests at a reduce cost from Atila Biosystems.
- **Funding statement:** This work is being supported by NIH by grant numbers 5U54CA254568-02 and 2U01AI096299-13. This work is also being supported by the intramural research program of the National Cancer Institute/NIH.
- **Data sharing statement:** Data will be shared in accordance to policies for NIH-funded research at the conclusion of the study and after the publication of the main scientific findings.

Figures

Figure 1. Enrollment visit procedures

Figure 2. Six-month follow-up visit procedures

Figure 3. Follow-up and clinical management of baseline HPV-positive results

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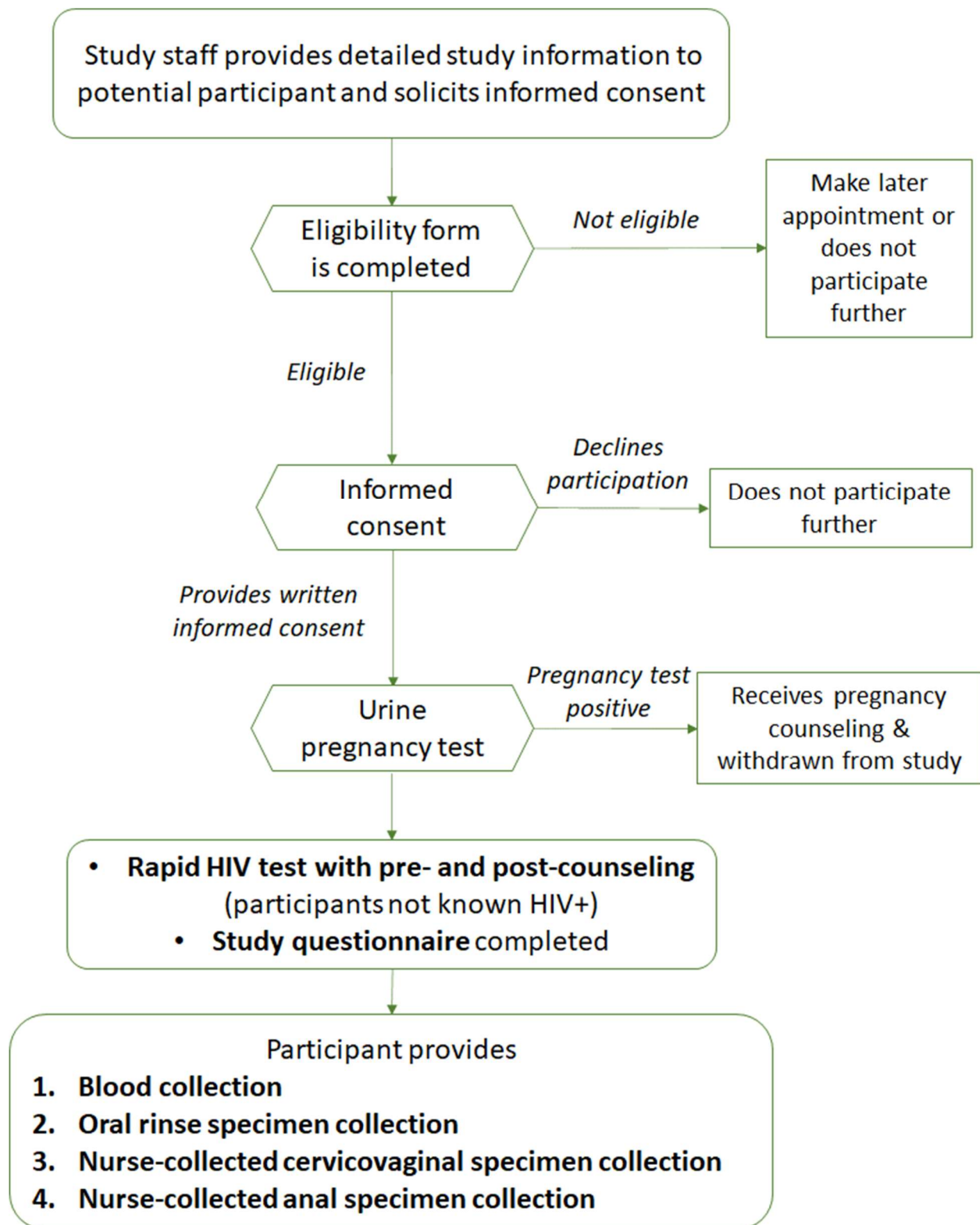
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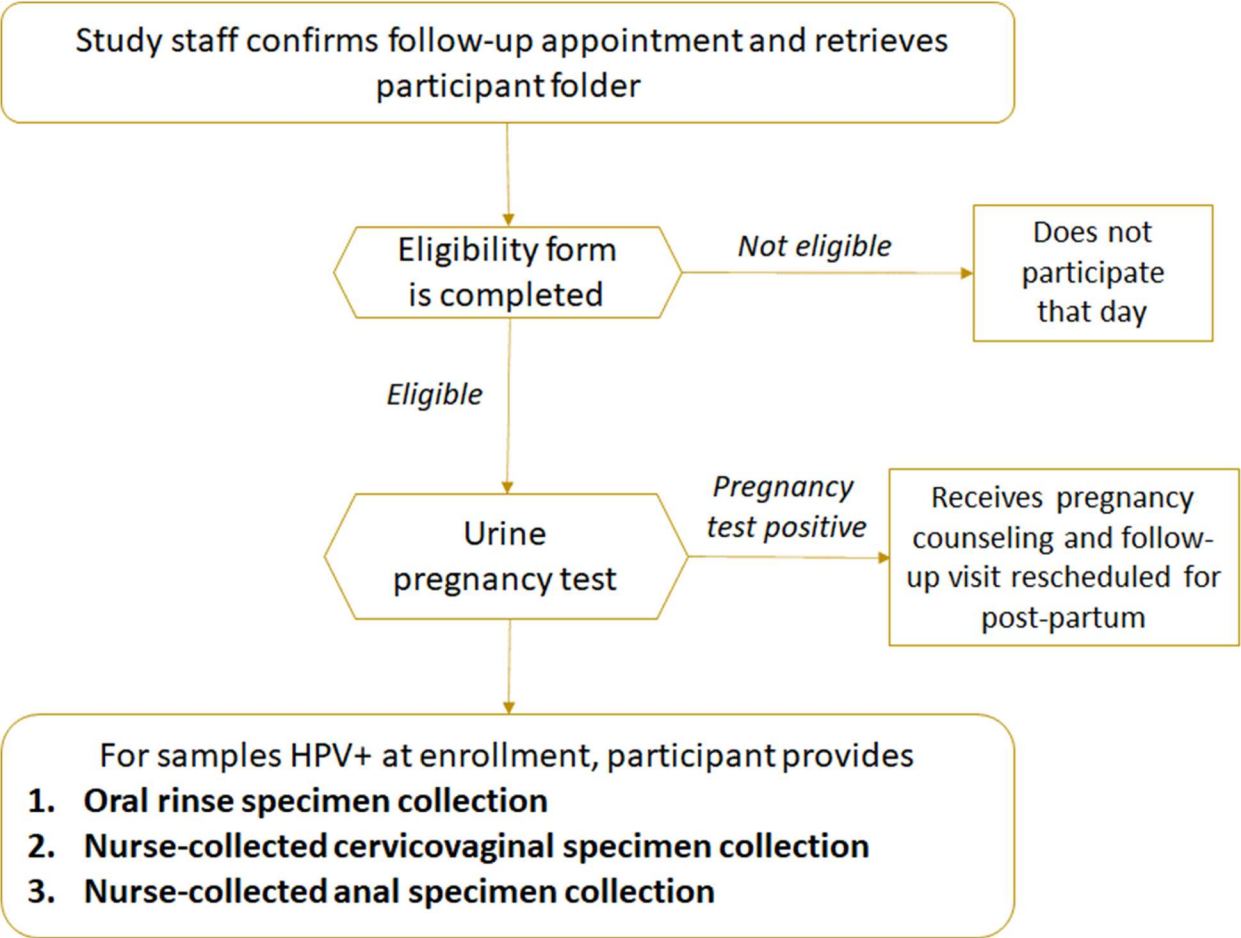
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Baseline Cervicovaginal, Oral, and/or Anal Specimen(s) Test HPV+ (1st visit)

6-Month Follow-Up Visit (2nd visit)

Oral HPV+

Anal HPV+

Cervicovaginal HPV+

Collect & Test Oral Specimen

Collect & Test Anal Specimen

Collect & Test Cervicovaginal Specimen

WLWH & Persistent HPV16/18+

Type-Specific hrHPV Persistence?

No

Yes

Anoscopy & Biopsy of Acetowhite Lesions (3rd visit)

Colposcopy & Biopsy of All Acetowhite Lesions (3rd visit)

AIN3+

WLWH & Persistent HPV16/18+

CIN3+

Treatment (4th visit)

Ablative Treatment {colpo visit}

Ablation Ineligible

Treatment (4th visit)

hrHPV+ = positive for HPV16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, and/or 68
 HPV+= hrHPV+ and/or positive for HPV53, HPV66, HPV6, and/or HPV11+

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Long-Term Human Papillomavirus Vaccination Effectiveness and Immunity in Rwandan Women Living with and without Human Immunodeficiency Virus: A Study Protocol

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Manuscripts

Long-Term Human Papillomavirus Vaccination Effectiveness and Immunity in Rwandan Women Living with and without Human Immunodeficiency Virus: A Study Protocol

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[ClinicalTrials.gov](https://clinicaltrials.gov) Identifier: NCT05247853

Abstract

Introduction. Prophylactic human papillomavirus (HPV) vaccines have been shown to be highly effective in protecting women against cervical infections, high-grade abnormalities, and cancer caused by the targeted HPV types. However, the evidence for their effectiveness in women living with human immunodeficiency virus (HIV) (WLWH) is less clear.

Methods. WLWH and HIV-negative women who likely did (birth cohorts 1996 and later) and WLWH and HIV[-] negative who likely did not (birth cohorts before 1996) receive HPV vaccination (n=3,028; 757 participants for each of the four groups). Between groups, we will compare cervicovaginal, anal, and oral prevalent and 6-12-month persistent HPV6/11/16/18 infections as measured using a modified AmpFire HPV genotyping assay that tests for 15 high-risk or intermediate-risk HPV genotypes, HPV6, and HPV11. We will also compare the HPV immune response in HPV-vaccinated WLWH to HPV-vaccinated HIV-negative women using an anti-HPV16 and -HPV18 ELISA. Vaccination status will be confirmed through national vaccination records.

Analysis. We will calculate point prevalence and prevalence of 6-12-month persisting infections by individual HPV-type specific infections and groups of infections for each anatomic site and for each group of women. Results will be stratified by age at vaccination, age at enrollment, and the number of doses (3 vs. 2) as well as other factors possibly associated with HPV prevalence. Differences in endpoints between groups, overall and between sub-groups, will be tested for statistical significance ($p < 0.05$) using Fisher's exact or Pearson chi-square test. Differences in geometric mean titers (GMT) and seropositivity will be tested for statistical significance using the Mann-Whitney and Fisher's exact tests, respectively.

Ethics and Dissemination. The study was approved by the Albert Einstein College of Medicine Institutional Review Board and the Rwanda National Ethics Committee. Results will be disseminated through publication in peer-reviewed journals.

Key Words: Human papillomavirus (HPV), vaccination, cervical, anal, oral, cancer, Rwanda

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STRENGTHS AND LIMITATIONS OF THIS STUDY

- The study is being conducted in Rwanda, a high cervical-cancer burden country
- National introduction of HPV vaccine in Rwanda in 2011 and relatively high prevalence of HIV makes a study of long-term HPV vaccination effective in women living with HIV (WLWH) possible.
- The study is not a randomized control trial, but an observational study of populations selected from different groups of women.
- A convenience sample of Rwandan WLWH living in and around Kigali will be enrolled and therefore the study population is not representative of all Rwanda or WLWH living elsewhere.
- The quality of HPV vaccination status ascertainment is uncertain given the use of paper registers and the long time period that has elapsed since vaccination.

INTRODUCTION

Current prophylactic vaccines against human papillomavirus (HPV), the necessary cause of virtually all cervical cancer [1, 2], are based on the self-assembly of recombinantly expressed L1 protein in cell lines into virus-like particles (VLPs) that resemble native viral capsids but lack the viral genome required for viral replication and infectivity. The first generation of prophylactic HPV vaccines, Gardasil® (Merck & Co, Kenilworth, NJ, USA)[3] and Cervarix™ (GlaxoSmithKline, Wavre, Belgium)[3], targeted HPV16 and HPV18 (HPV16/18), which cause approximately 70% of cervical cancers [4]. Gardasil® also targets HPV6 and HPV11 (HPV6/11), non-high-risk HPV types responsible for approximately 90% of anogenital warts (*Condylomata acuminata*)[5].

Several countries, including Australia [6-12], Scotland [13], Denmark [14], and the U.S. [15-17], were early adopters of HPV vaccination and have documented reductions in infections, diseases, and cervical abnormalities related to the HPV vaccine-targeted types. A meta-analysis on the impact of HPV vaccination found reductions in anogenital warts, HPV infections, cervical intraepithelial neoplasia (CIN) grade 2 (CIN2) or more severe diagnoses (CIN2+) among girls and women, and on anogenital warts diagnoses among girls, women, boys, and men [18]. Recent reports from Finland [19], Sweden [20], England [21], and Denmark [22] provide real-world evidence that HPV vaccination prevents invasive cervical cancer.

Cervical cancer was included as an acquired immunodeficiency syndrome (AIDS)-defining disease in adolescents and adults in 1993 [23, 24]. Women living with human immunodeficiency (WLWH) virus, the cause of AIDS, have a significantly elevated risk of cervical cancer [25, 26], due to an impaired immune response to HPV, compared to HIV-uninfected (HIV[-]) women.

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91 Meta-analyses of HIV/AIDS cohorts reported a 6-fold increased incidence of cervical cancer
92 compared to the general female population/HIV[-] women [25-27]. People living with HIV
93 (PLWH) are also at an increased risk of other HPV-related malignancies, notably anal and
94 oropharyngeal cancers [25, 28]. HIV co-infection has a profound impact on the natural history of
95 HPV, thereby increasing the risk of ICC. HIV co-infection increases the 1) likelihood of cervical
96 and anal HPV persistence and 2) likelihood of cervical and anal HPV infections progressing to
97 precancer [29, 30].

98 Data on HPV-vaccine efficacy and effectiveness in WLWH are lacking [31, 32]. To date, most
99 studies of HPV vaccine in WLWH focused on immunogenicity and safety. Generally, HPV
100 vaccination in PLWH has been well tolerated, safe, and resulted in good immune responses [33-
101 37]; one study found lower seroconversion and anti-HPV antibody titers among PLWH
102 compared to perinatally HIV-exposed, uninfected youth [37]. Studies have noted an impact of
103 HIV disease status (CD4 counts and viral suppression) on the immune responses to HPV
104 vaccination [38-40]. Studies have reported lower seroconversion and antibody titers in WLWH
105 not taking (vs. taking) antiretroviral therapy (ART) and with lower (vs. higher) CD4 counts [34,
106 39]. Similarly, another study found peak antibody titers to be 2- to 3-fold higher in mid-adult
107 WLWH with full HIV viral suppression compared to those not suppressed [40]. Interestingly,
108 higher anti-HPV18 titers but similar anti-HPV16 titers were reported in response to HPV
109 vaccination by Cervarix® compared to Gardasil® in PLWH but the differences in the immune
110 responses were due to differences in women (WLWH) but not men [41].

111 The few studies of HPV vaccine effectiveness in WLWH provided promising but inconclusive
112 evidence. Immunogenicity studies in WLWH have been of insufficient sample size to address
113 efficacy/effectiveness. Studies of HPV vaccination in select PLWH at high risk of anal cancer

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3 114 have been limited in sample size because of the high HPV anal exposure (and likely
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5 115 misclassification of exposure) to targeted HPV types prior to enrollment [42] so that few truly
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7 116 incident events could be observed. Of those studies that did have endpoints, one recent trial of
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9 117 Gardasil® in PLWH aged 27 years and older concluded that “This double-blind, randomized
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11 118 trial did not find a benefit from HPV vaccination to prevent anal HPV infection or anal high-
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13 119 grade squamous intraepithelial lesions” and favorable but inconclusive benefit for protection
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15 120 against oral HPV [43].
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20 121 The paucity of HPV-vaccine effectiveness data in WLWH is especially a concern for the
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22 122 prevention of cervical cancer in sub-Saharan Africa (SSA), where cervical cancer is the most
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24 123 common cause of cancer death in women [44]. Almost one quarter of the global burden of
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26 124 cervical cancer occurs in SSA and an estimated 20% of cervical cancer in SSA is attributable to
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28 125 HIV coinfection [27]. Evidence of the protective effects of HPV vaccination in SSA WLWH is
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30 126 needed, especially since most long-term public health planning to address cervical cancer in SSA
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32 127 depends on the unproven effectiveness of these vaccines.
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37 128 Rwanda, a central/east African country that experiences a high burden of cervical cancer [45], is
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39 129 the ideal locale to answer questions about the long-term effectiveness and immunogenicity of
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41 130 Gardasil® in WLWH for the following reasons. First, in 2011-13, Rwanda, through a donation
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43 131 from Merck, launched a national HPV vaccination program [46]. In 2011, over 92,000 girls in
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45 132 primary school grade six (~12 years old) were vaccinated with three doses of Gardasil®. During
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47 133 2012 and 2013, a catch-up vaccination program targeted girls in secondary school grade three
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49 134 (~15 years old) [46]. In 2014, HPV vaccination was supported by GAVI [47] and reverted to
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51 135 vaccinating 12-year-old girls [48]. In 2015, Rwanda switched from 3 doses to 2 doses, 6 months
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55 136 apart, for vaccinating 12-year-old girls. In all years, Rwanda achieved $\geq 90\%$ annual coverage
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137 with the recommended number of HPV vaccine doses in the target population [46, 47, 49, 50].

138 Thus, Rwanda is one of the earliest and most successful adopters of HPV vaccination globally.

139 Second, the prevalence of HIV in Rwanda is 2.7% among 15-30 year olds [51]. Approximately

140 half of the Rwanda population of 13 million is female, and one-quarter of them are between the

141 age of 15-30 years. Thus, there is approximately 450,000 WLWH aged 15-30 years living in

142 Rwanda [52]. The excellent HIV care program in Rwanda allows for easy and efficient

143 recruitment and following of WLWH in an observational study of HPV-vaccination impact.

144 Third, given Rwanda's nearly complete national coverage with Gardasil®, HPV-vaccinated and

145 unvaccinated WLWH are easily recruited based on birth year. Rwanda has excellent national

146 vaccination records so that retrospectively the small number of participants misclassified by

147 HPV vaccination status based on birth year subsequently can be correctly categorized.

148 The Einstein/Rwanda/DRC Consortium for Research in HIV/HPV/Malignancies therefore

149 launched an observational cohort of HPV-vaccinated and -unvaccinated WLWH and vaccinated

150 and unvaccinated HIV[-] women to study the long-term effectiveness of HPV vaccination on

151 cervicovaginal, anal, and oral HPV carriage in Rwandan WLWH in late 2021. We will compare

152 HPV-vaccinated WLWH to HPV-vaccinated HIV[-] women to measure the relative long-term

153 effectiveness and immunogenicity of HPV vaccination in WLWH. We will compare HPV-

154 vaccinated WLWH to HPV-unvaccinated WLWH to measure the reduction of HPV burden in

155 WLWH attributable to HPV vaccination. As a secondary goal, we will conduct a natural history

156 study to investigate determinants, including cervicovaginal microbiome, of short-term HPV

157 persistence in young WLWH and HIV[-] women living in a SSA setting.

METHODS

Study Population and Setting

At community health centers in Kigali, 3,028 women aged 18-28 years will be enrolled: 757 WLWH (Group 1) and 757 HIV[-] women (Group 2) who may have received HPV vaccination (birth cohorts 1996 and later) and 757 WLWH (Group 3) and 757 HIV[-] women (Group 4) who were older and unlikely to have received HPV vaccination (birth cohorts before 1996). Most Rwandan women born in 2003 or later will have been vaccinated with two doses of Gardasil®, women born between 1996 and 2002 will have been vaccinated with three doses of Gardasil®, and women born in 1995 or earlier will be unvaccinated. Within the HPV-vaccinated Groups 1 and 2, the study will enroll at least 274 women who received three doses and 274 women who received two doses of the HPV vaccine, with the remaining 209 not selected for the number of doses and may have received three or two doses.

We will recruit by HIV status and age, which will serve as a useful and very good proxy for HPV-vaccination status, which otherwise would be difficult to determine in real time at enrollment. HPV-vaccination status then will be confirmed by retrospective review of the HPV vaccination records kept by the Rwanda Ministry of Health (MoH), who are collaborating on this project. The number of women recruited will therefore be stratified by age (as proxy for vaccine status) and HIV status. Because this study has age-specific enrollment goals for WLWH and HIV[-] women, once those enrollment goals are met for each study group, the respective groups will be closed and other eligible women for that study group will be excluded from participation.

In Kigali, the study staff will work with five public health facilities that participate in the Central Africa International Epidemiology Databases to Evaluate AIDS (IeDEA) program (ca-iedea.org) and the WE-ACTx private health facility (**Table 1**). If needed, the staff will work with up to four

Table 1. Study enrollment sites

IeDEA Sites	Non-IeDEA sites
1. Busanza health center (HC)	1. Cor-unum HC
2. Gikondo HC	2. Kacyiru HC
3. Kicukiro HC	3. Remera HC
4. Nyarugunga HC	4. Rwampara HC
5. Rwanda Military Hospital	
6. WE-ACTx private clinic	

other public health clinics to recruit additional participants. We will recruit women attending collaborating health facilities and from the surrounding communities.

Study Design

Eligibility Criteria. Women are eligible to participate if they: live in Rwanda; are ages 18-28 years who are either known to be living with HIV or, if HIV status is unknown, consent to HIV testing to confirm HIV status; have had sex; are physically and mentally able and willing to participate in the study; and are willing to provide written and signed or thumb printed, informed consent. A summary of the target population by HIV and HPV vaccination statuses are shown in **Table 2**.

Women are ineligible to participate if they: have positive pregnancy test or report to be pregnant at the time of visit or less than 6 weeks post-partum (will be asked to make an appointment 6 or more weeks post-partum); have a history of hysterectomy and no longer have a cervix; have a history of treatment for cervical abnormalities after cervical cancer screening, have a history of

cervical cancer; report no previous sexual activity. Women who report menstruating at the time of visit will be asked to make a new appointment 2 weeks later.

Table 2. Target population by HIV status and age

HIV Status	Approximate number of HPV vaccine doses	Birth year	Age at year of study enrollment (years)		N	Group
			in 2021	in 2023		
WLWH	3 doses	1996-2002	19-25	21-26	274	1
	2 doses	2003-2005	16-18*	18-20	274	
	2 or 3 doses	1996-2005	19-25	18-26	209	
	subtotal				757	
	0 doses - unvaccinated	1993-1995	26-30*	28-31*	757	3
HIV[-]	3 doses	1996-2002	19-25	21-26	274	2
	2 doses	2003-2005	16-18*	18-20	274	
	2 or 3 doses	1996-2005	19-25	18-26	209	
	subtotal				757	
	0 doses - unvaccinated	1993-1995	26-30*	28-31*	757	4
TOTAL					3,028	

*Only women aged 18-28 are eligible for enrollment.

Enrollment Visit. The enrollment visit is summarized in **Figure 1**. After eligibility is confirmed and consent given, eligible, consenting women (participants) will provide a urine sample for pregnancy testing to confirm they are not pregnant. The consent form obtains the participants' permission to extract from medical records information related to their HIV care including CD4 count, viral load, ART use, and age of initiation of HIV care as well as HPV vaccination status. Participants recruited into Group 2 and 4, whose current HIV status is unknown, will have a rapid HIV test followed by a confirmatory test if the initial test result is invalid (inconclusive) or reactive (positive) according to the Rwanda National HIV Diagnostic Testing Guidelines.[53, 54] If the newly HIV-diagnosed participant chooses not to exit the study, she will be reclassified into either Group 1 or 3, depending on HPV vaccination status.

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212 Participants will complete a baseline questionnaire on socio-demographics and risk factors (e.g.,

213 sexual behavior) for HPV, undergo blood collection for HPV serology, and finally

214 cervicovaginal, anal, and oral specimen collection for HPV testing.

215 *Specimen Collection.* The study nurse first will collect one 5 mL tube of blood for EDTA plasma

216 from all participants. For the oral specimen collection, participants will alternate between a 5-

217 second squish and a 5-second gargle of 10 mL of saline for 30 seconds and then spit out the

218 specimen into sterile specimen container. The study nurse will then collect a cervicovaginal

219 specimen using the AmpFire specimen collection brush (Atila BioSystems, Mountain View, CA,

220 USA), placing it into the posterior vagina without a speculum, turning the brush 3 turns left and

221 right. Then the study nurse will withdraw the brush, place it into collection tube, snap off the

222 handle to break it, and seal the collection tube. Finally, the study nurse will collect an anal

223 specimen inserting a water-moistened Dacron swab into the anal canal, turning the swab 2-3

224 turns left and right, before removing the swab. The study nurse will insert the swab into the

225 collection tube, snap off the handle, and seal the collection tube.

226 *Six to twelve Month Follow-Up Visit of HPV-positive Women.* The follow-up visit is summarized

227 in **Figure 2**. Participants positive for HPV on any sample will have a 6-12-month follow-up visit

228 to measure 6-12-month HPV type-specific persistence - a surrogate endpoint recommended by

229 the World Health Organization (WHO) [55]. At young ages, the risk of precancer is very low,

230 but for safety purposes, participants will be offered colposcopy if they have a 6-12-month HPV

231 type-specific persistent high-risk HPV cervical infection and anoscopy if they have a 6-12-month

232 HPV type-specific persistent HPV16/18 anal infection. At this follow-up visit, participants will

233 complete a brief sexual history questionnaire to identify likelihood of potential new exposure to

234 HPV. If HIV negative at baseline, participants will also have a rapid HIV test. They then will

have cervicovaginal, anal, and/or oral specimens collected only from those tissue sites that were HPV positive at baseline for HPV testing as described.

Clinical Management. A summary of the management of HPV-positive results is shown in

Figure 3. Women with HPV type-specific persistent cervicovaginal HPV infection for one or more of the 13 high risk HPV (hrHPV) types (HPV16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, and 68), regardless of HIV status, will be referred for colposcopy and, as needed, treatment.

Following application of dilute acetic acid to the cervix, digital images of the cervix will be taken using a study-provided contemporary digital camera (e.g., Samsung A21S or similar cellular phone camera). Then colposcopy-guided biopsies will be taken of all acetowhite, visible cervical abnormalities [56]. Colposcopic impression and biopsy locations will be recorded.

Biopsies will be read by a local pathologist (and a second one if second opinion is required in order to guide care) and a US pathologist. All participants with a biopsy diagnosis of CIN grade 3 (CIN3) or more severe diagnoses (CIN3+) as rendered by any pathologist and WLWH with persistent HPV16- or HPV18 infection will undergo treatment by ablation or, if ineligible for ablative treatments, excision according to WHO guidelines [57-59]. In accordance with US management guidelines [60], those diagnosed with CIN2 will not undergo immediate treatment due the likelihood of its regression and low risk of ICC following a CIN2 diagnosis especially in young women [61], and the possible increased risk in negative reproductive outcomes such as pre-term delivery following treatment [62]. They will be advised to seek clinical follow-up in a year.

WLWH with persistent anal HPV16 or HPV18 infection will undergo anoscopy; HIV[-]

participants with persistent HPV16- or HPV18 infection will not undergo anoscopy because only

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3 257 ~10% of anal cancer is due to HPV16 or HPV18 among HIV[63], the low absolute risk of anal
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5 258 cancer [28], and the possibility of morbidity from treating anal abnormalities. A biopsy will be
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7 259 taken from acetowhite lesions. Anoscopy impression and location of biopsies will be recorded
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10 260 for clinical management files and saved in the participant record for study purposes. Those with
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12 261 a biopsy diagnosis of anal intraepithelial neoplasia (AIN) grade 3 (AIN3) or anal cancer will be
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14 262 treated or referred for anal cancer management, respectively. Those diagnosed with AIN grade 2
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16 263 (AIN2) will not undergo immediate treatment due to the low risk of anal cancer following a
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18 264 AIN2 diagnosis and the morbidity associated with treatment. Those with untreated AIN2 or 6-
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20 265 12-month persistent anal hrHPV by non-HPV16/18 types will be advised to seek clinical follow-
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22 266 up in a year.
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27 267 There are no management guidelines or evidence-based intervention for 6-12-month HPV type-
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29 268 specific persistent oral HPV infection. Therefore, participants will not receive any clinical
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31 269 intervention for HPV type-specific persistent oral infection.
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35 270 Other participants, including hrHPV-negative WLWH and/or women with 6-12-month persistent
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37 271 low-risk HPV infection, will exit the study without further follow-up visits. Those diagnosed
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39 272 with cervical or anal cancer will be referred for cancer care.
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43 273 *Study Outcomes.* Main outcomes will be prevalence and 6-12-month type-specific persistence of
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45 274 cervicovaginal (as an excellent proxy for cervical sampling [64]), anal, and/or oral infections by
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47 275 HPV6/11/16/18 as well as anti-HPV16 and -HPV18 geometric mean titers. Cervical and anal
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49 276 biopsy specimens diagnosed as CIN2 or more severe diagnoses (CIN2+) or AIN2 or more severe
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51 277 diagnoses AIN2+, respectively, will also be tested for HPV for a secondary analysis to measure
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53 278 the effects of HPV vaccination on the prevalence of HPV type-specific precursors to anogenital
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279 cancer. In an exploratory aim, we will examine risk factors, including the cervicovaginal
280 microbiome [65] and current and past HIV status [positive vs. negative and current antiretroviral
281 therapy, CD4 counts, and HIV viral load], for HPV type-specific persistence in WLWH and
282 HIV[-] women living in Rwanda.

283 *HPV Testing.* We will use a modified version of the AmpFire HPV genotyping assay
284 (Genotyping High Risk HPV Real Time Fluorescent Detection; Atila Biosystems, Mountain
285 View, CA, USA) for HPV genotyping of cervicovaginal, anal, and oral specimens. AmpFire uses
286 real-time PCR to detect 15 individual HPV genotypes including 13 hrHPV types and 2
287 intermediate-risk HPV types (HPV53 and HPV66). The modified AmpFire HPV genotyping
288 assay will detect the 17 individual HPV genotypes, including vaccine targeted HPV6 and
289 HPV11, in 5 assay reactions. The AmpFire testing platform was previously established at the
290 study lab and validated against other commercially available assays at Rwanda Military Hospital
291 (RMH)[66-69]. The assay will be run per the manufacturer's instructions.

292 Anogenital samples, collected into a dry tube, will be processed directly according to the
293 manufacturer's instructions. Residual lysed anogenital specimens will be neutralized according
294 to the manufacturer's protocol and stored at -20°C for future use.

295 Upon arrival to the lab, the oral rinse specimens will be stored at 4°C for processing the next
296 working lab day. Oral rinse specimens in saline will first be concentrated by centrifugation to
297 enrich the sample before processing and then stored frozen at -20°C until tested for HPV. After
298 testing, residual specimens will be discarded.

299 Formalin-fixed paraffin-embedded tissues diagnosed as CIN2+ or AIN2+ will similarly have
300 HPV genotyping using the AmpFire system according to the manufacturer's protocol.

HPV Serology. Anti-HPV16 and HPV18 IgG antibody geometric mean titers (GMTs) will be measured from plasma by an VLP-based ELISA using a previously described method [70-72]. All serum from Groups 1 and 2 (HPV-vaccinated groups) and a 10% sample from Groups 3 and 4 (HPV-unvaccinated groups (n=152, 76 from each group) will be tested. First, 20% of each group will be run at the HPV Serology Laboratory at the National Cancer Institute (NCI) Frederick National Laboratories, then the results will be replicated ($\geq 90\%$ correlation) at the study lab in Rwanda (masked to the original results), and then the remaining 80% of the testing will be finished at the study lab. As an additional quality control measure, a 10% random sample of specimens will be re-run at the HPV Serology Laboratory (masked to the original results) (inter-laboratory reliability) and another 10% random sample of specimens at the study lab (intra-laboratory reliability). Positive (e.g., IS standards [73, 74]) and negative controls (e.g., negative plasma) will be included in some testing plates to monitor assay performance per WHO recommendations [75].

Pathology. All histopathology slides will be scanned at the study lab and reviewed by a local pathologist, with a second review if second opinion is required, and then the Einstein study pathologist.[76] If the diagnoses are concordant, no further review of the case will be performed. If the diagnosis is discordant and at least one pathologist diagnoses CIN2+, the biopsy slide will be subjected to a joint review and consensus diagnosis. For negative/cervical intraepithelial neoplasia grade 1 pairs of diagnoses, which does not influence our analyses or the care of the participants, there will be no joint review.

Analysis and Statistical Power

Analyses. We will calculate point prevalence and the prevalence of persisting infection by individual HPV-type specific infections, with binomial 95% confidence intervals (95%CI), for each anatomic site and for the woman (all anatomic sites) for all 4 groups. We likewise will calculate point prevalence and the prevalence of persisting infection with 95%CI for all HPV types in aggregate and in sub-groups of HPV types according to the protection afforded by Gardasil®: Gardasil®-targeted types (HPV6, 11, 16, and 18), Gardasil®-untargeted types for which there might be cross-protection (HPV31, 33, and 45)[18, 77, 78], and Gardasil®-untargeted HPV types for which there is little or no evidence of cross-protection (HPV35, 39, 51, 52, 53, 56, 58, 59, 66, and 68). Results will be stratified by age and number (3 vs. 2) of doses as well as other factors possibly related to HPV prevalence. Differences in prevalence of these HPV type sub-groups (targeted, possible cross-protection, and untargeted) between the four study groups of women will be tested using Fisher's exact or Pearson chi-square test. Differences in prevalence of these HPV type sub-groups by age, number of doses (3 vs. 2), and other factors within the group, will be tested for statistical significance ($p < 0.05$) using a Fisher's exact or Pearson chi-square test.

Notably, study groups of participants are fundamentally different populations (vs. a randomized control trial that would recruit from the same population and, as result of randomization, enroll similar, representative populations in each arm). Specifically, there are known differences in age and therefore possible differences in sexual activity between Gardasil®-vaccinated (Group 1) vs. -unvaccinated WLWH (Group 2), and possible differences in sexual behaviors between Gardasil®-vaccinated WLWH (Group 1) vs. HIV[-] women (Group 3)(since HIV infection predominately is sexually transmitted in this population). Therefore, we will use a relative measure of effectiveness to account for the differences in age and possible differences in

exposure to HPV. We will use logistic regression to calculate the odds ratio (OR) of the point prevalence and the prevalence of 6-12 month persisting HPV infections of Gardasil®-targeted HPV types, individually (HPV6, 11, 16, *or* 18) and in aggregate (HPV6, 11, 16, *and* 18), vs. untargeted HPV genotypes for which there is no evidence of cross-protection, for each anatomic site individually (cervix, anus, OR oral cavity) and combined (cervix, anus, AND oral cavity). That is, we also will compare the ratio between study groups of participants as follows:

$$\frac{N_{\text{HPV6/11/16/18}}}{N_{\text{HPV35/39/51/52/53/56/58/59/66/68}}} [\text{vaccinated WLWH}] (\text{Group 1})$$
$$\frac{N_{\text{HPV6/11/16/18}}}{N_{\text{HPV35/39/51/52/53/56/58/59/66/68}}} [\text{unvaccinated WLWH or HIV[-] women}] (\text{Group 2 or Group 3})$$

or

$$\frac{N_{\text{HPV6/11/16/18}}}{N_{\text{HPV35/39/51/52/53/56/58/59/66/68}}} [\text{vaccinated WLWH}] (\text{Group 1})$$
$$\frac{N_{\text{HPV6/11/16/18}}}{N_{\text{HPV35/39/51/52/53/56/58/59/66/68}}} [\text{vaccinated HIV[-] women}] (\text{Group 3})$$

or

$$\frac{N_{\text{HPV6/11/16/18}}}{N_{\text{HPV35/39/51/52/53/56/58/59/66/68}}} [\text{vaccinated HIV[-] women}] (\text{Group 3})$$
$$\frac{N_{\text{HPV6/11/16/18}}}{N_{\text{HPV35/39/51/52/53/56/58/59/66/68}}} [\text{unvaccinated HIV[-] women}] (\text{Group 4})$$

This will help us to account/adjust for differences, above what can be achieved statistically, in HPV exposure, prevalence, and persistence due to differences in age (n.b., prevalence equals incidence times duration; prevalent infections tend to be more persistent with increasing age[79]) and sexual behaviors between groups. Additional logistic regression models may be used to adjust specifically on other factors, including age, number of doses, and sexual behaviors, to account for population differences between study groups.

Differences in GMT and seropositivity will be tested for statistical significance using the Mann-Whitney and Fisher’s exact tests, respectively. ANOVA and logistic regression models will be used to adjust for/assess the association of other factors (e.g., age at vaccination, age at

enrollment into the study, number of doses, current and past HIV status, and detection of HPV genotypes, etc.) with GMT and seropositivity, respectively.

Sample Size/Power. We made the following assumptions: 1) at least 30% prevalence of HPV infection (of any anatomic site) at baseline, 25% of which will be by Gardasil®-target HPV types and 55% will be untargeted HPV types, 2) a 10% loss to follow-up (LTFU) in 6-12 months, and 3) at least 70% of HPV infections persist for 6-12 months. We justify an HPV prevalence of at least 30% among WLWH less than 30 years of age in this study, based on 30% prevalence of high-risk HPV infection of the cervix alone in 30-34 year-olds WLWH from our previous cervical cancer screening study in Rwanda [80]. We expect that the HPV prevalence may be higher since the prevalence of HPV, like that of other sexually transmitted infections, tends to peak about 5-10 years after the median age of sexual initiation in a population [81, 82], which in Rwanda is around 17 years of age [83, 84]. However, we conservatively used 30% prevalence to ensure adequate statistical power. We justify our assumption of a maximum of 10% loss to follow-up over 6-12 months, given our experience working in Rwanda over the last ~20 years. We justify 70% of prevalent HPV infections persisting for 6-12 months, given that 36-50% of prevalent HPV infections persist 6-months in HIV[-] women [85, 86] and WLWH have an impaired immunity to HPV compared to HIV[-] women.[25, 26]

Sample size calculation relates to the relative effectiveness measure as discussed. Under above assumptions and accounting for 10% misclassification of HPV vaccine status, a sample size of 757 vaccinated and 757 unvaccinated WLWH will provide $\geq 80\%$ power ($p=0.05$) to detect an OR of ≤ 0.5 for persistent HPV6/11/16/18 infections relative to HPV35/39/51/52/53/56/58/59/66/68 (but not HPV31/33/45 because of possible cross protection), in Gardasil®-vaccinated (Group 1) vs. -unvaccinated WLWH (Group 3). Consequently, because

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there will be more participants with a prevalent infection than persistent HPV infection (because there are no losses to follow-up or HPV viral clearance for which to account), there will be $\geq 80\%$ power ($p=0.05$) to detect a OR of ≤ 0.75 in point prevalence of HPV6/11/16/18 infection relative to HPV35/39/51/52/53/56/58/59/66/68, in Gardasil®-vaccinated (Group 1) vs. -unvaccinated WLWH (Group 3).

Using the variance of GMT for HPV antibodies from Einstein *et al.* [87, 88], a sample size of 274 will provide 80% power ($p=0.05$) to detect a 30% difference in GMT between HPV-vaccinated WLWH (Group 1) and HIV[-] (Group 2) women for the same number of doses. If there is no appreciable difference in the GMT between those who got three or two doses, we can combine those groups with different doses. In that case, with a sample size of 548, there will be 80% power ($p=0.05$) to detect a 22% difference between HPV-vaccinated WLWH (Group 1) and HIV[-] women (Group 2).

Patient and Public Involvement

Patients or the public WERE NOT involved in the design, or conduct, or reporting, or dissemination plans of our research. The results of the study will be shared with interested parties including the Rwanda Ministry of Health and we will establish a Community Advisory Board, both of which will help disseminate our findings to the general Rwandan public.

DISCUSSION

Limitations

Several limitations in the proposed study are worth noting. First, our study is not a randomized control trial, but an observational study of populations selected from different groups of women.

We have proposed several approaches to account/adjust for those age differences but as with any observational study, these techniques may be unable to completely control for biases.

Populations recruited into this study are primarily from Kigali and therefore are not representative of all Rwanda, nor are they representative of other populations in SSA or elsewhere in the world. In 2018-9, almost 80% of Rwandan PLWH had a suppressed HIV viral load[89], a higher percentage than for PLWH populations living in many SSA and other countries [90]. Therefore, these results may not be generalizable to all WLWH populations, especially populations of WLWH who are severely immunocompromised.

We are using 6-12-month HPV type-specific persistence as a proxy for cancer risk but ideally, we want to measure the impact of Gardasil® on high-grade cervical and anal abnormalities as a more proximal surrogate for cancer risk. The population will be still too young to have large numbers of these endpoints although we will evaluate these endpoints in secondary analyses. This may warrant a follow-on, follow-up study to evaluate the longer-term effectiveness and immunogenicity in the cohort, especially in the WLWH.

Strengths

Despite noted limitations, there are some important strengths of the study to highlight. First, the early introduction of HPV vaccination that is documented through a national registry, excellent HIV care including a national database and patient tracking, and relatively high prevalence of HIV provides a unique opportunity to study the impact of HPV vaccination in WLWH, the population at the highest risk of cervical cancer [25-27] that therefore would gain the greatest benefit from it, living in Rwanda. Although a comparative effectiveness randomized control trial of different HPV vaccines is probably still warranted to determine how best to protect this most

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3 434 vulnerable population [31], these data may provide some of the first evidence of long-term
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5 435 effectiveness of HPV vaccination in WLWH and inform such a trial. A second strength is that
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7 436 the study is being conducted in Rwanda, a high cervical-cancer burden country [45]. The
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10 437 findings from this study, if positive, may encourage other high-burden countries to accelerate the
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12 438 introduction of HPV vaccination or at least target WLWH and PLWH populations. Finally, HPV
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14 439 DNA and serology testing are being done locally for this study could potentially be transferred
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16 440 and replicated in other LMICs that want to monitor and evaluate HPV-vaccine impact in their
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18 441 populations.
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22 442 Importantly, as a result of implementing this research protocol, we will continue to expand the
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24 443 local capacity to conduct state-of-the-art HPV and molecular epidemiologic research by 1)
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26 444 establishing next-generation sequencing (NGS) technology to perform cervicovaginal, anal, and
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28 445 oral microbiota characterization for this study that will enable locally conducted studies of the
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30 446 human genome and genomic testing for personalized medicine in Rwanda; 2) building upon
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32 447 current ELISA capabilities at the study lab to perform titration of plasma antibodies, a skill that
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34 448 can be applied to other studies of vaccine response; 3) enhancing data capture and management
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36 449 through increased capacity to use REDCap [91-93]; and 4) migrating current HPV vaccine
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38 450 records for participants living in Kigali into a common electronic database, which will allow us
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40 451 to conduct studies more easily on the impact of HPV vaccination on outcomes. Importantly, this
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42 452 will allow linkage of HPV-vaccination status to both the Rwanda National HIV Registry and the
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44 453 Rwanda Cancer Registry, the latter of which we, in collaboration with the Rwanda MoH, helped
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46 454 re-establish [94]. This will allow investigations of the long-term impact of HPV vaccination on
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48 455 cancer incidence in Rwandan WLWH and HIV[-] women and, as a result of herd protection,
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50 456 Rwandan men.
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3 457 As building research capacity in Rwanda is a major goal of this project, all members of the
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5 458 research team will be asked and supported to lead at least one analysis and one manuscript
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7 459 preparation, based on interests and expertise. Analytic and publication responsibilities will be
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9 460 divided equally and collaboratively among both Rwandan and U.S. investigators.
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ETHICS AND DISSEMINATION

The study was approved by the Albert Einstein College of Medicine Institutional Review Board (IRB#: 2021-13087) and the Rwanda National Ethics Committee.

All participant data collected will be entered or transferred to a secure REDCap database, with access to personal information restricted to the local staff. Study data will be stored in REDCap and maintained on password protected study computers behind the institution firewall.

Copies of signed consents will be stored in locked file cabinets in a locked room, with access restricted to study personnel only, at the study lab. Data will be entered or transferred to a secure REDCap database, with access to personal information restricted to the local staff.

Study data will be stored in REDCap and maintained on password protected study computers behind the institution firewall. REDCap is a secure, web-based application designed to support data capture for research studies providing: 1), an intuitive interface for validated data entry; 2) audit trails for tracking data manipulation and export procedures; 3) automated export procedures for seamless data downloads to common statistical packages; and 4) procedures for importing data from external sources. REDCap is hosted on a secure server and has undergone a Governance Risk & Compliance Assessment by All REDCap electronic data files shared with Albert Einstein College of Medicine will be maintained by the HIPAA-compliant Epidemiology Study Management and Informatics Core Facility (ESMI) at Einstein. The Albert Einstein College of Medicine policy on use of REDCap can be found at:

http://ric.einstein.yu.edu/ric_files/REDCap%20Appropriate%20Use%20Policy.pdf

Medical, screening, and preventive services, all of which are minimally invasive, safe, and outpatient, and have been done in millions of people, provided by the study are on par with or

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3 483 better than international standards-of-care, most of which have very low risks of even minor
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5 484 adverse events or harms. Positive test results and diagnoses may result in psychologic distress
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8 485 and anxiety. Of note, pregnancy and HIV testing may cause pre- and post-test anxiety. Therefore,
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10 486 pre- and post-testing counseling will be provided as needed. Any participant who tests HIV
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12 487 positive will be referred to the health facility's HIV clinic for HIV management following the
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14 488 Rwanda MoH's HIV management guidelines.

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18 489 We will publish a series of reports in peer-review scientific journals to disseminate these results.

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20 490 We will share the results with the Rwanda MoH. Data from this study will be made available by
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22 491 request in accordance with NIH policies and Rwandan laws.

23 24 25 26 492 **ETHICS STATEMENT**

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29 493 Patient consent for publication not required.

30 31 32 494 **ACKNOWLEDGEMENTS**

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35 495 We acknowledge and thank the Rwandan women who participate in this study. We also thank
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37 496 the research teams at Albert Einstein College of Medicine, the Einstein-Rwanda Research &
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39 497 Capacity Building Program, and the U.S. National Cancer Institute for their contributions in
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41 498 developing and implementing this research protocol.
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- **Contributorship statement:** GM, PEC, and KA designed the study initially. GM, AP, PEC, and KA refined and finalized the overall study design and wrote the grant proposal that is the basis for this protocol. GM, FS, NH, AM, JCG, AP, KA, and PEC drafted and finalized the written protocol. GM, FS, NH, AM, JCG, BM, FK, AP, PT, KA, and PEC implemented it. PEC drafted this manuscript. All authors have read, revised, and approved the final manuscript. GM and FS are co-first authors; KA and PEC are co-senior authors.
- **Competing interests statement:** Dr. Castle is the Director of the Division of Cancer Prevention at the NCI, the NIH institute that funds this research. However, Dr. Castle has recused himself from any decisional or financial authority over this grant or any extramural HPV grants funded by the NCI. Other authors claim no competing interests. The study is receiving HPV genotyping tests at a reduce cost from Atila Biosystems.
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- **Data sharing statement:** Data will be shared in accordance to policies for NIH-funded research at the conclusion of the study and after the publication of the main scientific findings.

Figures

Figure 1. Enrollment visit procedures

Figure 2. Follow-up visit procedures

Figure 3. Follow-up and clinical management of baseline HPV-positive results

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Figure 1. Enrollment Visit

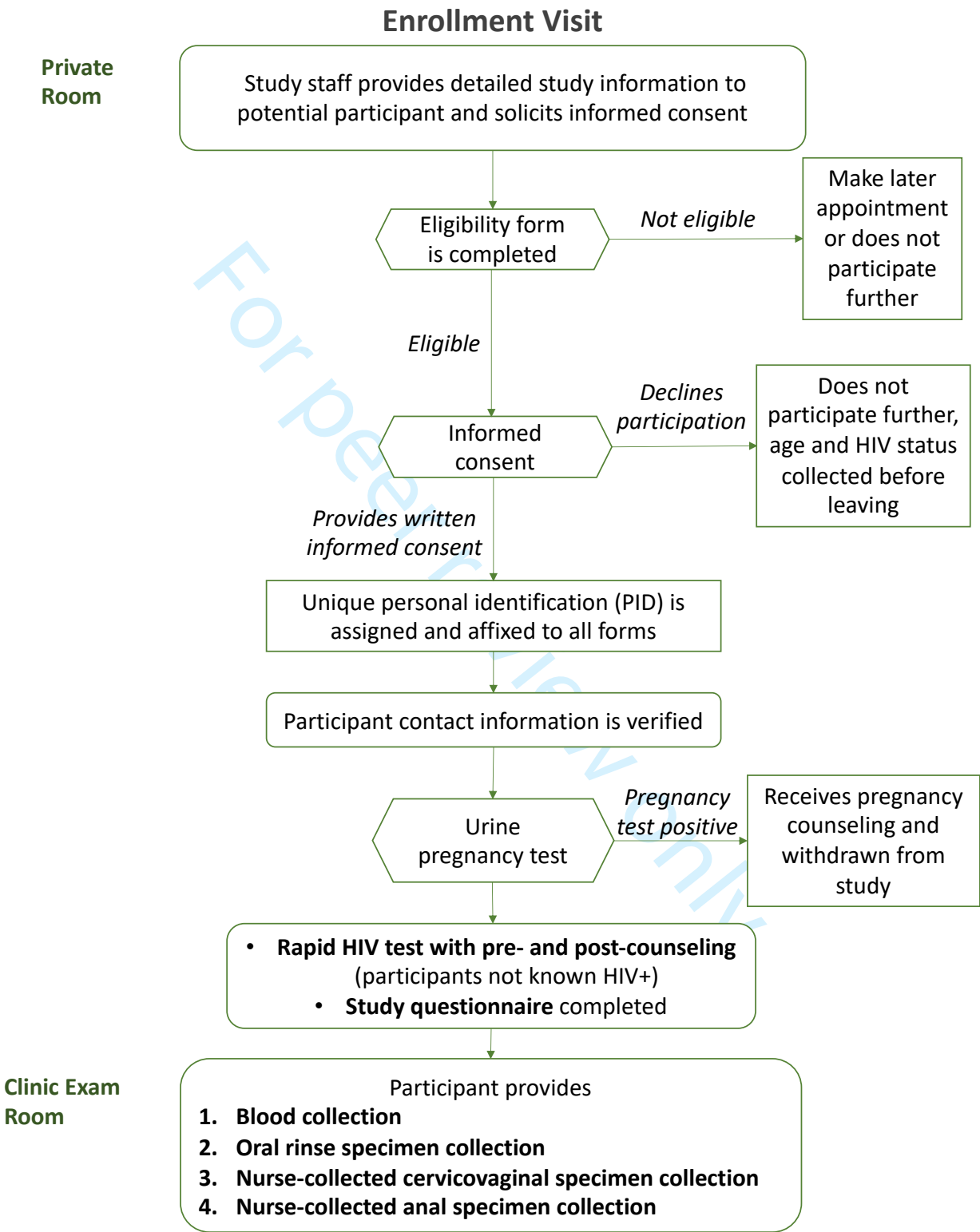


Figure 2. Follow-up Visit

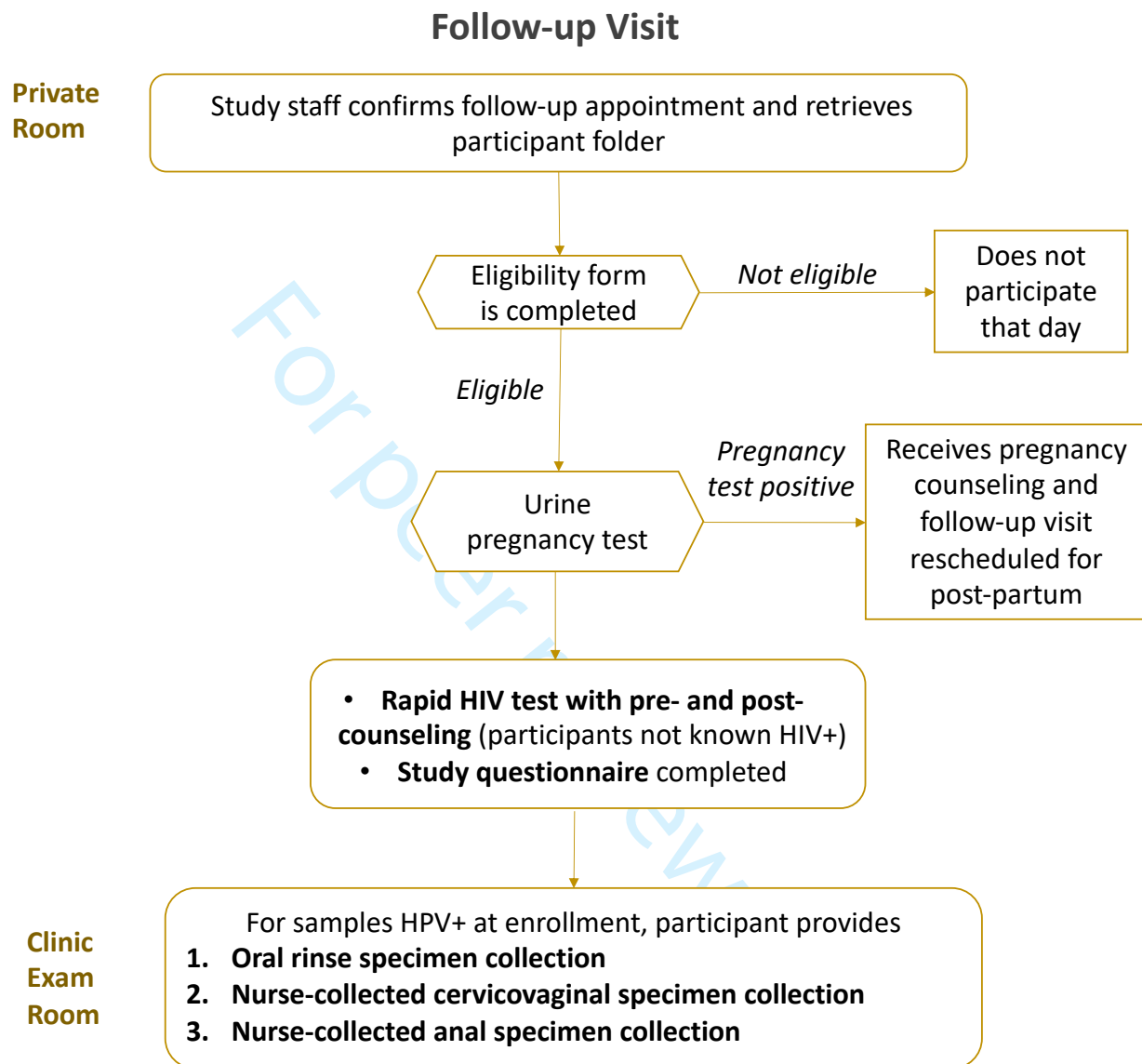


Figure 3. Follow-up and clinical management of baseline HPV-positive results

